

1 **SPECIES AUTHENTICATION IN POULTRY MEAT PRODUCTS BY NEXT-**  
2 **GENERATION SEQUENCING**

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19 product mislabelling

20

21 **Abstract**

22 The growing demand for poultry meat and the complexity of the supply chain affect traceability.  
23 Therefore, the aim of this study was to apply DNA metabarcoding to verify the labelling compliance  
24 of multi-species poultry meat commercial products .  
25 Overall, the molecular identifications conducted in this study confirm that all products contain the  
26 species declared in the label and all the non-conformities regard the addition of one or more  
27 undeclared meat species which could reflect both unintentional and fraudulent behaviors. In  
28 particular, the presence of undeclared species, such as swine and bovine, were highlighted in 8/13  
29 (60%) of the samples. Such composition? pattern could be due to technological purposes or accidental  
30 contamination linked to inappropriate sanitation practices during processing. However, the presence  
31 of undeclared species can affect the ability to choose for consumers with of specific needs (e.g., ethic,  
32 religious) or health risk (e.g., cardiovascular diseases, obesity) and should be not neglected. Results  
33 of this study show that metabarcoding is a promising tool to identify meat species in mixtures.  
34 Therefore, its application by competent companies and institutions, could help to innovate the food  
35 management system with the creation of a favorable environment for the protection and respect of  
36 the consumer needs.

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## 39 INTRODUCTION

40 Meat consumption has been shifting towards poultry, driven by two different forces. In lower income  
41 developing countries, poultry has lower price compared to other meats, while in high-income  
42 countries poultry meats are considered more convenient to prepare and perceived as a healthier food  
43 choice with a risk reduction of cardiovascular diseases (Falkovskaya & Gowen, 2020; Marangoni et  
44 al., 2015; OECD-FAO Agricultural Outlook 2021-2030). Poultry meat is the meat of domesticated  
45 birds, such as turkey and chicken, and poultry products foods such as sausages, patties, hamburgers  
46 are gaining growing interest among consumers due to their convenience in preparation (ready to  
47 cook), handling, and storage (Barbut, 2012; Kennedy et al., 2004). However, in these kinds of  
48 products, the ingredients are naturally less traceable due to international trade, market globalization,  
49 and long and complex food supply chains. In addition, with the booming of e-commerce, the  
50 opportunities for their fraud increased and mixed meat products are often considered among the most  
51 frequently adulterated foods (Di Pinto et al., 2015; Hassoun et al., 2020; Walker et al., 2013). In  
52 general, food frauds involving partial or full species substitutions are expected to increase economic  
53 gain with high-priced species being substituted by cheaper ones or even illegally trade matrices  
54 (Barbarossa et al., 2016). On the other hands, accidental species substitution could occurs and may  
55 be associated with unintentional cross-contamination in processing plants sharing common  
56 machinery or equipment to produce different meat products or improper human handling (Keyvan et  
57 al., 2017). Whether intentional or not, the incorrect description of meat products is an issue of primary  
58 importance not only for economic value, but also for the potential public health risks. Indeed species  
59 substitution could have a direct impact on health of consumers when meat not compliance with  
60 hygiene requirements or even coming from illicit trade is used(Vidal Junior et al., 2020). Moreover,  
61 species substitution affects the possibility for the consumers to choose products based on ethical  
62 issues as sustainable production, animal health and wellness, health problems or religious laws  
63 (Bertolini et al., 2015). The implementation of control systems is fundamental, and several methods,

64 including molecular, chromatography, spectroscopy, and/or spectrometry, as well as imaging  
65 approaches, have been used for the authentication of meat products (Ballin, 2010; Ellis et al., 2015;  
66 Fengou et al., 2021; Ropodi et al., 2017). However, considering the high stability and highly  
67 specificity of DNA present in almost all tissue types, molecular approaches are considered the most  
68 appropriate allowing the differentiation even in cases of closely related species. Many DNA-based  
69 methods such as DNA sequencing, species specific PCR, randomly amplified polymorphic DNA  
70 (RAPD), restriction-fragment-length polymorphism (RFLP), real-time PCR, Droplet digital PCR  
71 (ddPCR) loop-mediated isothermal amplification (LAMP), and touchdown PCR (TD-PCR) have  
72 effectively and largely been developed, tested and used for identification and differentiation of animal  
73 species in meat products both in single ingredient commodities and in complex matrices (Cai et al.,  
74 2017; Di Pinto et al., 2015, 2019; Haider et al., 2012; Kumar et al., 2017; Lin et al., 2019; Nischala  
75 et al., 2022). However, these analytical techniques require knowledge about which species to search  
76 for, and therefore are not appropriate for detecting all the species used in mixed meat products.  
77 Currently, DNA metabarcoding, the combination of DNA barcoding with Next Generation  
78 Sequencing platforms (NGS), could play an important role in food authentication without the  
79 requirement for previous knowledge of the supply chain, production process, ingredients or about the  
80 species to search for. Thank to these new generation of sequencers, all the DNA molecules extracted  
81 from the matrices can be simultaneously amplified and sequenced allowing species identification also  
82 in complex foods containing multiple ingredients. Although application of metabarcoding to trace  
83 ingredients is still in its infancy, several studies have tested this approach for species identification in  
84 different food products including dairy (Ribani et al., 2018), seafood (Giusti et al., 2017; Piredda et  
85 al., 2022), commercial plant (Bruno et al., 2019), herbal medicinal (Anthoons et al., 2021), candies  
86 (Muñoz-Colmenero et al., 2017) honey (Prosser & Hebert, 2017; Wirta et al., 2021), probiotics (Patro  
87 et al., 2016) and pet food (Palumbo et al., 2020; Preckel et al., 2021). Few studies applied  
88 metabarcoding to investigate the species composition in artificially prepared mixtures or commercial

98 meat and meat products (i.e., sausages, balls, canned luncheon meat, minced meats, kebab) made of  
 99 several animal species including beef, camel, horse, sheep, deer, swine and/or poultry (Cottenet et  
 100 al., 2020; Dobrovolny et al., 2019; Pan et al., 2020; Preckel et al., 2021; Xing et al., 2019). Despite  
 101 the paucity, these studies have shown that metabarcoding is a promising tool to species authentication  
 102 also able to reveal the presence of unexpected taxa in addition to those declared.

103 In this study, the DNA metabarcoding approach will be applied to multi-species poultry meat products  
 104 to verify the declared list of ingredient in term of animal species Results of this work, by highlighting  
 105 the potential application of DNA metabarcoding for food authentication and traceability, could  
 106 innovate the food management system throughout the supply chain.

98

## 99 **2. MATERIALS AND METHODS**

### 100 **2.1 Sampling**

101 A total of 10 packaged and 3 unpackaged poultry meat products samples including sausages, cutlets  
 102 hamburgers and meat patties, reporting in the ingredient list as manufactured using pure chicken (6),  
 103 pure turkey (2), chicken and turkey meat (4) and chicken, turkey, and swine meat (1) (Table1)

<b>Sample ID</b>	<b>Type of product</b>	<b>Type of production</b>	<b>Packaged or unpackaged</b>	<b>Market/supermarket????</b>	<b>Declared species</b>
<b>Sample 01</b>	Sausage	Artisanal			Chicken
<b>Sample 02</b>	Hamburger	Artisanal			Chicken
<b>Sample 03</b>	Patties	Industrial			Chicken, milk traces
<b>Sample 04</b>	Hamburger	Industrial			Chicken
<b>Sample 05</b>	Hamburger	Industrial			Chicken
<b>Sample 06</b>	Hamburger	Industrial			Chicken, milk traces
<b>Sample 07</b>	Hamburger	Industrial			Chicken, Turkey, Bovine proteins
<b>Sample 08</b>	Sausage	Artisanal			Chicken, Turkey

Sample 09	Cutlet	Industrial			Chicken, Turkey, milk traces
Sample 10	Hamburger	Industrial			Chicken, Turkey
Sample 11	Sausage	Industrial			Turkey, Chicken, Swine
Sample 12	Hamburger	Industrial			Turkey
Sample 13	Hamburger	Industrial			Turkey

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105 were purchased from different markets and supermarkets in the Apulia region (SE, Italy). Samples  
106 were stored at -20 °C until processed. Positive control was generated using an artificial DNA pool  
107 constructed from 50 ng of VERYfinder Poultry Pure DNA Extract – HEAT TREATED MEAT  
108 (Generon, Italy), 50 ng of VERYfinder Turkey Pure DNA Extract – HEAT TREATED MEAT  
109 (Generon, Italy), 50 ng of VERYfinder Bovine Pure DNA Extract – HEAT TREATED MEAT  
110 (Generon, Italy), and 50 ng of VERYfinder Swine Pure DNA Extract – HEAT TREATED MEAT  
111 (Generon, Italy).

## 112 2.2. DNA extraction, purification, and sequencing

113 Total genomic DNA was extracted from all samples starting from aliquots of 25 mg of each sample  
114 using the DNeasy Blood and Tissue Kit (QIAGEN, Hilden, Germany) following producer  
115 instructions. To verify the purity of the extraction reagents, blank negative control (no added tissue)  
116 was included. DNA concentration and purity were established by evaluating the ratio A260 nm/A280  
117 nm using a BioPhotometer D30 filter (Eppendorf, Milan, Italy). Then, the DNA was amplified using  
118 the primer pairs previously tested by Pan et al. (2020), consisting in mini-COI-F: 5'-  
119 GGTCACAATCATAAAGATATTGG-3' (Folmer et al., 1994) and mini-COI-R:5'-  
120 ACTATAAAGAAGATTATTACAAAGGC-3' (Pan et al., 2020), amplifying a fragment of about  
121 136bp of the cytochrome oxidase I (COI) mitochondrial gene. The sequencing was carried out on  
122 the Illumina NextSeq platform by LGC Genomics GmbH (Berlin, Germany) (2×150 bp). PCR  
123 negative controls (no template) were included during the amplification step of library preparation.

124 Raw sequences were deposited in the Sequence Read Archive (SRA) under the BioProject (under  
125 submission).

126 Illumina paired-end raw reads were pre-processed to generate Amplicon Sequence Variants (ASVs)  
127 using DADA2 R package (Callahan et al., 2016). Briefly, primers were removed, forward and reverse  
128 trimmed based on the Quality score and, the reads filtered, were then used to train the error model  
129 using machine learning approach. Then forward and reverse were dereplicated to generate unique  
130 sequences and denoised (collapsed) in amplicon sequence variants (ASVs) applying the trained error  
131 model. Finally forward and reverse reads were merged and checked for chimera sequences.  
132 Representative sequence for each ASV were taxonomic assigned blasting the representative  
133 sequences against GenBank in remote mode using the standalone blast + suite (Altschul et al., 1990;  
134 Camacho et al., 2009) and assignments with a similarity of <90%, representing potential low-quality  
135 reads, were discarded. Sequences assigned in the range 100-98% of similarity were assigned at  
136 species level (Barbuto et al., 2010) and merged. Molecular results were then compared with the list  
137 on ingredients reported on the labels.

### 138 **3. RESULTS**

139 The Illumina sequencing of the 13 meat samples generated a total of 5,695,822 raw reads and filtering  
140 reduced the dataset to 5,473,877 reads. Positive control sample confirmed the efficiency of the  
141 primers and generated 396,128 raw reads reduced to 376,484 (Suppl. Table 1). Taxonomic  
142 assignments revealed the presences of four species *Bos taurus*, *Gallus gallus*, *Meleagris gallopavo*  
143 and *Sus scrofa*.

144 The comparison between the results of the molecular identification and ingredient lists confirmed that  
145 all the samples contained the species reported on the label. In eight cases 8/13 (61,5%) the presence  
146 of unexpected species was highlighted. All the six samples labeled as pure chicken were found to  
147 contain additional species: turkey in 4/6 cases (67%), turkey, swine and bovine in 2/6 (33%). As  
148 regards the 2 samples of turkey hamburger, the analysis confirmed the presence of turkey, but in one

149 case (50%) also the presence of chicken. Within the 4 chicken and turkey products, one sample (25%)  
150 revealed also the presence of bovine. Finally, in the chicken, turkey and swine sausages molecular  
151 data confirmed the species reported on the label (Figure 1 and Table 1). In the sample Sample 04  
152 (chicken hamburger) the presence of the avian feather mites *Proctophyllodes sylviae* (Acari:  
153 Astigmata), an important symbiont of birds, was even detected.

#### 154 **4. DISCUSSION**

155 The globalization of meat supply and the consequent increase in the complexity of supply chains has  
156 significantly increased the risks of food fraud. The European Union (EU) is one of the world's largest  
157 poultry meat producers. However, EU imports high value poultry products, including breast meat and  
158 poultry preparations, mainly from Brazil, Thailand, and Ukraine, while the EU exports poultry  
159 products of lower value (European Union, 2022). Such complexity reduces the ability of both  
160 regulators and industry to effectively oversee food supply chains, creating further confusion and  
161 weakness that can facilitate inadequate practices. Furthermore, considering the reduced worldwide  
162 regulatory monitoring that appears to have occurred during the pandemic, the COVID-19 pandemic  
163 played a role in the observed increase in food fraud incidents from January to June 2020 compared to  
164 the same period in 2019 (Brooks et al., 2021).

165 Overall, the molecular identifications conducted in this study, focusing on poultry meat products,  
166 confirm that all products contain the species declared in the label and all the non-conformities regard  
167 the addition of one or more undeclared meat species which could reflect both unintentional and  
168 fraudulent behaviors. The primers pair used in this study were proved to be able to identify 51 edible  
169 animal species (swine, bovine, poultry, ovine, caprine, and some fish and shrimp), also including  
170 *Homo sapiens* (Pan et al., 2020); however, only four edible species (chicken, turkey, swine, and  
171 bovine) have been detected in the analyzed products. In particular, in this study, we revealed a high  
172 rate of non-conformity (61.5%) as found by Xing et al., 2019 who reported a non-conformity rate of  
173 59% in processed meat and poultry products. Despite the reduced number of samples, this study



174 represents one of the few studies available for poultry products and it corroborates the hypothesis that  
175 processed foods with no morphological structure are vulnerable to species substitution, either  
176 intentionally or unintentionally, practiced at any stage of the supply chain (Lianou et al., 2021).  
177 However, we have to consider that chicken, turkey, swine, and bovine, represent the top consumed  
178 meat types worldwide and are expected to be routinely present in the butchery, so their presence in  
179 the products could be related to an accidental introduction due to the fact that different raw materials  
180 are processed within the same processing plants (Di Pinto et al., 2019; Marchetti et al., 2021). Indeed,  
181 cross-contamination, can easily occur when improperly cleaned equipment is used to process meat of  
182 multiple species. This hypothesis could be the reason why, according to our results, all the samples  
183 purchased from butcher's shop contain four types of meat independently to those declared in label  
184 and suggesting more risks of incident in 'artisanal' products than in industrial ones. On the other  
185 hand, the addition driven by economic benefit cannot be excluded since undeclared meat could be  
186 intentionally and illegally incorporated into the products for technological purposes. In particular, the  
187 presence of bovine or porcine DNA could be due to the fraudulent addition to poultry of water  
188 containing proteins of porcine or bovine origin, aimed at aid carcasses water retention (Fuseini et al.,  
189 2017; Lianou et al., 2021). Yet, in the unpackaged sausage sample, pork casings could have been  
190 used to contain the products. In addition, more mechanically recovered meat (MRM), often produced  
191 from pork and chicken carcass, is added as cheap protein source to meat products such as sausages,  
192 hamburgers, or cured meats (Surowiec et al., 2011). Similarly, the undeclared bovine presence could  
193 have been due to intentional addition of non-fat dry milk powder to increase taste and to improve  
194 binding qualities (Di Pinto et al., 2015, 2019). Whatever was the reason or source, the non-conformity  
195 due to species substitution to the presence of undeclared species has always consequences for the  
196 consumers, and this is especially important in the case of poultry meat products which are often  
197 chosen and purchased on the bases of specific needs. Indeed, poultry products are allowed in presence  
198 of strict religious restrictions on the consumption of pork and bovine, as is the case of Muslim food

199 laws (Halaal) and in Jewish food laws (Kashrut) that forbid swine meats, or Hindus that abstain from  
200 eating beef meat (Ng et al., 2022). For this reason, the simultaneous presence of bovine and swine  
201 (three samples) and bovine (two samples) found in our samples in addition to poultry meats, should  
202 be seriously considered. Indeed, Europe's societies are undergoing change and, even if Italy cannot  
203 be considered as multiethnic or multicultural country right away, we can predict an increase of  
204 consumers with different cultural values for which the undeclared presence of bovine and swine will  
205 have more impact and weight in comparison with the traditional Italian consumers.

206 In addition to ethical aspects, species substitution includes violation of the EU traceability  
207 requirements of the transparent food labelling systems set forth respectively in Reg. EC 178/02 and  
208 Reg. EU 1169/2011, as well as the code of conduct on the management of food allergens established  
209 by the recent Commission Regulation (EU) 2021/382 (Mottola et al., 2022). A full traceability of  
210 ingredients is fundamental for people with allergies to milk or with allergic reactions to gelatin  
211 (Caponetto et al., 2013; Zin et al., 2021). Moreover, although the allergy to meat itself has historically  
212 been considered quite rare, cases of allergy to meat from mammals and birds, beef, pork, lamb, and  
213 poultry have become more common starting around 20 years ago (Marques et al., 2021; Wilson &  
214 Platts-Mills, 2018, 2019).

215 Despite the European Union Commission defines food fraud as “*any suspected intentional action by*  
216 *businesses or individuals for the purpose of deceiving purchasers and gaining undue advantage*  
217 *therefrom, in violation of the rules referred to in Article 1(2) of Regulation (EU) 2017/625”*, to date  
218 from a regulatory point of view, it lacks a specific body of legislation and a clear and shared definition  
219 of "food fraud", as well as details concerning the approaches of discriminating between accidental  
220 and intentional. In the routine analysis of samples in public laboratories, mass concentrations below  
221 1% (w/w) are generally reported as possible process contaminants and do not constitute a violation  
222 of declaration since substitution at such low levels should not have an economic advantage (Al-  
223 Kahtani et al., 2017). However, this requires specific discipline and great inspection attention. Indeed,

224 any possible accidental or low-level presence linked to the unintentional presence and accidental  
225 traces of a type of food product with another species during processing and handling must in any case  
226 be indicated on the label as "May contain traces". On the contrary, to prevent or react on food fraud-  
227 incidents, companies need to plan mitigation practices and prevention strategies for species  
228 replacement practices, making use of the Food Fraud Vulnerability Assessment, an effective measure  
229 for specific risk management for the food industries, food authorities and consumers (Marchetti et  
230 al., 2021). A comprehensive food fraud and adulteration prevention program could be a decisive and  
231 fundamental development factor in the innovation of the processed poultry sector for more accurate  
232 and truthful labeling (Di Pinto et al., 2019). The main tools for controlling vulnerabilities include the  
233 traceability plan vulnerability and severity analyses and assessment of risk, evaluation of the  
234 preventive measures in place, identification of critical points for controlling origin of the fraud,  
235 establishing a system for monitoring and critical limits, corrective actions and verification and  
236 validation of the system. Specifically, the development of standardized tests (Pan et al., 2020) play a  
237 crucial role in meat product authentication and in the mitigation of food fraud related to species  
238 substitution.

239 Our outcomes show that metabarcoding is a promising tool to identify meat species in mixtures which  
240 often contain multiple animals including species not routinely used, which were not suspected to be  
241 present or for which real-time PCR methods are not available (Piredda et al., 2022). In addition, the  
242 high sensitivity of metabarcoding approach could also help the estimation of hygienic conditions of  
243 meat supply chain. Indeed, Pan et al. (2020) detected the presence of fly and cockroach in their  
244 artificial meat mixture samples, probably due to a contamination of laboratory working environment,  
245 showing that the application of metabarcoding on food products could trace not only the mislabeling  
246 but also the history of environmental and hygienic-sanitary conditions. In this sense, very positive  
247 outcomes emerge from our poultry samples in which none 'unexpected' eukaryotic taxa are detected  
248 except for one sample in which we detected trace of avian feather mites belonging to

249 *Proctophyllodidae*. However, such presence cannot be related to a scarce hygienic condition since  
250 the general avian slaughter process operations does not remove avian skin, and thus could justify the  
251 *Proctophyllodidae* presence. Furthermore, it does not represent a sanitary risk given that, to date, this  
252 avian mite has not been identified as harmful for human health. Similarly, the presence of  
253 *Proctophyllodidae* seems to be not an animal welfare issue given that they are bird ectosymbionts  
254 that play an important biological function in cleaning bird feathers (Dona et al., 2019). Interestingly,  
255 also traces of human DNA are absent in our samples suggesting that the strong obligation of masks  
256 and gloves probably due to pandemic times, could have avoid the occasional contamination with the  
257 operator's saliva from talking while work with a consequent improvement of safety.

258 Despite the great potential of metabarcoding approach, the semiquantitative nature of metabarcoding  
259 approach is well known and the limitations for DNA quantification have been reported in several  
260 field including in meat products (Cottenet et al., 2020; Preckel et al., 2021). Different reasons  
261 contribute to this bias as tissue type, genome size, copy number for nuclear regions, number of  
262 mitochondrial in cells/tissues/organs (Ren et al., 2017). Moreover, since metabarcoding is a PCR-  
263 based method, variations in primer binding capacity, could overestimate low abundant taxa with  
264 higher primers affinity or underestimate high abundant taxa with lower primer affinity, especially in  
265 complex food matrices with competitive affinity of animal species. On the other hand, in most of the  
266 cases, the labels of sampled products didn't include the proportion of animals used in the mixtures  
267 so, a qualitative approach for the comparison remains the most practicable way for the metabarcoding  
268 assessments in meat products.

269

## 270 **5 CONCLUSIONS**

271 Poultry meat product species substitution result from the combination of opportunities, motivations,  
272 and inadequate control measures. The poultry meat is the fastest growing segment in the world meat  
273 market (Roiter et al., 2021) and it is necessary to conduct baseline studies of the current state, identify

274 strengths and weaknesses of the supply chain specific, build vulnerability assessment and critical  
275 control point system. The development and application of strategies and tools for traceability are  
276 required especially for processed poultry meat products because these products are often the target of  
277 consumers with of specific needs (e.g., ethic, religious) or health risk (e.g., cardiovascular diseases,  
278 obesity, diabetes) that should be respected. The innovative approach of DNA metabarcoding could  
279 be a suitable method helping the authentication of animal species in mixed meat products. Its  
280 application in routine assays may verifying compliance with food labeling for the protection of  
281 consumers and contribute to the achievement of the European Green Deal objectives for the food  
282 systems.

283



284

285 **Figure 1. Comparison between species reported in label and metabarcoding identification.** Number and  
 286 type of species reported in Label (yellow on the left) and Metabarcoding identification (blue on the right)  
 287 with unexpected species highlighted in bold. Mislabeled samples are indicated with asterisk (\*)

288

289 **Table 1.** Details of information of poultry products and molecular identification by metabarcoding

<b>Sample ID</b>	<b>Type of product</b>	<b>Type of production</b>	<b>Declared species</b>	<b>Molecular Identification</b>
Sample 01	Sausage	Artisanal	Chicken	Chicken, Turkey, Bovine, Swine
Sample 02	Hamburger	Artisanal	Chicken	Chicken, Turkey, Bovine, Swine
Sample 03	Patties	Industrial	Chicken, milk traces	Chicken, Turkey
Sample 04	Hamburger	Industrial	Chicken	Chicken, Turkey
Sample 05	Hamburger	Industrial	Chicken	Chicken, Turkey
Sample 06	Hamburger	Industrial	Chicken, milk traces	Chicken, Turkey
Sample 07	Hamburger	Industrial	Chicken, Turkey, Bovine proteins	Chicken, Turkey, Bovine
Sample 08	Sausage	Artisanal	Chicken, Turkey	Chicken, Turkey, Bovine, Swine
Sample 09	Cutlet	Industrial	Chicken, Turkey, milk traces	Chicken, Turkey, Bovine
Sample 10	Hamburger	Industrial	Chicken, Turkey	Chicken, Turkey
Sample 11	Sausage	Industrial	Turkey, Chicken, Swine	Chicken, Turkey, Swine
Sample 12	Hamburger	Industrial	Turkey	Turkey
Sample 13	Hamburger	Industrial	Turkey	Chicken, Turkey

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291 **References**

- 292 Akhbarizadeh, R., Dobaradaran, S., Nabipour, I., Tajbakhsh, S., Darabi, A. H., & Spitz, J. (2020).  
293 Abundance, composition, and potential intake of microplastics in canned fish. *Marine*  
294 *Pollution Bulletin*, 160, 111633. <https://doi.org/10.1016/j.marpolbul.2020.111633>
- 295 Al-Kahtani, H. A., Ismail, E. A., & Ahmed, M. A. (2017). Pork detection in binary meat mixtures  
296 and some commercial food products using conventional and real-time PCR techniques.  
297 *Food chemistry*, 219, 54–60.
- 298 Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignments  
299 earch tool. *Journal of Molecular Biology*, 215, 403–410. [https://doi.org/10.1016/S0022 -](https://doi.org/10.1016/S0022-)  
300 [2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
- 301 Anthoons, B., Karamichali, I., Schrøder-Nielsen, A., Drouzas, A. D., de Boer, H., & Madesis, P.  
302 (2021). Metabarcoding reveals low fidelity and presence of toxic species in short chain-of-  
303 commercialization of herbal products. *Journal of Food Composition and Analysis*, 97,  
304 103767.
- 305 Ballin, N. Z. (2010). Authentication of meat and meat products. *Meat science*, 86(3), 577–587.
- 306 Barbarossa, C., De Pelsmacker, P., Moons, I., & Marcati, A. (2016). The influence of country-of-  
307 origin stereotypes on consumer responses to food safety scandals: The case of the horsemeat  
308 adulteration. *Food Quality and Preference*, 53, 71–83.
- 309 Barbut, S. (2012). Convenience breaded poultry meat products—New developments. *Trends in Food*  
310 *Science & Technology*, 26(1), 14–20.
- 311 Barbuto, M., Galimberti, A., Ferri, E., Labra, M., Malandra, R., Galli, P., & Casiraghi, M. (2010).  
312 DNA barcoding reveals fraudulent substitutions in shark seafood products: The Italian case  
313 of “palombo” (*Mustelus* spp.). *Food research international*, 43(1), 376–381.



314 Bertolini, F., Ghionda, M. C., D'Alessandro, E., Geraci, C., Chiofalo, V., & Fontanesi, L. (2015). A  
315 next generation semiconductor based sequencing approach for the identification of meat  
316 species in DNA mixtures. *PloS one*, *10*(4), e0121701.

317 Brooks, C., Parr, L., Smith, J. M., Buchanan, D., Snioch, D., & Hebishy, E. (2021). A review of  
318 food fraud and food authenticity across the food supply chain, with an examination of the  
319 impact of the COVID-19 pandemic and Brexit on food industry. *Food Control*, *130*,  
320 108171.

321 Bruno, A., Sandionigi, A., Agostinetto, G., Bernabovi, L., Frigerio, J., Casiraghi, M., & Labra, M.  
322 (2019). Food tracking perspective: DNA metabarcoding to identify plant composition in  
323 complex and processed food products. *Genes*, *10*(3), 248.

324 Cai, Y., He, Y., Lv, R., Chen, H., Wang, Q., & Pan, L. (2017). Detection and quantification of beef  
325 and pork materials in meat products by duplex droplet digital PCR. *PLoS One*, *12*(8),  
326 e0181949.

327 Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. A., & Holmes, S. P.  
328 (2016). DADA2: High-resolution sample inference from Illumina amplicon data. *Nature*  
329 *Methods*, *13*(7), 581–583. <https://doi.org/10.1038/nmeth.3869>

330 Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., & Madden, T. L.  
331 (2009). BLAST+: Architecture and applications. *BMC Bioinformatics*, *10*(1), 421.  
332 <https://doi.org/10.1186/1471-2105-10-421>

333 Caponetto, P., Fischer, J., & Biedermann, T. (2013). Gelatin-containing sweets can elicit  
334 anaphylaxis in a patient with sensitization to galactose- $\alpha$ -1, 3-galactose. *The Journal of*  
335 *Allergy and Clinical Immunology: In Practice*, *1*(3), 302–303.

336 Chuah, L.-O., He, X. B., Effarizah, M. E., Syahariza, Z. A., Shamila-Syuhada, A. K., & Rusul, G.  
337 (2016). Mislabelling of beef and poultry products sold in Malaysia. *Food Control*, *62*, 157–  
338 164.

339 Cottenet, G., Blancpain, C., Chuah, P. F., & Cavin, C. (2020). Evaluation and application of a next  
340 generation sequencing approach for meat species identification. *Food Control*, *110*, 107003.

341 Di Pinto, A., Bottaro, M., Bonerba, E., Bozzo, G., Ceci, E., Marchetti, P., Mottola, A., & Tantillo,  
342 G. (2015). Occurrence of mislabeling in meat products using DNA-based assay. *Journal of*  
343 *Food Science and Technology*, *52*(4), 2479–2484. <https://doi.org/10.1007/s13197-014-1552->  
344 [y](https://doi.org/10.1007/s13197-014-1552-y)

345 Di Pinto, A., Mottola, A., Marchetti, P., Savarino, A., & Tantillo, G. (2019). Fraudulent species  
346 substitution in e-commerce of protected denomination origin (pdo) products. *Journal of*  
347 *Food Composition and Analysis*, *79*, 143–147.

348 Dona, J., Proctor, H., Serrano, D., Johnson, K. P., Oploo, A. O., Huguet-Tapia, J. C., Ascunce, M.  
349 S., & Jovani, R. (2019). Feather mites play a role in cleaning host feathers: New insights  
350 from DNA metabarcoding and microscopy. *Molecular ecology*, *28*(2), 203–218.

351 Ellis, D. I., Muhamadali, H., Haughey, S. A., Elliott, C. T., & Goodacre, R. (2015). Point-and-  
352 shoot: Rapid quantitative detection methods for on-site food fraud analysis—moving out of  
353 the laboratory and into the food supply chain. *Analytical Methods*, *7*(22), 9401–9414.

354 European Union (2022) [https://ec.europa.eu/info/food-farming-fisheries/animals-and-animal-](https://ec.europa.eu/info/food-farming-fisheries/animals-and-animal-products/animal-products/poultry_en)  
355 [products/animal-products/poultry\\_en](https://ec.europa.eu/info/food-farming-fisheries/animals-and-animal-products/animal-products/poultry_en) (30th May 2022)

356 Falkovskaya, A., & Gowen, A. (2020). Literature review: Spectral imaging applied to poultry  
357 products. *Poultry Science*, *99*(7), 3709–3722.

358 Fengou, L.-C., Lianou, A., Tsakanikas, P., Mohareb, F., & Nychas, G.-J. E. (2021). Detection of  
359 meat adulteration using spectroscopy-based sensors. *Foods*, *10*(4), 861.

360 Folmer, O., Black, M., Hoeh, W., Lutz, R., & Vrijenhoek, R. (1994). DNA primers for amplification  
361 of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates.  
362 *Molecular Marine Biology and Biotechnology*, *3* (3), 294–299.

363 Fuseini, A., Wotton, S. B., Knowles, T. G., & Hadley, P. J. (2017). Halal meat fraud and safety  
364 issues in the UK: a review in the context of the European Union. *Food ethics*, 1(2), 127–  
365 142.

366 Giusti, A., Armani, A., & Sotelo, C. G. (2017). Advances in the analysis of complex food matrices:  
367 Species identification in surimi-based products using Next Generation Sequencing  
368 technologies. *PLOS ONE*, 12(10), e0185586. <https://doi.org/10.1371/journal.pone.0185586>

369 Haider, N., Nabulsi, I., & Al-Safadi, B. (2012). Identification of meat species by PCR-RFLP of the  
370 mitochondrial COI gene. *Meat Science*, 90(2), 490–493.

371 Hassoun, A., Måge, I., Schmidt, W. F., Temiz, H. T., Li, L., Kim, H.-Y., Nilsen, H., Biancolillo, A.,  
372 Ait-Kaddour, A., & Sikorski, M. (2020). Fraud in animal origin food products: Advances in  
373 emerging spectroscopic detection methods over the past five years. *Foods*, 9(8), 1069.

374 Kennedy, O. B., Stewart-Knox, B., Mitchell, P., & Thurnham, D. (2004). Consumer perceptions of  
375 poultry meat: A qualitative analysis. *Nutrition & Food Science*.

376 Keyvan, E., İPLİKÇİOĞLU, G. Ç., ÇINAR, B. K., BİLGİN, N., & ŞİRELİ, U. T. (2017).  
377 Identification of meat species in different types of meat products by PCR. *Ankara*  
378 *Üniversitesi Veteriner Fakültesi Dergisi*, 64(4), 261–266.

379 Kumar, Y., Bansal, S., & Jaiswal, P. (2017). Loop-mediated isothermal amplification (LAMP): A  
380 rapid and sensitive tool for quality assessment of meat products. *Comprehensive Reviews in*  
381 *Food Science and Food Safety*, 16(6), 1359–1378.

382 Lianou, A., Papakonstantinou, M., Nychas, G.-J. E., & Stoitsis, J. (2021). Fraud in meat and poultry  
383 products. In *Food Fraud* (pagg. 85–108). Elsevier.

384 Lin, C.-C., Tang, P.-C., & Chiang, H.-I. (2019). Development of RAPD-PCR assay for identifying  
385 Holstein, Angus, and Taiwan Yellow Cattle for meat adulteration detection. *Food Science*  
386 *and Biotechnology*, 28(6), 1769–1777.

- 387 Marangoni, F., Corsello, G., Cricelli, C., Ferrara, N., Ghiselli, A., Lucchin, L., & Poli, A. (2015).  
388 Role of poultry meat in a balanced diet aimed at maintaining health and wellbeing: An  
389 Italian consensus document. *Food & nutrition research*, 59(1), 27606.
- 390 Marchetti, P., Mottola, A., Tantillo, G., Castrica, M., & Di Pinto, A. (2021). Detection of  
391 undeclared presence of bovine milk in buffalo yogurt. *Journal of Dairy Science*, 104(4),  
392 4056–4061.
- 393 Marques, M. L., Falcão, I., Labrador Horrillo, M., Falcão, H., & Cunha, L. (2021). *Milk and cow's*  
394 *meat allergy in a child: A clinical case*.
- 395 Mottola, A., Piredda, R., Catanese, G., Lorusso, L., Ciccarese, G., & Di Pinto, A. (2022). Species  
396 authentication of canned mackerel: Challenges in molecular identification and potential  
397 drivers of mislabelling. *Food Control*, 137, 108880.
- 398 Muñoz-Colmenero, M., Martínez, J. L., Roca, A., & Garcia-Vazquez, E. (2017). NGS tools for  
399 traceability in candies as high processed food products: Ion Torrent PGM versus  
400 conventional PCR-cloning. *Food chemistry*, 214, 631–636.
- 401 Ng, P. C., Ahmad Ruslan, N. A. S., Chin, L. X., Ahmad, M., Abu Hanifah, S., Abdullah, Z., &  
402 Khor, S. M. (2022). Recent advances in halal food authentication: Challenges and strategies.  
403 *Journal of Food Science*, 87(1), 8–35. <https://doi.org/10.1111/1750-3841.15998>
- 404 Nischala, S., Vaithyanathan, S., Ashok, V., Kalyani, P., Srinivas, C., Aravind Kumar, N., &  
405 Vishnuraj, M. (2022). Development of a Touchdown—Duplex PCR Assay for  
406 Authentication of Sheep and Goat Meat. *Food Analytical Methods*, 1–8.
- 407 OECD-FAO Agricultural Outlook 2021-2030. [https://www.fao.org/publications/oecd-fao-](https://www.fao.org/publications/oecd-fao-agricultural-outlook/2021-2030/en/)  
408 [agricultural-outlook/2021-2030/en/](https://www.fao.org/publications/oecd-fao-agricultural-outlook/2021-2030/en/). [accessed 27 April, 2022]

- 409 Palumbo, F., Scariolo, F., Vannozzi, A., & Barcaccia, G. (2020). NGS-based barcoding with mini-  
410 COI gene target is useful for pet food market surveys aimed at mislabelling detection.  
411 *Scientific reports*, *10*(1), 1–8.
- 412 Pan, Y., Qiu, D., Chen, J., & Yue, Q. (2020). Combining a COI Mini-Barcode with Next-  
413 Generation Sequencing for Animal Origin Ingredients Identification in Processed Meat  
414 Product. *Journal of Food Quality*, 2020.
- 415 Patro, J. N., Ramachandran, P., Barnaba, T., Mammel, M. K., Lewis, J. L., & Elkins, C. A. (2016).  
416 Culture-independent metagenomic surveillance of commercially available probiotics with  
417 high-throughput next-generation sequencing. *MSphere*, *1*(2), e00057-16.
- 418 Piredda, R., Mottola, A., Cipriano, G., Carlucci, R., Ciccarese, G., & Di Pinto, A. (2022). Next  
419 Generation Sequencing (NGS) approach applied to species identification in mixed processed  
420 seafood products. *Food Control*, *133*, 108590.  
421 <https://doi.org/10.1016/j.foodcont.2021.108590>
- 422 Preckel, L., Brünen-Nieweler, C., Denay, G., Petersen, H., Cichna-Markl, M., Dobrovolny, S., &  
423 Hochegger, R. (2021). Identification of Mammalian and Poultry Species in Food and Pet  
424 Food Samples Using 16S rDNA Metabarcoding. *Foods*, *10*(11), 2875.
- 425 Prosser, S. W., & Hebert, P. D. (2017). Rapid identification of the botanical and entomological  
426 sources of honey using DNA metabarcoding. *Food Chemistry*, *214*, 183–191.
- 427 Ren, J., Deng, T., Huang, W., Chen, Y., & Ge, Y. (2017). A digital PCR method for identifying and  
428 quantifying adulteration of meat species in raw and processed food. *PLoS One*, *12*(3),  
429 e0173567.
- 430 Ribani, A., Schiavo, G., Utzeri, V. J., Bertolini, F., Geraci, C., Bovo, S., & Fontanesi, L. (2018).  
431 Application of next generation semiconductor based sequencing for species identification in  
432 dairy products. *Food Chemistry*, *246*, 90–98.  
433 <https://doi.org/10.1016/j.foodchem.2017.11.006>

- 434 Roiter, L., Vedenkina, I., & Eremeeva, N. (2021). *Analysis of the market potential of poultry meat*  
435 *and its forecast. 937(2), 022104.*
- 436 Ropodi, A. I., Panagou, E. Z., & Nychas, G.-J. E. (2017). Multispectral imaging (MSI): A  
437 promising method for the detection of minced beef adulteration with horsemeat. *Food*  
438 *Control, 73, 57–63.*
- 439 Surowiec, I., Fraser, P. D., Patel, R., Halket, J., & Bramley, P. M. (2011). Metabolomic approach  
440 for the detection of mechanically recovered meat in food products. *Food Chemistry, 125(4),*  
441 *1468–1475.*
- 442 Vidal Junior, P. O., Menezes, A. C. R., de Souza, L. M. P., Guimarães, A. G., & de Cassia Vieira  
443 Cardoso, R. (2020). Trade and Safety Issues of Raw Beef from the Countryside of Bahia  
444 State, Brazil. *Journal of Public Health Research, 9(3), jphr.2020.1752.*  
445 <https://doi.org/10.4081/jphr.2020.1752>
- 446 Walker, M. J., Burns, M., & Burns, D. T. (2013). Horse meat in beef products—Species  
447 substitution 2013. *Journal of the Association of Public Analysts, 41, 67–106.*
- 448 Wilson, J. M., & Platts-Mills, T. A. (2018). Meat allergy and allergens. *Molecular immunology,*  
449 *100, 107–112.*
- 450 Wilson, J. M., & Platts-Mills, T. A. (2019). Red meat allergy in children and adults. *Current*  
451 *opinion in allergy and clinical immunology, 19(3), 229.*
- 452 Wirta, H., Abrego, N., Miller, K., Roslin, T., & Vesterinen, E. (2021). DNA traces the origin of  
453 honey by identifying plants, bacteria and fungi. *Scientific reports, 11(1), 1–14.*
- 454 Xing, R.-R., Wang, N., Hu, R.-R., Zhang, J.-K., Han, J.-X., & Chen, Y. (2019). Application of next  
455 generation sequencing for species identification in meat and poultry products: A DNA  
456 metabarcoding approach. *Food Control, 101, 173–179.*

457 Zin, Z. M., Sarbon, N. M., Zainol, M. K., Jaafar, S., Shukri, M. M., & Rahman, A. H. A. (2021).

458 *Halal and Non-Halal Gelatine as a Potential Animal By-Products in Food Systems:*

459 *Prospects and Challenges for Muslim Community.* 536, 530–540.

460