

1 **Fat globule membranes in ewes' milk: the main enzyme activities during lactation**

2 Mina Martini*¹, Iolanda Altomonte¹, Rossana Pesi², Maria Grazia Tozzi², Federica Salari¹

3 ¹Physiological Science Department, University of Pisa, Viale delle Piagge 2 – 56124 Pisa (Italy)

4 ²Biology Department, University of Pisa, via San Zeno 51, 56127 Pisa (Italy)

5 *mmartini@vet.unipi.it

6 phone number +39502216897 fax +39 050.2216901

7

8 **Abstract**

9 Studies on milk fat globule membranes (MFGMs) have primarily been carried out on human and
10 bovine milk. An investigation of the proteins in sheep MFGM during lactation could provide
11 information regarding the role of MFGM enzymes and milk quality. A trial was carried out on
12 seven Massese ewes, from which individual milk samples were taken during lactation. All the
13 enzyme activities investigated (oxidase activity of xanthine oxidoreductase, gamma-
14 glutamyltranspeptidase, alkaline phosphatase and 5'-nucleotidase) were affected by the lactation
15 phase, with the exception of the dehydrogenase activity of xanthine oxidoreductase (XOR). A
16 higher oxidase activity (XO) of XOR was found when the diameter of milk fat globules was
17 smaller. In addition, the absence of XO in colostrum led to the hypothesis that its antibacterial
18 role is independent of its activity. The increase in alkaline phosphatase activity during lactation
19 requires further research in order to better define the criteria for pasteurized sheep milk.

20

21 **1. Introduction**

22 Milk fat globules (MFGs) are enveloped by a triple membrane, called the milk fat globule
23 membrane (MFGM). The MFGM consists of a single inner layer originating from the
24 endoplasmic reticulum and a double layer, arising from the membrane of the secretory cell
25 (Mather & Keenan, 1998). The amount of MFGM in milk changes depending on the number and
26 diameter of globules (Martini, Salari, Pesi, & Tozzi, 2010). The MFGM supplies many proteins
27 and enzymes, some of which have beneficial biological effects (Dewettinck, Rombaut,
28 Thienpont, Le, Messens, & Van Camp, 2008), whereas others have been linked to human
29 diseases (Riccio, 2004).

30 Research on MFGMs has mainly been carried out on cows' and human milk (Mather, 2000;
31 Liao, Alvarado, Phinney & Lönnerdal, 2011), whereas there have only been a few studies on
32 MFGM proteins in sheep (Martini, Salari, Pesi & Tozzi, 2010; Pisanu, et al., 2011).

33 To date, the exact physiological role of many MFGM enzymes has not been fully
34 understood. The aim of this study was to investigate the activities of MFGM enzymes during

lactation in order to check their link with the size of the native milk fat globules secreted and with colostrum and milk nutritional and hygienic quality.

2. Materials and methods

2.1. Animals and sampling

A trial was carried out on seven Massese reared in Tuscany (Central Italy). All ewes were reared on the same farm and were homogeneous in terms of parity and feeding. The subjects were kept indoors at the tenth day before partum. The experiment lasted 120 days from 10 hours postpartum (day 0); 11 samples of milk were taken from each individual during lactation, producing a total of 77 samples.

2.2. Morphometric analysis of milk fat globules

The diameter and number of MFGs per mL were measured following the method of Scolozzi, Martini, & Abramo (2003). Our method enables the MFGs to be characterized directly, using an image analyser system, whereas other methods involve an indirect analysis using the refractive index (Michalski, Briard, & Michel, 2001). The detection limit of the microscope and the image analyser do not allow globules smaller than 0.4 μm to be measured. Although globules with a diameter $<1 \mu\text{m}$ are numerous, and comprise 80% of the globules (Heid & Keenan, 2005) they only represent a few percent of the total milk fat volume (Walstra, 1969; Michalski, Ollivon, Briard, Leconte & Lopez, 2004).

2.3. Extraction MFGM proteins and enzyme assays

The cream was isolated from the skim following a macroversion of Patton and Huston's method (1986). This method, carried out on fresh milk, enabled us to isolate the MFGs, thus avoiding the denaturation of the MFGM proteins. To obtain an extract containing MFGM proteins, the cream was gently stirred for two hours in the presence of 1% triton X-100, and then centrifuged at 100,000xg for two hours at 10°C (Camici, Fini, & Ipata, 1985 modified). Three layer were obtained after the centrifugation: a pellet made up of MFGM lipids, a supernatant containing the MFGM proteins and an upper layer made up of the triglycerides of the core.

Total membrane proteins (TMP) were measured in the supernatant according to Bradford (1976). In the supernatant from each sample, an assay was made in duplicate of the dehydrogenase activity (XDH) of the enzyme XOR, oxidase activity (XO) of the XOR, gamma-glutamyltranspeptidase (γ -GT) alkaline phosphatase (AP), and 5' nucleotidase (5'-N) activities according to Benboubetra, Baghiani, Atmani, and Harrison (2004), Bergmeyer (1983), Huseby and Strömme (1974) and Fini, Camici, Minelli, Floridi, and Ipata (1986), respectively.

69

70 2.4. Statistical analysis

71 The results of the morphometric analysis of the MFGs and of the enzyme activities were
72 analysed following ANOVA for repeated measurements, regarding the sampling time as a fixed
73 effect and the subject as a random effect. To verify the relationships between enzymes and
74 MFGs size, Pearson's correlation method was applied. Significant differences were considered at
75 $P \leq 0.05$. The statistical analysis was carried out using JMP software (2002).

76

77 3. Results and discussion

78 A greater amount of TMP was detected at 10 hours post partum (4.23 mg mL⁻¹ of
79 supernatant; Table 1). XO of the XOR, a protein that derives from the endoplasmic reticulum
80 (Bianchi et al., 2009), was not observed in the mammary secretion in the first few hours (day 0;
81 Table 1). This was probably due either to the lower expression of XOR in colostrum as reported
82 in the literature for cow and human milk (Reinhardt & Lippolis, 2008; Liao et al., 2011) or to the
83 activation and deactivation of the enzyme during lactation as found by Benboubetra et al. (2004).
84 In addition, XOR has been reported as having anti-bacterial properties by catalyzing chemical
85 reactions (Harrison, 2006). In any case, we did not detect XO at the day 0. Thus, we
86 hypothesized that the anti-bacterial properties of the enzyme are not due to this activity, at least
87 at the beginning of lactation when the defence role of the newborn mainly involves other
88 components such as immunoglobulins.

89 XO increased during lactation (Table 1) after the first 10 hours and 15th day ($P \leq 0.01$),
90 reaching a climax on the 45th day (3.06 mU mg⁻¹), corresponding to a decrease in the average
91 diameter of the measured MFGs (Figure 1). In addition, a negative Pearson's correlation between
92 XO and the diameter of MFGs (-0.159; $P \leq 0.01$) was found, confirming the positive link
93 between XO and smaller globules found in a previous study (Martini et al., 2010). This suggests
94 that the link between XO and the diameter of MFGs is probably due to the fact that the enzyme
95 XOR could be involved in the secretion of the globules (McManaman, Russell, Schaack,
96 Orlicky, & Robenek, 2007).

97 In fact, XO is supposed to control MFGs secretion either by interactions with butyrophilin
98 and adipophilin (Mather & Keenan, 1998) or through the generation of reactive oxygen species
99 which work as signals for butyrophilin. The typical structure of receptor of the protein
100 butyrophilin support this hypothesis (Ogg, Weldon, Dobbie, Smith, & Mather, 2004). Since the
101 intracellular precursors of the MFGs (cytoplasmic droplets) increase in size by intracellular
102 fusions, the higher XO would increase the speed of release of the globules by the mammary

103 cells, resulting in the secretion smaller MFGs. In contrast, XDH did not vary significantly during
104 lactation, as previously described in bovine milk by Heinz and Reckel (1983), who reported that
105 the mammary gland and milk contain the enzyme mainly as XO.

106 AP is the most important enzyme indicator for the correct pasteurization of milk (Rankin,
107 Christiansen, Lee, Banavara, & Lopez-Hernandez, 2010). In our study, AP showed a progressive
108 increase in activity during lactation, at 90 days reaching a climax that was about 40 times higher
109 than the initial value of activity (day 0), as reported for ovine milk (Rankin et al., 2010). Since
110 the legal limit for the test in terms of negativity for the activity of AP is currently defined only
111 for cows' milk (Rankin et al., 2010) and since there is a considerable variation in AP activity
112 with advancing lactation, further research is required in order to better define the extent of heat
113 treatment in sheep milk.

114 Transmembrane glycoprotein 5'-N is an enzyme that plays a major role in the catabolism
115 of mononucleotides and dinucleotides (Fini et al., 1986), however its role in milk has not yet
116 been clarified. In our study, the activity of 5'-N showed an increasing trend during lactation,
117 from a minimum of 4.77 to a maximum of 47.34 mU mg⁻¹ of MFGM proteins on the 3rd and
118 60th day respectively (Table 1).

119 γ -GT is known to be one of the colostrum enzymes and its activity is positively correlated
120 with the content of immunoglobulins in ruminants, therefore it is considered to be indicative of
121 the quality of this initial secretion (Zarrilli et al., 2003). In fact, in our study, γ -GT showed the
122 maximum activity in the colostrum phase, with peaks of activity between the third and tenth days
123 (Table 1), decreasing, and then remaining constant thereafter.

124

125 4. Conclusions

126 All the enzyme activities assayed, with the exception of XDH were affected by lactation. A
127 higher XO was found when the diameter of the milk fat globules is smaller. With regard to AP,
128 the significant increase in its activity during lactation requires further research in order to better
129 define the pasteurization criteria of sheep milk.

130

131 Acknowledgements

132 This work was supported by the Research Project of National Interest (PRIN 2007).

133

134

135 **References**

- 136
- 137 Benboubetra, M., Baghiani, A., Atmani, D., & Harrison, R. (2004). Physicochemical and kinetic
 138 properties of purified sheep's milk xanthine oxidoreductase. *Journal of Dairy Science*, 87,
 139 1580–1584.
- 140 Bergmeyer, H. U. (1983). *Methods of enzymatic analysis*. Deerfield Beach, FL, USA: Verlag
 141 Chemie Academic Press.
- 142 Bianchi, L., Puglia, M., Landi, C., Matteoni, S., Perini, D., Armini, A., Verani, M., Trombetta,
 143 C., Soldani, P., Roncada, P., Greppi, G., Pallini, V., & Bini, L. (2009). Solubilization methods
 144 and reference 2-DE map of cow milk fat globules. *Journal of Proteomics*, 72, 853 – 864.
- 145 Bradford, MM. (1976). A rapid and sensitive method for the quantitation of microgram
 146 quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*,
 147 72, 248–254.
- 148 Camici, M., Fini, C., & Ipata, P.L. (1985). Isolation and kinetic properties of 5'-nucleotidase
 149 from guinea pig skeletal muscle. *Biochimica et Biophysica Acta*, 840, 6-12.
- 150 Dewettinck, K., Rombaut, R., Thienpont, N., Le, T. T., Messens, K., & Van Camp, J. (2008).
 151 Nutritional and technological aspects of milk fat globule membrane material. *International*
 152 *Dairy Journal*, 18, 436–457.
- 153 Fini, C., Camici, M., Minelli, A., Floridi, A., & Ipata, P. L. (1986). Concanavalin A induced
 154 inhibition of 5'-nucleotidase from guinea pig skeletal muscle and bull seminal plasma: a
 155 comparative study. *International Journal of Biochemistry*, 18, 683–689.
- 156 Harrison, R. (2006). Milk xanthine oxidase: Properties and physiological roles. *International*
 157 *Dairy Journal*, 16, 546-554.
- 158 Heid, H.W., & Keenan, T.W. (2005). Intracellular origin and secretion of milk fat globules.
 159 *European Journal of Cell Biology*, 84, 245-258.
- 160 Heinz, F., & Reckel, S. (1983). Xanthine Oxidase. In Bergmeyer, H.U. (Ed.), *Methods of*
 161 *Enzymatic Analysis* (3rd ed.) (pp. 210-216) New York, NY, USA: Academic Press.
- 162 Huseby, N. E., & Strömme, J. H. (1974). Practical points regarding routine determination of
 163 gamma-glutamyl transferase (γ -GT) in serum with a kinetic method at 37 °C. *Scandinavian*
 164 *Journal of Clinical and Laboratory Investigation*, 34, 357–361.
- 165 JMP. (2002). *User's guide* (version 5.0 for PC). SAS Institute: Cary, NC, USA.
- 166 Liao, Y., Alvarado, R., Phinney, B. & Lönnnerdal, B. (2011). Proteomic Characterization of
 167 Human Milk Fat Globule Membrane Proteins during a 12 Month Lactation Period. *Journal of*
 168 *Proteome Research*, 10, 3530–3541.
- 169

170 Martini M., Salari, F., Pesi, R., & Tozzi, M. G. (2010). Relationship between activity of some fat
 171 globule membrane enzymes and the lipid fraction in ewes' milk: Preliminary studies.
 172 *International Dairy Journal* 20, 61–64

173 Mather, I. H. (2000). A review and proposed nomenclature for major proteins of the milk-fat
 174 globule membrane. *Journal of Dairy Science*, 83, 203–247.

175 Mather, I. H., & Keenan, T. W. (1998). Origin and secretion of milk lipids. *Journal of Mammary*
 176 *Gland Biology and Neoplasia*, 3, 259–273.

177 McManaman, J. L., Russell, T. D., Schaack J., Orlicky, D. J., Robenek, H. (2007). Molecular
 178 determinants of milk lipid secretion. *Journal of Mammary Gland Biology and Neoplasia* 12,
 179 259–268.

180 Michalski, M. C., Briard, V., & Michel, F. (2001). Optical parameters of milk fat globules for
 181 laser light scattering measurements. *Lait*, 81, 787–796.

182 Michalski, M. C., Ollivon, M., Briard, V., Leconte, N., & Lopez, C. (2004). Native fat globules
 183 of different sizes selected from raw milk: thermal and structural behavior. *Chemistry and*
 184 *Physics of Lipids*, 132, 247–261.

185 Ogg, S. L, Weldon A. K., Dobbie L., Smith A. J. H., & Mather I. H. (2004). Expression of
 186 butyrophilin (Btn1a1) in lactating mammary gland is essential for the regulated secretion of
 187 milk–lipid droplets. *PNAS*, 27, 10084 –10089.

188 Patton, S., & Huston, G. E. (1986). A method for isolation of milk fat globules. *Lipids*, 21, 170–
 189 174.

190 Pisanu, S., Ghisaura, S., Pagnozzi, D., Biosa, G., Tanca, A., Roggio T., Uzzau S. & Addis M. F.
 191 (2011). The sheep milk fat globule membrane proteome. *Journal of Proteomics*, 74, 350-358.

192 Rankin, S.A., Christiansen, A., Lee, W., Banavara, D. S., & Lopez-Hernandez, A. (2010). The
 193 application of alkaline phosphatase assays for the validation of milk product pasteurization.
 194 *Journal of Dairy Science*, 93, 5538–5551.

195 Reinhardt, T.A., & Lippolis, J. D. (2008). Developmental Changes in the Milk Fat Globule
 196 Membrane Proteome During the Transition from Colostrum to Milk. *Journal of Dairy Science*
 197 91, 2307–2318.

198 Riccio, P. (2004). The proteins of the milk fat globule membrane in the balance. *Trends in Food*
 199 *Science and Technology*, 15, 458–461.

200 Scintu, M. F., Daga, E., & Ledda, A. (2000). Evaluation of spectrophotometric and fluorometric
 201 methods for alkaline phosphatase activity determination in ewe's milk. *Journal of Food*
 202 *Protection*, 63, 1258–1261.

203 Scolozzi, C., Martini, M., & Abramo, F. (2003). A method for identification and characterization
 204 of ewe's milk fat globules. *Milchwissenschaft*, 58, 490–493

205 Walstra, P., (1969). Studies on milk fat dispersion. The globule size distribution of cow's milk.
206 *Netherlands Milk and Dairy Journal*, 28, 3-9.

207 Zarrilli, A., Micera, E., Lacarpia, N., Lombardi, P., Pero, M. E., Pelagalli, A., d'Angelo, D.,
208 Mattia, M., & Avallon, L. (2003). Evaluation of goat colostrum quality by determining
209 enzyme activity level. *Livestock Production Science*, 83, 317–320.

210

211 Table 1 Activities of milk fat globules membrane enzymes (U mg⁻¹ of milk fat globule membrane (MFGM) proteins) and total membrane proteins (TMP; mg mL⁻¹
 212 of supernatant) in ewes' milk during lactation.

213

		Days in milk											SEM ^a
		10 h	3	6	10	15	20	30	45	60	90	120	
XO ^b	(mU mg ⁻¹)	ND	0.50B	0.70B	1.23B	0.62B	1.81A	1.75AB	3.06A	2.78A	1.87A	0.86B	0.001
XDH ^c	(mU mg ⁻¹)	0.76	0.60	0.18	0.78	0.11	0.52	0.92	1.15	0.87	0.66	0.25	0.001
γ-GT ^d	(U mg ⁻¹)	2.83b	4.91a	4.90a	4.74a	3.31ab	3.00b	2.77b	2.86b	3.64ab	3.19ab	3.51ab	1.656
AP ^e	(mU mg ⁻¹)	3.79C	7.54C	21.42C	22.73C	31.79BC	72.58BC	81.37B	128.33AB	148.83A	150.56A	140.76A	0.072
5'-N ^f	(mU mg ⁻¹)	5.22B	4.77B	9.70B	12.51B	14.42B	19.82B	20.99B	28.30AB	47.34A	34.17A	22.84B	0.015
TMP	(mg mL ⁻¹)	4.23A	2.56B	2.15BC	3.10B	2.16B	1.92C	2.06C	2.18B	2.04C	2.32B	2.69B	0.771

214 ^a Standard error of the model

215 ^b Oxidase activity of the enzyme xanthine oxidoreductase

216 ^c Dehydrogenase activity of the enzyme xanthine oxidoreductase

217 ^d Gamma-glutamyl transpeptidase

218 ^e Alkaline phosphatase

219 ^f 5'-nucleotidase

220 ^g Not determined.

221 Different letters indicate statistical differences across a row at $P \leq 0.01$ (A, B, C) and $P \leq 0.05$ (a, b).

222 Figure 1. Average diameter (µm) and number per mL of milk fat globules during lactation.
223 Different letters indicate statistical differences among bars at $P\leq0.01$

