

Vaporization kinetic study of lavender and sage essential oils.

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Abstract

The thermal behavior of lavender and sage essential oils (EOs) and also that of their main components (camphor and 1,8-cineole) were studied by thermogravimetry (TG) at different scan rates under inert gas atmosphere up to about 140 °C. All the samples investigated undergo a single-step evaporation starting from ambient temperature without evidence of side decomposition processes. On the basis of the onset temperatures (T_{on}) extrapolated from the TG curves the following increasing stability trend is assessed:

1,8-cineole < sage < lavender < camphor.

However, the difference of T_{on} values among the first three samples was found to be slight. To the aim of assessing a thermal stability scale among the samples, a kinetic analysis of evaporation was performed. A first approach was based on a simple model-fitting kinetic method, widely used in the past for studying evaporation of EOs. The results were compared with those obtained with a more reliable approach based on the integral isoconversional method by Kissinger–Akahira–Sunose in which the activation energy (E_a) versus reaction extent (α) plot of each sample was obtained. Because of the compensation effect, neither activation energy nor the pre-exponential factor, alone, could be considered to compare the stabilities. Lastly, the Arrhenius kinetic rate constant values at given temperatures within the experimental range were calculated, and camphor was found to be the more stable component, while lavender seems to vaporize faster than any other sample tested.

Keywords Essential oils, Lavender, Sage, 1,8-cineole, camphor, TG, Modulated TG, Vaporization kinetics, Kissinger-Akahira-Sunose method

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32 **Introduction**

33 Essential oils (EOs), also called volatile or ethereal oils, are hydrophobic liquids containing volatile
34 compounds obtained from plant material (flowers, buds, seeds, leaves, twigs, bark, herbs, wood, fruits
35 and roots) that have gained great popularity as alternatives for artificial additives or
36 pharmacologically relevant agents in food, cosmetic and pharmaceutical industries or simply as
37 flavoring material.

38 Sage and lavender are spontaneous herbs belonging to the Lamiaceae family and their EOs, whose
39 composition includes in both cases camphor and 1,8-cineole as principal components, show important
40 properties. In particular, sage possesses well-recognized biological activity as antibacterial,
41 antioxidant [1] and anti-inflammatory agent [2], and lavender exhibits antioxidant activity in
42 conservation of food [3] and pharmaceutical properties, as anticonvulsive [4], antidepressant [5],
43 antiviral [6], antibacterial [3] and antifungal [7, 8].

44 Although the use of EOs dates back to 4500 BC, it is specially from the Middle Ages that they have
45 become common in the everyday life, due to the development by the Arabs of the steam (SD) or
46 hydro-distillation (HD) techniques, which still remain the main methods of extraction of EOs.

47 During the last years, EOs have expanded their applications in specific fields as therapeutic agents in
48 biomedical [9] or in new applications in nanotechnology or synthesis [10], justifying the efforts to
49 develop efficient and low-cost methodologies to isolate these valuable renewable products from
50 different herbs. In this respect, it is noteworthy that the yield, extraction time and chemical
51 composition of EOs are influenced not only by parameters like growing environmental conditions,
52 genetic variety and organ of the plant sample, but also by the extraction approach used to their
53 isolation.

54 Novel extraction techniques, namely “green extractions,” have been developed in order to maximize
55 the yield percent, while the time processing and energy demand are reduced. The success of these
56 approaches is based on their capability to produce EOs with the same quality and sensorial properties
57 as those obtained by the conventional techniques but in a faster and cheaper way. In this respect,

microwave (MW)-assisted extraction technology can be considered a convenient methodology to obtain essential oils from several herbs [11,12,13,14,15,16].

Currently, lavender and sage EOs are primarily produced from leaves and flowers by conventional extraction methods, namely HD and SD [17]. Although the distillation techniques are both, simple and low cost, there are severe limitations in their use because of thermolability of compounds (high operating temperature), hydrolysis of water-sensitive compounds (prolonged extraction time) and incomplete extraction.

To the aim to overwhelm these limitations, our innovative MW-assisted HD method, where the electromagnetic energy is applied directly inside the aqueous medium by a coaxial dipole antenna, was successfully applied to extraction of EOs from sage and lavender [14].

At the same time, we realized that, in order to furnish a rational base to the prevention of the bioactivity of sage and lavender EOs, of their characteristic aromatic properties, and to validate the efficiency of our extraction method, the thermal stability and the rate of volatilization of EOs and of their principal pure components should be carefully investigated.

As a matter of fact, although the vapor pressure is the fundamental quantity determining the volatility of a pure compound [18], in the case of a complex mixture of volatiles, such as EOs [19], the vapor composition at equilibrium also reflects the molecular interactions determining, in turn, the activity coefficients of each component. Moreover, the vaporization of a volatile mixture can occur in non-equilibrium conditions giving rise to a situation, even more difficult to predict, where the volatility of each component is affected by the corresponding evaporation rate [20]. Finally, degradation (pyrolytic or oxidative) of one or more components can also occur. Each of the above-mentioned aspects proves crucial in ruling the prevention of original properties of EOs and for their safe and proper handling during product formulation.

Many studies have been performed in the past, aimed to the determination of the evaporation rate at ambient temperature or slightly higher. For instance, Rudolfi and co-worker developed a rapid

83 method for determining the evaporation rate of EOs using a conventional gas chromatographic
84 technique [20].

85 In the recent past, we successfully used thermal analysis techniques to investigate the drying oils with
86 particular reference to their drying, polymerization and oxidative degradation [21, 22] and EOs'
87 thermal stabilities [14]. However, in the literature few studies report a comprehensive analysis of
88 thermal behavior, vaporization characteristic and stability of EOs [23,24,25,26]. Hazra et al. [23, 24]
89 dealt with the vaporization kinetics of different EOs and their main components by carrying out single
90 thermogravimetric experiment (TG) at a single heating rate, assuming a zero-order reaction by simply
91 observing the profiles of the corresponding derivative curves (DTG) and by using a model-fitting
92 approach. However, the reliability of this approach was found questionable by valuable members of
93 the Kinetic Committee of the International Confederation of Thermal Analysis and Calorimetry
94 (ICTAC) in several papers and reviews published in 2000 [27], 2011 [28] and 2014 [29].

95 In this study, the thermal behavior of lavender and sage EOs and of their main components (camphor
96 and 1,8-cineole) were investigated by TG at different heating rates, similarly to what has been made
97 in the past with commercial vegetable oils by Dweck and Sampaio [30]. Subsequently, aiming at
98 assessing a thermal stability scale among this series of EOs and related components a kinetic analysis
99 of evaporation was performed. To this purpose, we decided first to check the reliability of a simple
100 model-fitting method (as made in the past [23, 24]) and compare the results with those obtained with
101 a more reliable approach based on the integral isoconversional method by Kissinger–Akahira–Sunose
102 (KAS) [31, 32]. This approach was successfully applied to the vaporization kinetics of the pesticides
103 such as alachlor and metolachlor whose activation energy was found substantially constant over the
104 whole range of the extent of reaction, α [33]. After the kinetic analysis of evaporation of the four
105 samples (lavender, sage, 1,8-cineole and camphor) was performed and the corresponding stability
106 scale was assessed, a comparison with that based on the extrapolated onset temperature was made.

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109 **Theoretical background of Kinetic Analysis**

110 Substances in the condensed phase (like EOs or EOs components) may undergo vaporization with or
 111 without decomposition, and the reaction rate can be monitored as a function of temperature (from
 112 non-isothermal and constant heating rate, $\beta = dT/dt$, TG experiments) through the change in the extent
 113 of reaction, defined as $\alpha = (m_0 - m_T)/(m_0 - m_f)$, where m_0 , m_T and m_f are the initial sample mass, the
 114 sample mass at a given temperature and the sample mass at the end of a reaction, respectively. The
 115 kinetic approach starts by assuming that the general equation describing the rate of a reaction
 116 involving a substance in the condensate state $d\alpha/dt$ is a function of both the two variables T and α :

$$117 \quad \quad \quad d\alpha/dt = k(T)f(\alpha) \quad (1)$$

118 where the $k(T)$ temperature function often has the meaning of the Arrhenius specific rate constant and
 119 $f(\alpha)$ is the so-called differential model function, whose mathematical expressions representing the
 120 reaction mechanisms are available from literature [29, 34, 35]. For a non-isothermal TG experiment
 121 under constant heating rate, using the Arrhenius equation for the temperature dependence of $k(T)$, it
 122 yields:

$$123 \quad \quad \quad d\alpha/dt = (d\alpha/dT)\beta = A \exp(-E_a/RT)f(\alpha) \quad (2)$$

124 where A and E_a are the pre-exponential factor and the activation energy, respectively, and along with
 125 the selected $f(\alpha)$ are sometimes called the kinetic triplet. Rearranging Eq. (2) with the view to separate
 126 the two variables T and α , and considering that $g(\alpha) = \int d\alpha/f(\alpha)$:

$$127 \quad \quad \quad g(\alpha) = (A/\beta) \int_0^T \exp(-E_a/RT) dT = (AE_a/R\beta) \int_x^\infty \exp(-x)/x^2 dx = (AE_a/R\beta) p(x), \quad (3)$$

128 where $g(\alpha)$ is called the integral model function and $x = E_a/RT$, and the integration limits were
 129 correspondingly transformed. Since the temperature integral $p(x)$ in Eq. (3) has no exact mathematical
 130 solution it is often replaced by approximations, one of the most common being the asymptotic series
 131 expansion considered in the Coats-Redfern (CR) model-fitting method [36, 37], represented by the
 132 following Eq. (4):

$$133 \quad \quad \quad \ln[g(\alpha)/T^2] = \ln\{(AR/\beta E_a)[1 - (2R\langle T \rangle/E_a)]\} - E_a/RT, \quad (4)$$

134 where $\langle T \rangle$ is the average of the experimental temperature range. From the intercept and slope of the
 135 linear regression obtained by plotting the left hand side of Eq.(4) against the reciprocal absolute
 136 temperature the corresponding A and E_a have been determined, respectively. The most suitable model
 137 function(s) can be selected on the bases of the best linear fit of Eq. (4). Once the so-called kinetic
 138 triplet is determined using this model-fitting method, the α vs. T curve can be reconstructed starting
 139 from α_0 (equal to 0.025 in this study) and using as differential increments of α the expression obtained
 140 rearranging Eq. (2):

$$141 \quad d\alpha_j = A_j/\beta \exp(-E_{a,j}/RT) f_j(\alpha) dT \quad (5)$$

142 where the subscript j refers to the mechanism selected according to the procedure previously
 143 described. A good agreement between the experimental and reconstructed α vs. T curves should
 144 confirm the reliability of the approach used. By contrast, model-free methods that use TG data
 145 obtained from experiments performed under different heating rates were found to provide more
 146 reliable results. Among them one of the most popular is the integral isoconversional method firstly
 147 proposed by Kissinger-Akahira-Sunose [30], based on the following equation:

$$148 \quad \ln(\beta/T_\alpha^2) = \text{Const} - E_\alpha/RT_\alpha. \quad (6)$$

149 At each given value of α , a single value of activation energy can be calculated from the slope of the
 150 regression line obtained by plotting the left hand side of Eq. (6) against the reciprocal isoconversional
 151 temperature (T_α). By applying the CR model-fitting method a pair of A and E_a values can be obtained
 152 for each reaction model using a single-heating rate experiment. Wide ranges of pair values were
 153 usually found when all the usual reaction models were considered, and a strong linear correlation
 154 (called compensation effect) is found between them in the following form:

$$155 \quad \ln A_i = aE_i + b \quad (7)$$

156 where the subscript i refers to each of all the reaction model. Once a and b parameters have been
 157 determined at each heating rate using a linear regression procedure, these values were replaced in Eq.
 158 (7) by the $\langle a \rangle$ and $\langle b \rangle$ values, as averages of the pair values obtained in the five replicates (one for

each TG experiment at a given value of β). Then, the E_i values were replaced by the isoconversional E_α values in order to determine the corresponding $\ln A_\alpha$ values at each given value of α [38].

161

162 **Experimental**

163 **Materials**

164 The leaves and stems of fresh lavender and sage plants were collected in October 2014 from a
165 courtyard located in the city of Pisa, (Tuscany, Italy). These raw plant materials were harvested daily
166 during the experimental period and used as substrate to obtain their EO. Deionized water obtained
167 with a Milli-Q system (Millipore, Bedford, MA, USA) was used as solvent for the EO extraction.

168 **Instruments**

169 The TG/DTG experiments were carried out using a TA Instruments Thermobalance model Q5000IR.
170 TG measurements were performed from about 25 to 150 °C under inert flowing nitrogen atmosphere
171 (25 mL min⁻¹) using Pt crucibles. Five TG experiments at different heating rates (2, 4, 6, 8 and 10 K
172 min⁻¹) were performed for each sample, being the experiment at 2 K min⁻¹ that used to study the
173 thermal behaviour. Mass calibration was performed using certified mass standards, in the range from
174 0 to 100 mg, supplied by TA Instruments. The amount of sample in each measurement varied between
175 1.2 and 1.5 mg. Temperature calibration was based on the Curie point of paramagnetic metals. A
176 multipoint calibration with five Curie points from reference materials (Alumel, Ni, Ni83%Co17%,
177 Ni63%Co37%, Ni37%Co63%) was performed.

178

179 **Results and discussion**

180 The EOs were analyzed by gas chromatography (GC-FID) and gas chromatography-mass
181 spectrometry (GC-MS) techniques in a previous work [14] and their compositions are shown in Fig.
182 1.

183 The TG/DTG curves of the oily samples of lavender, sage, 1,8-cineole and camphor carried out at 2
184 K min⁻¹ under a nitrogen flow are displayed in Fig. 2. All the samples undergo only one step of mass
185 loss starting from ambient temperature without appreciable differences among them, except for
186 lavender, for which two distinguished and consecutive steps of mass loss are found. In order to better
187 study these processes, the peaks of reaction rate, $d\alpha/dT$, were deconvoluted (Fig. 3) using two
188 Gaussian peaks exponentially modified to make them asymmetrical. A multiparameter least-square
189 approach was used to determine the best fit between the models (in blue and red in Fig. 3b for the
190 first and the second peaks, respectively). The first one, who involves about 80% of the mass loss from
191 ambient temperature to 75.6 °C, was studied in detail from a kinetic point of view, and the results
192 obtained were compared with those deriving from the single step of the others. So, after the TG data
193 have been collected at all heating rates (from 2 to 10 K min⁻¹) the extrapolated onset vaporization
194 temperatures (T_{on}) were determined with the aim to find a parameter able to assess a stability scale
195 among the samples tested. The plot of the T_{on} values as a function of heating rate (β) is given in Fig.
196 4 along with the regression (dotted) lines. The value for lavender is referred to the first step, even if
197 no difference is found (within the estimated temperature uncertainties) with the value obtained using
198 the TG portion related to the first step only. As it can be seen, extrapolation of these values at null
199 heating rate provides similar values around 30 °C for sage, lavender and 1,8-cineole, while only the
200 value of camphor is higher than 40 °C, thus suggesting that this could be the more stable among the
201 oily samples investigated. However, this result could be misinterpreted since it is mainly due to the
202 difference in the shapes of their TG curves, with particular reference to its slowest initial mass loss
203 with respect to those of the other three samples (Fig. 2): Camphor loses 20% of its mass in a
204 temperature range of about 20–25 °C, while the other three give the same result in only 5–7 °C. On
205 the other hand, looking carefully at the slopes of the linear portion of the TG curves around the
206 inflection point, that of 1,8-cineole is more negative than those of lavender and sage which, in turn,
207 is similar to that of its main component camphor (see Fig. 2). Since the slopes are expected to be

208 proportional to the rates of vaporization, the above observation suggests that 1,8-cineole vaporizes
209 faster than the other samples.

210 In any case, the thermal behaviour study of these samples must be considered preliminary, but not
211 sufficient to provide exhaustive information on their stabilities. So, we carried out a kinetic analysis
212 of vaporization of these samples, starting with a model-fitting approach very similar to that already
213 applied in the past [23, 24]. To this purpose, the CR method, based on processing data from a single
214 TG run according to Eq. (4), was considered. Four best model functions were selected for
215 vaporization of all samples on the basis of the best linear fit (the highest values of R^2 , squares of the
216 correlation coefficients) and reported in Table 1 along with the Arrhenius parameters (E_a and A). The
217 first set of data were determined from TG experiments performed at 2 K min^{-1} , while a second set of
218 data were obtained from TG runs carried out at 10 K min^{-1} , with the view to verify possible
219 interferences of the heating rate. The same four models were selected for the vaporization of the four
220 samples regardless of the heating rate considered. Moreover, no significant differences can be
221 attributed to the four R^2 values, thus concluding that the four models should be considered equivalent,
222 even though the pair values associated to each model were found, especially in some cases,
223 remarkably different. In order to find the most suitable models, a reconstruction of the α vs. T curves
224 was made, according to Eq. (5), and compared to the experimental one (Fig. 5) at the lowest and
225 highest heating rates considered. All the reconstructed curves fit quite well with the experimental
226 ones in a significant range of extent of conversion, except for the case of sage according to model D3
227 (plots c, d), showing a remarkable deviation from the corresponding experimental curves. This
228 behavior does not allow to univocally select the most suitable model, thus indicating that this
229 approach fails to provide a thermal stability scale among the investigate EOs (Table 2).

230 A comparison with the literature was only possible for lavender EO whose evaporation was found to
231 reach 50% at $76\text{ }^\circ\text{C}$ (constant temperature) in 13 min [20]. In our study, this percentage of
232 vaporization was reached between 50 and $75\text{ }^\circ\text{C}$ depending on the heating rate used (2 and 10 K min^{-1}
233 in Fig. 5a, b plots, respectively).

234 Dollimore and co-worker claimed that evaporation of 1,8-cineole, lavender and other 11 EOs is a
 235 “non-activated zero-order” process. However, they simply derived this result by observing the
 236 closeness of vaporization enthalpies from Clausius–Clapeyron equation with the activation energy
 237 obtained by assuming a zero-order reaction [23, 24]. Our approach, though inadequate to find a
 238 reasonable model function for vaporization of the investigated samples, disagrees with the zero-order
 239 reaction assumption. Incidentally [24], these authors also claim that their method could furnish the
 240 vapor pressure curve for multicomponent systems, provided that the exact composition or the average
 241 molecular weight is known. In our opinion, this conclusion is very questionable since activity
 242 coefficients (unknown) may strongly affect the properties of the mixture.

243 The E_a versus α dependencies for vaporization of lavender (limited to the first step only), sage, 1,8-
 244 cineole and camphor calculated according to KAS are displayed in Fig. 6, where the relative
 245 uncertainties associated with the E α values (RUE, %), expressed as error bars, were estimated to be
 246 always lower than 8%. The E_a versus α curves show a decreasing trend, typical of the reversible
 247 processes [35], similarly to what previously observed for dehydration of different materials
 248 [39,40,41,42]. The results substantially confirm the stability order inferred on the basis of the T_{on} ,
 249 since the higher is the energy barrier (for vaporization of camphor), the slower is the vaporization
 250 rate, while 1,8-cineole and sage seem to have the highest (and comparable) reaction rates. On the
 251 other hand, these results match the thermochemical data related to phase transition for both camphor
 252 and 1,8-cineole [43]. In fact, the higher volatility of 1,8-cineole with respect to camphor could be
 253 expected on the basis of its lower boiling temperature (449.5 against 482.3 K) or its lower standard
 254 molar enthalpy of vaporization at 298.15 K (49.0 or 53.2 against 54.5 kJ mol⁻¹).

255 However, the thermal stability (or volatility) cannot be assessed only on the basis of the activation
 256 energy. Actually, it is commonly found that E_a obtained using a model-fitting method is not
 257 independent of pre-exponential factor, but the two quantities are strictly correlated through the
 258 compensation effect previously described in Eq. (6). To confirm this, the natural logarithm of A α is
 259 plotted versus the extent of conversion (Fig. 7) showing the same trends of Fig. 7.

260 Lastly, in order to have a clearer idea of the stability order, kinetic rate constants $k_{\text{vap}}(\langle T \rangle_{\alpha,\beta})$ were
261 calculated according to the well-known Arrhenius equation, at the temperatures $\langle T \rangle_{\alpha,\beta}$ [average
262 between $T(\alpha = 0.2)$ and $T(\alpha = 0.8)$ at a given heating rate β]. These values are summarized in Table
263 3 for vaporization of all four samples. The $k_{\text{vap}}(\langle T \rangle_{\alpha,\beta})$ values show that camphor seems to be the
264 most stable, thus confirming the results of thermal behavior (the highest T_{on}). Furthermore, lavender
265 seems to vaporize faster than any other sample, while no significant difference are worth noting
266 between the values referred to sage and 1,8-cineole, which can be considered to have comparable
267 stability within the errors associated with these results.

268

269

270 **Conclusions**

271 Thermal behavior and vaporization kinetics of EOs was scarcely investigated in the past, and only
272 few data are available in literature on this topic. In addition, kinetic methods applied in this study
273 require a priori assumptions not supported by experimental evidence and are now out of date. An
274 early model-fitting method was used within this work only as preliminary approach for comparison
275 purpose.

276 Our investigation involves the application of a modern kinetic approach to vaporization of essential
277 oils (from thermogravimetric data) based on an integral isoconversional method, which meets the
278 recommendations of the Kinetic Committee of ICTAC. In the case of lavender, where two-step
279 vaporization occurred, the first step was considered for the kinetic computations, and the results
280 obtained were in accordance with those of vaporization for the other samples. It was possible to verify
281 that the Arrhenius pair values (namely, activation energy and pre-exponential factor) do not vary
282 appreciably with the extent of conversion (at least in the range between 0.20 and 0.80). The kinetic
283 results based on the $k_{\text{vap}}(\langle T \rangle_{\alpha,\beta})$ values seem to confirm in some way the first preliminary observation
284 on the thermal behavior, thus revealing that camphor is the more stable among the samples examined.
285 The stabilities of the other samples based on the thermal behavior (on T_{on} reported in Fig. 4) seem to

286 be comparable, thus revealing that in this case the TG is not able to discriminate among vaporization
287 of substances with similar volatility. On the other hand, slight differences have been observed taking
288 into account the $k_{\text{vap}}(\langle T \rangle_{\alpha,\beta})$ values listed in Table 3. In particular, the following increasing order of
289 stability could be hypothesized for the three remaining samples: lavender < 1,8-cineole \leq sage. In any
290 case, lavender seems to vaporize faster than any other sample, while no significant difference is worth
291 noting between the values referred to sage and 1,8-cineole, which can be considered to have
292 comparable stability within the errors associated with these results.

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408

Caption of the figures

Fig. 1 Main compounds of lavender and sage essential oils extracted by microwaves, at extraction time of 30 min

Fig. 2 TG (**a**) and DTG (**b**) curves of evaporation of the EOs and components investigated under inert nitrogen atmosphere at 2 K min^{-1}

Fig. 3 TG (**a**) and $d\alpha/dT$ (**b**) versus T curves of lavender at 2 K min^{-1}

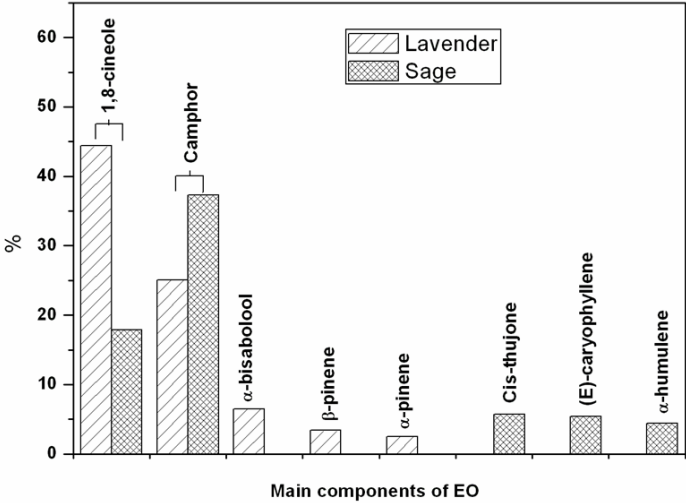
Fig. 4 Onset evaporation temperatures (T_{on}) extrapolated from the TG curves of the Eos and components investigated at corresponding heating rate β

Fig. 5 Experimental and reconstructed α vs. T curves for vaporization of lavender, sage, camphor and 1,8-cineole. In order to reconstruct the curves kinetic data taken from experiments at 2 K min^{-1} (plots **a**, **c**, **e** and **g**, respectively) and at 10 K min^{-1} (plots **b**, **d**, **f** and **h**, respectively) were used

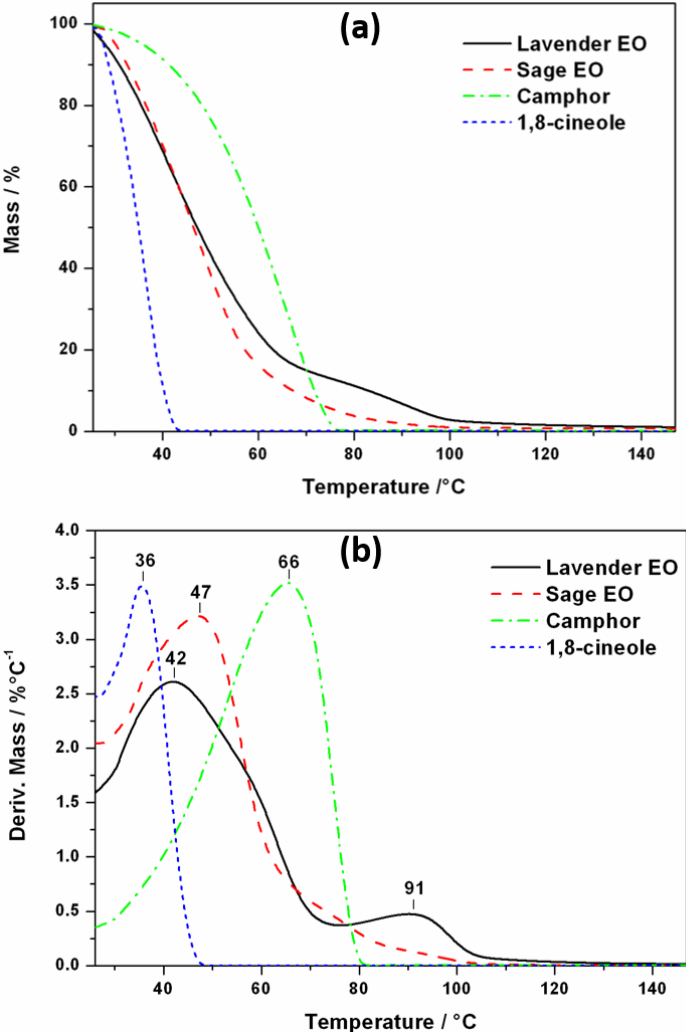
Fig. 6 Conversion dependency of activation energy of evaporation for the Eos and components investigated determined according to the integral isoconversional KAS method

Fig. 7 Conversion dependency of pre-exponential factor of evaporation for the EOs and components investigated

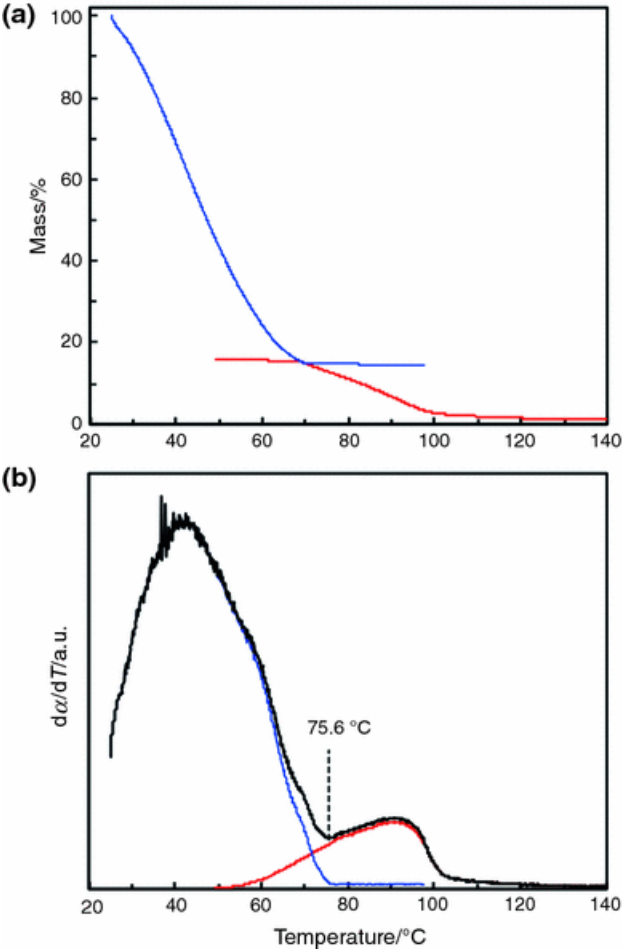
432 **Fig.1**



433 **Fig. 2**
434

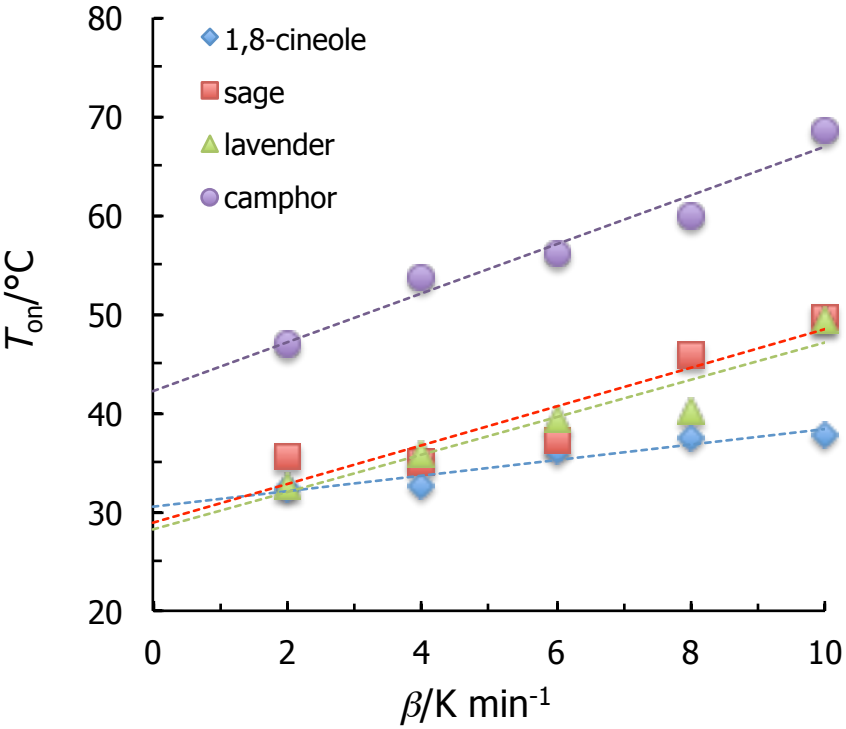


436 **Fig. 3**

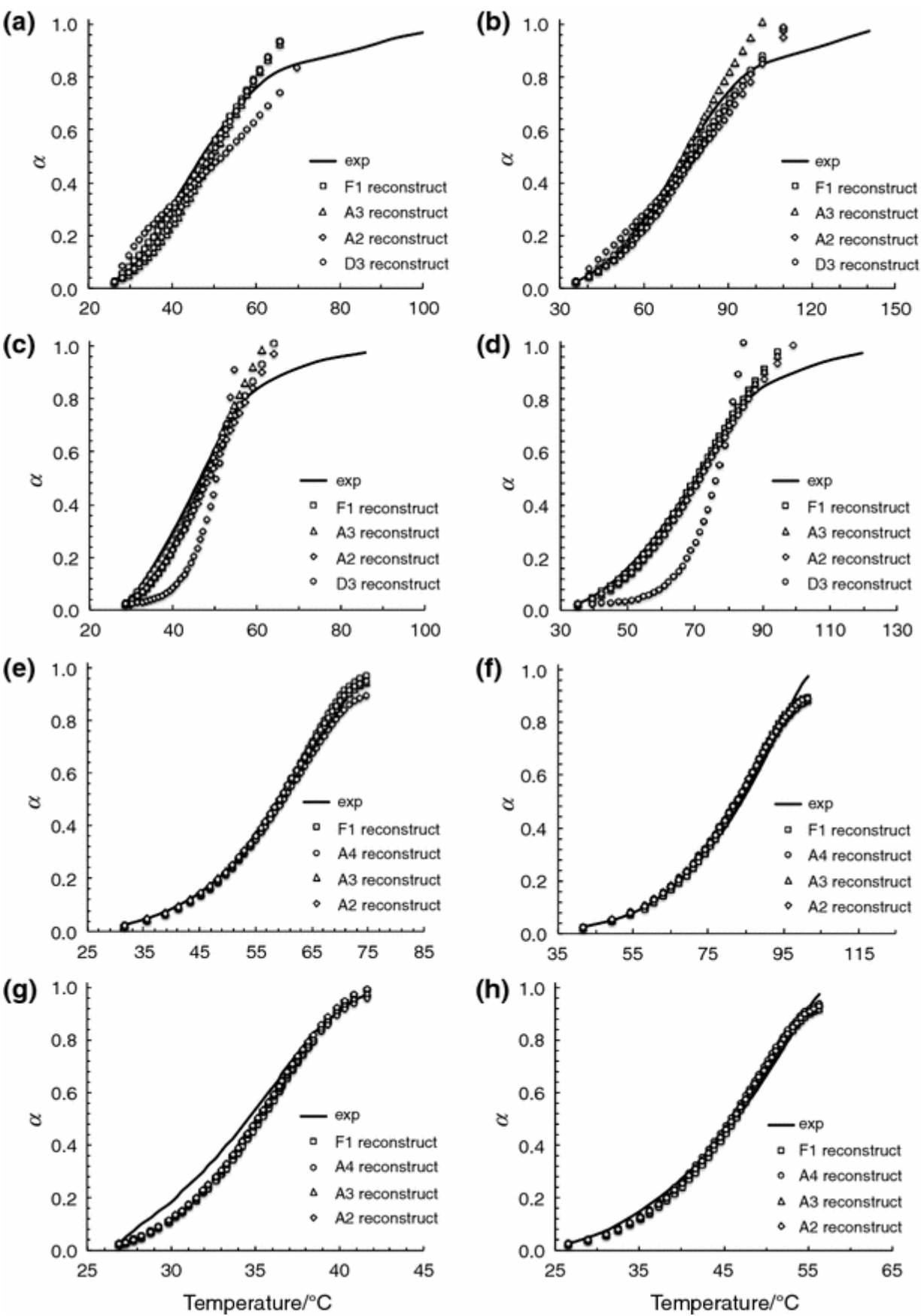


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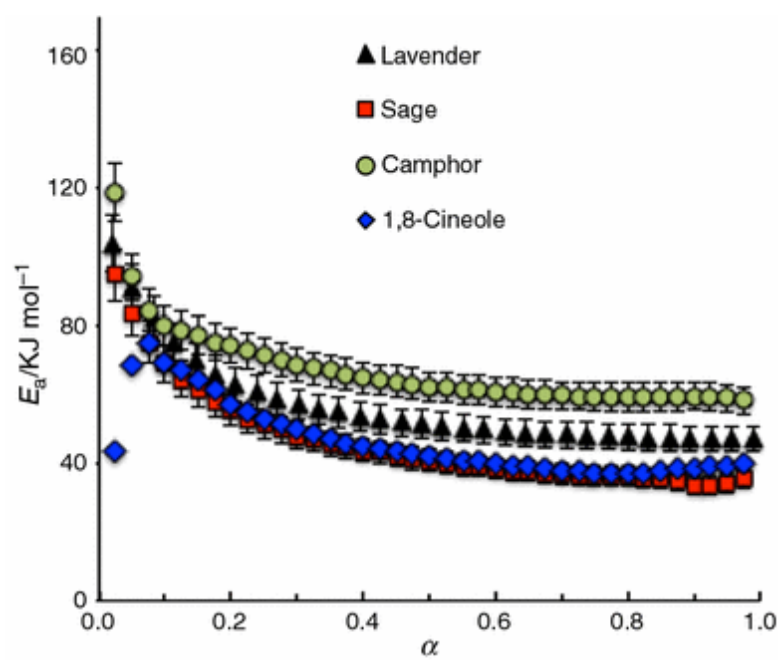
438 **Fig. 4**



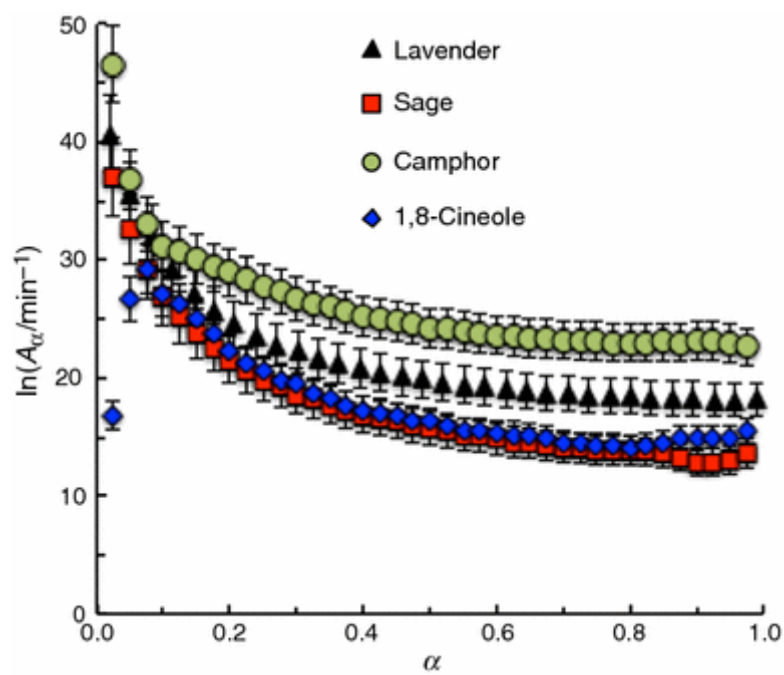
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442 Fig. 6



443
444 Fig. 7



445

446 **Table 1 Kinetic triplets for the most suitable mechanisms determined by the CR method according to Eq. (4) for vaporization of lavender,**
447 **sage, 1,8-cineole and camphor**

Parameter	From TG experiment/2 K min ⁻¹				From TG experiment/10 K min ⁻¹			
<i>Lavender</i>								
Best models ^a	F1	D3	A3	A2	F1	D3	A3	A2
<i>R</i> ^{2b}	0.8022	0.7640	0.6984	0.7640	0.8942	0.8635	0.7994	0.8566
<i>E</i> _a /kJ mol ⁻¹	47.2	86.7	12.0	20.8	40.8	76.6	9.7	17.5
<i>A</i> /min ⁻¹	5.2 × 10 ⁶	0.7	4.5	170	9.0 × 10 ⁴	1.9 × 10 ⁹	1.0	17
<i>Sage</i>								
Best models ^a	F1	D3	A3	A2	F1	D3	A3	A2
<i>R</i> ^{2b}	0.8161	0.7725	0.7609	0.7911	0.9043	0.8698	0.8548	0.8835
<i>E</i> _a /kJ mol ⁻¹	68.1	124	19.1	31.3	51.4	96	13.3	22.8
<i>A</i> /min ⁻¹	1.9 × 10 ¹⁰	9.8 × 10 ¹⁸	86	1.1 × 10 ⁴	5.9 × 10 ⁶	1.3 × 10 ¹³	4.0	1.5 × 10 ²
<i>1,8-Cineole</i>								
Best models ^a	F1	A4	A3	A2	F1	A4	A3	A2
<i>R</i> ^{2b}	0.8343	0.8256	0.8287	0.8316	0.9199	0.9196	0.9198	0.9198
<i>E</i> _a /kJ mol ⁻¹	250	49.9	68.3	105	114	24.5	34.4	54.2
<i>A</i> /min ⁻¹	1.2 × 10 ³⁶	3.6 × 10 ⁷	6.2 × 10 ¹⁰	1.5 × 10 ¹⁷	7.7 × 10 ¹⁷	6.4 × 10 ²	3.4 × 10 ⁴	8.5 × 10 ⁷
<i>Camphor</i>								
Best models ^a	F1	A4	A3	A2	F1	A4	A3	A2
<i>R</i> ^{2b}	0.9960	0.8371	0.8305	0.8388	0.9917	0.9319	0.9282	0.9250
<i>E</i> _a /kJ mol ⁻¹	88	17.9	25.7	41.2	70.9	13.4	19.8	32.6
<i>A</i> /min ⁻¹	9.5 × 10 ¹²	28	6.4 × 10 ²	2.6 × 10 ⁵	2.6 × 10 ⁹	2.9	33	3.6 × 10 ³

448 Relative standard deviations (RSD) associated with E_a and A values were always lower than 6%

449 ^aSelected on the basis of the best linear fit (among 13 candidate models) obtained by applying a linear regression procedure to the $\ln[g(\alpha)/T^2]$
450 versus $1/T$ data according to Eq. (4)

451 ^bValue referred to the best four linear fits obtained by plotting $\ln[g(\alpha)/T^2]$ versus $1/T$ according to Eq. (4)

452

453 **Table 2 Regression parameter values related to Eq. (6) at each TG experiment at a given value of β**

Parameter	Data obtained at a given value of β					Averages, $\langle a \rangle$ and $\langle -b \rangle$
	2 K min ⁻¹	4 K min ⁻¹	6 K min ⁻¹	8 K min ⁻¹	10 K min ⁻¹	
a	0.396	0.395	0.397	0.395	0.398	0.396
$-b$	3.5	3.4	3.4	3.5	3.5	3.5
SD(a) ^a	0.006	0.009	0.007	0.010	0.012	
SD(b) ^b	0.5	0.6	0.5	0.6	0.6	
SD($y:x$) ^c	1.3032	1.3306	1.3131	1.3997	1.4347	
R^2	0.9964	0.9910	0.9955	0.9887	0.9859	
F obs. value	3832	2845	3451	2556	2015	

454 ^aStandard deviation of the slope

455 ^bStandard deviation of the intercept

456 ^cStandard deviation of the estimate of $y(\ln A_i)$ by the regression line through Eq. (6)