

Lysozyme activity in donkey milk

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ABSTRACT

The aim of this work was to expand the knowledge on the lysozyme activity of donkey milk in relation to the lactation stage, age of the animals and pasteurization treatment. Raw and pasteurized bulk milk samples were collected monthly for one year and were analysed in relation to lysozyme activity. Individual raw milk samples were collected from 12 jennies, at three lactation stages: three, six and nine months. The results showed that the lactation stage and the age of the animals influenced the lysozyme activity of donkey milk. The highest lysozyme activity values were found at early lactation and in animals older than 15 years. We confirm that low-temperature long-time pasteurization of donkey milk does not reduce the activity of this enzyme. In addition, calcium, ash and some fatty acids, and in particular the saturated fatty acids, are linked to a higher lysozyme activity of donkey milk at early lactation.

1. Introduction

Lysozyme (LZ) is a glycoside hydrolase, which has shown antimicrobial activity against a

27 great variety of bacteria and is able to inhibit viruses, parasites and fungi (Benkerroum, 2008). This
28 powerful enzyme is part of the innate immune system and is present in many secretions (tears, saliva,
29 urine, mucus and milk) and is also produced by macrophages, neutrophils and dendritic cells (Ragland
30 & Criss, 2017). Regarding the LZ content in milk, LZ concentrations of between 0.3 and 1.1 g L⁻¹
31 have been reported in humans, while in the milk of ruminants, LZ is only present in trace amounts
32 (Altomonte, Salari, Licitra, & Martini, 2019; Benkerroum, 2008). Surprisingly, LZ represents one of
33 the main whey protein fractions in donkey milk and its presence is one of the key components of
34 interest in asinine milk (Martini, Altomonte, Licitra, & Salari, 2018). Among different donkey breeds,
35 the LZ content ranges from 1 to 3.7 g L⁻¹. In particular, the LZ content has been reported to be about
36 1 g L⁻¹ in Ragusano (Vincenzetti et al., 2008), 1.5 g L⁻¹ in Amiata (Caroli et al., 2015), 2.2 g L⁻¹ in
37 Martina Franca and in Jiangyue (Guo et al., 2007; Vincenzetti et al., 2011) and 1-3 g L⁻¹ in the Serbian
38 Balkan breed (Gubić et al., 2016). Analysis of the LZ content of milk obtained by four cross-breed
39 jennies has shown mean values of 3.7 g L⁻¹ (Chiavari, Coloretti, Nanni, Sorrentino, & Grazia, 2005).
40 In donkey milk, the LZ content seems to decrease during lactation (Gubić et al., 2016; Polidori &
41 Vincenzetti, 2010).

42 LZ activity is linked to its ability to catalyse the hydrolysis of the 1,4-β-linkages between N-
43 acetylmuramic acid and N-acetylglucosamine in peptidoglycan, which is the major component of the
44 gram-positive bacterial cell wall, causing bacterial lysis (Alhazmi, Stevenson, Amartey, & Qin,
45 2014). By virtue of its activity, LZ contributes to the low bacterial count of donkey milk and seems
46 to exert antimicrobial functions synergistically with other milk components, such as lactoferrin,
47 lactoperoxidase, N-acetyl-β-D glucoaminidase, immunoglobulins and some fatty acids (FAs)
48 (Brumini, Criscione, Bordonaro, Vegarud, & Marletta, 2016; Nazzaro, Orlando, Fratianni, &
49 Coppola, 2010). In vitro, LZ has shown resistance to acid pH and human gastrointestinal enzymes
50 (Tidona et al., 2014) and anti-inflammatory and anti-tumor actions (Lee, Ku, Na, & Bae, 2015; Mao
51 et al., 2009). On animal models, LZ has also been found to limit bacterial infections and to promote
52 the growth of probiotic bacteria associated with gut health (Huang et al. 2018).

53 An interesting aspect of LZ antibacterial activity is its use in the food industry. Traditionally
54 LZ from hen egg white has been widely used as a natural preservative in order to extend the shelf life
55 of food products (Silvetti, Morandi, Hintersteiner, & Brasca, 2017). However recently, donkey milk
56 has been tested as an antimicrobial additive in dairy products and has shown excellent results in
57 preventing cheese-blowing during ripening as an alternative to hen egg white LZ, which may cause
58 allergic reactions in egg allergic patients (Cosentino, Paolino, Freschi, & Calluso, 2013; Niro et al.,
59 2017). Understanding the potential application of donkey milk LZ in the food industry entails
60 knowledge of which factors affect its variability and its resistance to technological processes. Addo
61 & Ferragut (2015) reported the resistance of LZ activity in donkey milk to moderate heat treatments
62 and storage, however only a few studies have been carried out on the effects of thermal treatments.
63 In addition, LZ activity in donkey milk is less studied compared to other sources such as other
64 mammalian milk or hen egg white, and factors that affect its activity have been poorly investigated.
65 The aim of this work was to expand the knowledge on the activity of LZ in donkey milk in relation
66 to the lactation stage, age of the animals and pasteurization treatment. A possible correlation between
67 LZ activity and the chemical composition of donkey milk was also evaluated.

68

69 **2. Material and methods**

70 *2.1 Animals and samples collection*

71 The research was conducted on a herd rearing a total of about 160 Amiata donkey, an Italian
72 breed native to Mount Amiata, in Tuscany (central Italy). Animals were reared in a semi-intensive
73 system and were fed with mixed hay ad libitum and about 2.5 kg per day per head of commercial
74 pelleted concentrate formulated for dairy jennies. The farm produced pasteurized milk for human
75 consumption in accordance with the requirements of Regulation (EC) No 853/2004. During the first
76 month of lactation, all the milk was left for the foal. One month after delivery, the jennies were
77 routinely machine-milked once per day and four hours before being milked, the foals were separated
78 from the jennies. Bulk milk samples from 40 jennies (of which 8 primiparous, 7 secondiparous and

79 25 multiparous; aged on average 9.5 years) were collected once a month for one year and at each
80 sampling, two raw and two pasteurized samples (low-temperature long-time: 65 °C for 30 min) were
81 analysed in terms of LZ activity. In addition, individual raw milk samples were collected from 12
82 healthy jennies, at three lactation stages: three, six and nine months. The animals were homogeneous
83 in terms of foaling date and were aged between 9 and 20 years. All the samples were immediately
84 refrigerated at 4 °C and then analysed within 24 hours.

85

86 *2.2 Milk analysis*

87 Individual milk samples were analysed for dry matter, fat and lactose by infrared analysis
88 (MilkoScan; Italian Foss Electric, Padova, Italy) and for total protein, casein, ash and calcium content,
89 using methods of the Association of Official Analytical Chemists (AOAC, 1990). Milk fat extraction
90 was performed following Rose-Gottlieb's method (AOAC, 2000) and methyl esters of fatty acids
91 were prepared according to Christie (1982). A Perkin Elmer Clarus 480 (Perkin Elmer, Norwalk, CT,
92 USA) equipped with a flame ionization detector and a capillary column (ThermoScientific TR-FAME
93 60 m × 0.25 mm ID; film thickness 0.25 µm, Fisher Scientific UK) were used. The peak areas of
94 individual FAs were identified using a FAs standard injection (Food Industry FAME Mix – Restek
95 Corporation, 110 Benner Circle, Bellefonte, PA16823) and quantified as the percentage of total FAs.

96

97 *2.3 LZ activity analysis*

98 The LZ activity was evaluated in both bulk and individual milk samples using a commercial
99 fluorimetric method on a microplate (EnzChek Lysozyme Assay Kit, Thermo Fisher Scientific,
100 Waltham, MA, USA). The test uses a suspension of *Micrococcus lysodeikticus* labelled with
101 fluorescein. This microorganism is sensitive to the lytic activity of LZ which leads to a variation in
102 the intensity of the fluorescence measured at ~485/530 nm (excitation/emission). Milk was diluted
103 and no defatting methods were used. The results were compared with a LZ standard curve and
104 expressed in U ml⁻¹.

105

106 2.4 Statistical analysis

107 The results of the LZ activity on individual samples were analysed using ANOVA for repeated
108 measurements, considering the three sampling times (three, six and nine months of lactation) and the
109 age of the jennies (< 9 years, between 9 to 15 years and > 15 years) as fixed effects; mean and standard
110 deviation of milk composition were also calculated. The LZ activity of bulk milk samples was
111 analysed using ANOVA considering the milk heat treatment as the fixed effect (raw or pasteurized
112 milk). Least significance means were compared by the t-test. Pearson's correlations were also carried
113 out for each stage of lactation. The significance level was set at $P < 0.05$. Statistical analysis was
114 carried out using JMP software (SAS Institute, 2002).

115

116 3. Results

117 Table 1 reports the average chemical composition of individual donkey milk samples
118 evaluated throughout the entire lactation. Despite being low in lipids, donkey milk provides a good
119 contribution of unsaturated fatty acids (over 46%) and Omega 3. Table 2 shows the individual LZ
120 activity during lactation and between the different age classes of animals. The lactation stage
121 influenced the LZ activity of donkey milk. A statistically higher value was found at three months of
122 lactation and a lower value at six months, while an increasing trend was observed at nine months. LZ
123 activity was also influenced by the age of the animals, and higher values were found as the age of
124 animals increased. In particular, a significantly higher LZ activity was found in donkeys older than
125 15 years compared to those younger than 9 years. With regard to the pasteurization treatment, no
126 significant differences were found in the LZ activity between the raw and the pasteurized milk
127 (1340.82 ± 371.078 and 1402.50 ± 294.722 U mL⁻¹ respectively, with a SEM equal to 154.295).
128 Significant correlations between LZ activity and the chemical composition of donkey milk were
129 found during lactation: the most numerous and significant correlations were found at the third month
130 of lactation (Table 3).

131 In the first stage of lactation, significant correlations were found between LZ activity and both
132 the calcium and ash contents. In addition, the results of the correlation analysis between LZ activity
133 and FAs showed that the content of some saturated fatty acids (SFAs) (C6:0, C10:0, C12:0, C14:0
134 and C16:0), total SFAs, short chain fatty acids (SCFAs) and medium chain fatty acids (MCFAs) was
135 correlated positively with the LZ activity. In addition, another two unsaturated fatty acids
136 C18:2n6(t9,t12) and C18:3n3(c9,c12,c15) showed positive correlations with LZ activity. Differently,
137 the content of the following unsaturated fatty acids: C18:1(t9), C18:2n6(c9,c12), C20:1 and C20:4n6,
138 total polyunsaturated fatty acids (PUFAs) and long chain fatty acids (LCFAs) was correlated
139 negatively with LZ activity. Finally, LZ activity was correlated positively with the total n-3 and
140 negatively with the total n-6 fatty acids.

141

142 **4. Discussion**

143 The results on the gross composition are in agreement with our previous studies on Amiata
144 donkey (Martini, Altomonte, Salari, & Caroli, 2014; Ragona et al., 2016). As shown in Table 1, the
145 donkey milk was characterized by a low total protein content and in particular by a low casein content.
146 These characteristics contribute to the hypoallergenic properties of donkey milk which make it a valid
147 alternative for people with an allergy to cow milk proteins (Martini et al., 2018). The high content of
148 lactose, in addition to making it more palatable, contributes to the development of intestinal flora
149 (Venema, 2012).

150 Our data on the LZ activity in Amiata milk were slightly lower than those reported in previous
151 studies on individual milk samples from the same breed (Pilla, Daprà, Zecconi, & Piccinini, 2010;
152 Ragona et al., 2016) and similar compared to the results reported for bulk milk of the Ragusano
153 donkey (Conte, Foti, Malvisi, Giacobello, & Piccinini, 2012). Higher values (11531 U ml^{-1}) have
154 been reported by Addo & Ferragut (2015) in a pool of milk from donkeys between the first and third
155 month of lactation. The apparent variability of results found in the literature is due to differences in
156 sampling methods and analyses. In our study, the trend of LZ activity during lactation is similar to

157 that reported by Pilla et al. (2010). Regarding the age of the animals, Qureshi & Enbergs (2012)
158 reported that donkeys older than 10 years exhibited significantly higher LZ activity values compared
159 to donkeys younger than 10 years, in agreement with our findings. Similar results were also found in
160 a study carried out on horses, in which 10-14 year-old mares showed significantly higher LZ activity
161 values compared to 5-9 year-old mares (Sarwar, Enbergs, & Klug, 2001). Comparing LZ activity in
162 raw and pasteurized milks, our results showed that pasteurization treatment did not affect the LZ
163 activity. Similarly, other studies applying similar temperature treatments (Coppola et al., 2002;
164 Chiavari et al., 2005) or higher temperatures and shorter times (70-90 °C for 1 min) did not find
165 changes in the LZ activity of donkey milk (Coppola et al., 2002; Addo & Ferragut, 2015).

166 The positive correlation between LZ activity and calcium content found in this study was in
167 agreement with the findings of Sarwar et al. (2001) in mare milk. These results may be related to the
168 ability of equid milk LZ to bind calcium, which seems to stabilize the LZ structure and increase the
169 antimicrobial activity (Brumini et al., 2016). The strong positive correlations found between LZ
170 activity and SFAs and FAs with less than 17 carbon atoms strengthen the hypothesis of Brumini et
171 al. (2016) regarding the synergistic activity of LZ with some FAs known to be antibacterial such as
172 C12:0. In addition, we found other positive correlations with LZ activity and C10:0, C14:0 and C16:0
173 FAs, the latter having already been reported to be bactericidal FAs (Sprong, Hulstein, & Van der
174 Meer, 2001; Desbois & Smith, 2010). This relationship could strengthen the milk antibacterial
175 activity when the immature immune system of the offspring still needs protective factors. Further
176 confirmation of the synergistic action between LZ and milk FAs is provided by a study on egg white
177 LZ which showed that the lipophilization of LZ with SCFAs improves the bactericidal action against
178 gram-negative bacteria without decreasing its effect on gram-positive bacteria (Liu, Sugimoto,
179 Azakami, & Kato, 2000). Further studies are needed to clarify the negative relationships we observed
180 between lysozyme and PUFAs and LCFAs.

181 **5. Conclusions**

182 In Amiata donkey milk, LZ activity was found to be influenced by the lactation stage and the

183 age of the animals. In particular, the highest LZ activity values were found at early lactation and in
184 animals older than 15 years. This study confirms that the low-temperature long-time pasteurization
185 treatment of donkey milk does not impact on the variability in the activity of this enzyme. In addition,
186 both calcium, ash and some fatty acid contents, and in particular the SFAs, are linked to a higher
187 activity of donkey milk LZ at early lactation.

188

189 **Funding**

190 This work was supported by PRA 2017 (Ateneo Research Project, University of Pisa)

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288

289 Table 1. Gross composition and fatty acids classes of individual donkey milk. Data are expressed as
 290 mean value and standard deviation.

	Mean	Standard deviation
Gross composition (g 100mL⁻¹)		
Dry matter	9.80	1.095
Total Protein	1.65	0.201
Casein	0.81	0.099
Fat	0.33	0.193
Lactose	6.97	0.194
Ash	0.33	0.055
Calcium	0.09	0.035
Fatty acids classes (g 100 g of fat⁻¹)		
SFAs ¹	53.16	7.650
MUFAs ²	27.07	6.767
PUFAs ³	19.77	2.766
SCFAs ⁴ (\leq C10)	12.90	3.221
MCFAs ⁵ (C11-C17)	42.45	4.594
LCFAs ⁶ (\geq C18)	44.65	6.789
Total n-3	5.16	1.453
Total n-6	14.54	3.514
Lysozyme activity (U mL⁻¹)	1670.11	411.454

291 ¹SFAs: saturated fatty acids; ²MUFAs: monounsaturated fatty acids; ³PUFAs: polyunsaturated fatty
 292 acids; ⁴SCFAs: short chain fatty acids; ⁵MCFAs: medium chain fatty acids; ⁶LCFAs: long chain fatty
 293 acids.

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295 Table 2. Average lysozyme (LZ) activity in relation to lactation stage and age of the animals.

Month of lactation	3	6	9	SEM
LZ activity (U mL ⁻¹)	1842.21a	1433.84b	1733.46ab	446.058
Age of the animals	<9	9-15	>15	SEM
LZ activity (U mL ⁻¹)	1547.83b	1610.57ab	1824.11a	446.058

296 Within a row, means without a common lowercase letter differ at P < 0.05.

297

298 Table 3. Correlations between milk composition and lysozyme activity at three months of lactation.

Component	Lysozyme activity (U mL ⁻¹)
Ash	0.684*
Calcium	0.613*
C6:0	0.813**
C10:0	0.666*
C12:0	0.816**
C14:0	0.789**
C16:0	0.793**
C18:1(t9)	0.726*
C18:2n6(t9,12)	0.828**
C18:2n6(c9,12)	0.884***
C18:3n3(c9,12,15)	0.732*
C20:1	-0.696*
C20:4n6	-0.787**
SFAs ¹	0.868**
PUFAs ²	-0.727*
SCFAs ³ (≤C10)	0.677*
MCFAs ⁴ (≥C11≤C17)	0.857**
LCFAs ⁵ (≥C18)	-0.877***
Total n-3	0.738*
Total n-6	-0.884***

299 ¹SFAs: saturated fatty acids; ²PUFAs: polyunsaturated fatty acids; ³SCFAs: short chain fatty acids;

300 ⁴MCFAs: medium chain fatty acids; ⁵LCFAs: long chain fatty acids.

301 Only significant correlations are shown

302 Significance levels: *P < 0.05; **P < 0.01, ***P < 0.001

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