# RESEARCH

# Vitamin D measurement and effect on outcome in a cohort of patients with heart failure

Federica Saponaro<sup>1,2</sup>, Alessandro Saba<sup>1,3</sup>, Sabina Frascarelli<sup>1</sup>, Concetta Prontera<sup>4</sup>, Aldo Clerico<sup>4</sup>, Marco Scalese<sup>5</sup>, Maria Rita Sessa<sup>6</sup>, Filomena Cetani<sup>2</sup>, Simona Borsari<sup>2</sup>, Elena Pardi<sup>2</sup>, Antonella Marvelli<sup>7</sup>, Claudio Marcocci<sup>2</sup>, Claudio Passino<sup>4</sup> and Riccardo Zucchi<sup>1</sup>

Correspondence should be addressed to F Saponaro: federica.saponaro@gmail.com

## **Abstract**

Objectives: The aims of this paper were to evaluate the levels of Vitamin D (VitD) in patients with heart failure (HF), compared to a control group, to assess the effects of VitD on HF outcome and to compare VitD measurement between LIAISON immunoassay and HPLC-MS-MS methods in this population.

Design and Methods: We collected clinical, biochemical and outcome data from 247 patients with HF and in a subgroup of 151 patients, we measured VitD both with LIAISON and HPLC-MS-MS.

Results: HF patients had statistically lower 25OHD levels  $(45.2\pm23.7\,\mathrm{nmol/L}\,\mathrm{vs}\ 58.2\pm24.0\,\mathrm{nmol/L}$ , P<0.001) and a statistically higher prevalence of VitD insufficiency  $(61.1\%\,\mathrm{vs}\ 39.5\%,\,P<0.001)$  and deficiency  $(24.7\%\,\mathrm{vs}\ 6.6\%,\,P<0.001)$ , compared to healthy controls. There was a significant inverse relationship between baseline 25OHD and risk of HF-related death, with a HR of 0.59  $(95\%\,\mathrm{Cl}\ 0.37-0.92,\,P=0.02)$ , confirmed in a multivariate adjusted analysis. Kaplan–Meier survival analyses showed that VitD insufficiency was associated with reduced survival in HF patients (log rank P=0.017). There was a good agreement between LIAISON and HPLC-MS-MS (Cohen's kappa coefficient 0.70), but the prevalence of VitD insufficiency was significantly higher with the former compared to the latter method  $(58.3\%,\,n=88\,\mathrm{vs}\ 55.6\%,\,n=84,\,P<0.001)$ . LIAISON underestimated the 25OHD levels and showed a mean relative bias of  $-0.739\%\,\mathrm{with}\ 95\%$  of limits of agreement  $(-9.00\,\mathrm{to}\ +7.52\%)$ , when compared to HPLC-MS-MS. Conclusions: 25OHD levels adequately measured by HPLC-MS-MS showed to be low in HF population and to be correlated with HF-related risk of death.

#### **Key Words**

- vitamin D
- 25-hydroxyvitamin D
- mass spectrometry coupled to high performances liquid chromatography
- ▶ immunoassay
- ▶ hypovitaminosis D

Endocrine Connections (2018) 7, 957–964

## Introduction

Heart failure (HF) is a complex and chronic condition and despite advances in therapeutic options, its prognosis remains poor, with high mortality and morbidity. Vitamin D (VitD) has been recently emerged as a factor for HF risk and outcome evaluation (1, 2).

CC

VitD is a fat-soluble vitamin, which functions as a steroid hormone; it is involved in bone and skeletal homeostasis, and it was found to have many others extraskeletal functions, even on cardiovascular system (3, 4, 5, 6). Indeed, hypovitaminosis D is associated with

http://www.endocrineconnections.org https://doi.org/10.1530/EC-18-0207 © 2018 The authors Published by Bioscientifica Ltd This work is licensed under a Creative Commons Attribution 4.0 International License.

<sup>&</sup>lt;sup>1</sup>Department of Surgical, Medical, Molecular and Critical Area Pathology, Laboratory of Biochemistry, University of Pisa, Pisa, Italy

<sup>&</sup>lt;sup>2</sup>Endocrinology Unit 2, University of Pisa, Pisa, Italy

<sup>&</sup>lt;sup>3</sup>Laboratory of Clinical Pathology, University Hospital of Pisa, Pisa, Italy

<sup>&</sup>lt;sup>4</sup>Fondazione Toscana Gabriele Monasterio, Pisa, Italy

<sup>&</sup>lt;sup>5</sup>Institute of Clinical Physiology, National Council of Research, Pisa, Italy

<sup>&</sup>lt;sup>6</sup>Laboratory of Endocrinology, University Hospital of Pisa, Pisa, Italy

<sup>&</sup>lt;sup>7</sup>Department of Translational Research and of New Surgical and Medical Technologies, University of Pisa, Pisa, Italy

958



an increased risk of hypertension, coronary artery disease, ischemic heart disease and stroke. Moreover, it is known that it is involved in the pathophysiology of HF, with different mechanisms (downregulation of the reninangiotensin system, enhancement of insulin secretion and sensitivity, protection against angiogenesis and modulation of inflammatory processes) and low levels of VitD are associated with worse prognosis (7, 8, 9, 10, 11). A limitation in comparing the large amount of literature data on VitD and HF is the variety in VitD status assessment. A near to ideal analytical methods of VitD measurement is strongly required, which could be selective and sensitive, performed using a universal and standardized method and considering the different VitD metabolites (more than 50 VitD physiological products have been described) (5). Traditionally, radioimmunoassay and chromatography have been used for VitD measurement and the majority of routine analyses are performed all over the world by immunoassay techniques, which allow nonchromatographic quantification of total 25 Hydroxyvitamin D (25OHD). In recent years, liquid chromatographytandem mass spectrometry (LC-MS-MS) has been arisen as the 'gold standard' method due to its high sensitivity and accuracy and the possibility to measure multiple VitD compounds at the same time (12, 13).

The aims of this paper are (i) the evaluation of the prevalence of hypovitaminosis by measuring 25OHD levels with HPLC-MS-MS in a cohort of patients with HF compared with a control group of healthy subjects, (ii) the assessment of the effect of VitD on HF outcome, (iii) the comparison of 25OHD measurement between LIAISON immunoassay and HPLC-MS-MS methods in a population of HF patients.

# **Materials and methods**

## **Patients**

Between January 2012 and October 2014, we retrieved information about 247 patients with systolic dysfunction (left ventricular ejection fraction, LVEF <50%), referred to the Division of Cardiovascular Medicine of the Fondazione G. Monasterio in Pisa for HF management.

The study was approved by the Local Ethical Committee of Pisa. Consent has been obtained from each patient and healthy subject, after full explanation of the purpose and nature of all procedures used.

All patients fulfilled the European Society of Cardiology (ESC) criteria for the diagnosis of HF and ethical approval was obtained. Each patient underwent comprehensive characterization and the following data were available from basal evaluation: age, gender, BMI, NYHA class, blood pressure, ejection fraction, PTH levels and albumin-adjusted serum calcium.

In all patients, 25OHD levels were measured by highperformance liquid chromatography (HPLC-MS-MS). 25OHD levels were also measure by LIAISON assay in 151 patients, in whom an aliquot of the same sample used for HPLC-MS-MS analyses was available and had been stored at -20 $^{\circ}$ C.

To retrieve data about patient's follow-up, a telephone survey was conducted: all patients were recalled and data about hospitalizations for HF, major cardiovascular events, worsening of failure and exitus were collected.

#### **Assays**

# Mass spectrometry coupled to high-performance liquid chromatography (HPLC-MS-MS)

We measured 25OHD levels by HPLC-MS-MS in the whole group (n=245) of HF patients (blood samples – plasma – collected at the baseline evaluation and stored at -20°C). Moreover, we measured 25OHD levels by HPLC-MS-MS in a group of healthy people (n=76) from blood donors of Cisanello Hospital, Pisa.

To validate our HPLC-MS-MS method, we bought from DEQAS (the Vitamin D External Quality Assessment Scheme) a set of samples and compared 'true' DEQAS 25OHD values with our measurement. To achieve a good performance, laboratories are required to have 75% of assessable results within ±25% of the true values from DEQAS.

The accurate measurement of 25 hydroxyvitamin D3 (25OHD3) status was carried out by isotope dilution mass spectrometry coupled to high-performance liquid chromatography (HPLC-MS-MS), by using the MSMS VitD Kit from PerkinElmer. It is well known that HPLC-MS-MS offers a good quantification accuracy and the contribution of interfering compounds to the final results is limited (14).

Chemicals: solvents for sample preparation and HPLC elution, such as acetonitrile (LC-MS grade), methanol (LC-MS grade), ultra-pure water (LC-MS grade) and formic acid (for mass spectrometry ≈ 98%), were purchased from Sigma-Aldrich.

Instrumentation: the instrument consisted in an Agilent 1290 Infinity UHPLC system, including autosampler, binary pump and column oven, coupled to an AB-Sciex API 4000 triple quadrupole mass spectrometer (Concord, ON, Canada), equipped with an APCI source.



This work is licensed under a Creative Commons Attribution 4.0 International License

http://www.endocrineconnections.org https://doi.org/10.1530/EC-18-0207

© 2018 The authors Published by Bioscientifica Ltd



Chromatography was performed by a PerkinElmer Brownlee Supra C18  $3\,\mu\text{m}$ ,  $50\times2.1\,\text{mm}$  HPLC column, protected by a PerkinElmer Brownlee Supra C18 Guard Column.

HPLC-MS-MS conditions: the separation was carried out under gradient conditions by using methanol as solvent A, and water as solvent B, both containing 0.1% formic acid. The MS method, based on positive mode multiple reaction monitoring (MRM), made use of the following quantifying transitions: 25OHD<sub>3</sub>, 401.4→159.1; <sup>2</sup>H<sub>3</sub>-25OHD<sub>3</sub> (<sup>2</sup>H<sub>3</sub>-25OHD<sub>3</sub>, IS),  $404.4 \rightarrow 162.1$ ;  $401.4 \rightarrow 159.1$ ;  ${}^{2}H_{6}-25OHD_{3}$  ( ${}^{2}H_{6}-25OHD_{3}$ ). Remarkable is the use of <sup>2</sup>H<sub>6</sub>-25OHD<sub>3</sub>, a surrogate analyte which simulates 25OHD<sub>3</sub> in the real matrices containing this latter compound, used in the preparation of either calibration standard solutions or quality controls (QCs). Calibration curve and QCs: solutions containing <sup>2</sup>H<sub>6</sub>-25OHD<sub>3</sub> at six different concentration levels (4.7, 9.4, 18.8, 37.5, 75, 150 ng/mL) were used as standard solutions to build the calibration curve. They were obtained by reconstituting with water the lyophilized calibrators provided with the kit. Thereafter, a  $100\,\mu\text{L}$  aliquot of the resulting solution was added to 200 µL of acetonitrile containing 0.1% formic acid and a suitable amount of IS (daily precipitation solution, DPS). QCs were prepared as described for the calibration solutions, starting from the suitable lyophilized matters provided with the kit. In these solutions <sup>2</sup>H<sub>6</sub>-25OHD<sub>3</sub> was at three different concentration levels (9.4, 37.5, 75 ng/mL).

Sample preparation: plasma samples were stored at  $-20^{\circ}$ C. A  $100\,\mu$ L aliquot of thawed sample was added with DPS, as for calibrators and QC.

The intra-assay CV was specified by the producer and was <4.6% while the inter-assay CV was <4.7%

# Liaison 250HD total assay

In a subgroup of HF patients (n=151), from whom we had a second aliquot stored at  $-20^{\circ}$ C of the same plasma sample used for HPLC-MS-MS measurement, we quantified 25OHD with LIAISON 25OHD assay, a direct competitive chemiluminescence immunoassay (CLIA), which provides a quantitative determination. VitD binds to its binding protein and during the first phase, (first incubation), it dissociates from it by buffer 10% ethanol and surfactant and binds to the specific antibody on the solid phase. A tracer (VitD linked to an isoluminol derivative) is then added after 10 min. Unbound materials are washed out after incubation. A chemiluminescent reaction starts, when starter reagents are added. A photomultiplier

measures light signal as relative lights unit (RLU), which is inversely correlated to the 25OHD concentration.

Assay specificity has been estimated according to CLSI EP 7-A2 guidelines (15). The LIAISON 25OHD assay has an analytical sensitivity of 4 ng/mL. The intra-assay CV was specified by the producer and was 2–4%. The inter-assay CV was 7%. In our laboratory, the imprecision of Liaison 25OHD total assay (DiaSorin) evaluated with 12 control samples throughout 12 months (25OHD concentration from about 19 to 70 ng/mL) ranged from 4.6 to 10.2%.

#### Statistical analyses

Categorical variables are expressed as number of cases and percentages. Continuous variables are expressed as mean values and standard deviations or median values and interquartile range (IQR - 25° and 75° quartiles) as appropriate. Difference between groups was tested by chi-square test or paired t-test as appropriate. Agreement between LIAISON and HPLC-MS-MS method was measured by concordance correlation coefficient and by graphical display of the data and the reduced major axis of the data (the reduced major axis or SD line goes through the intersection of the means and has slope given by the sign of Pearson's r and the ratio of the standard deviations. The SD line serves as a summary of the center of the data). Bland-Altman plot was used to identify mean bias (the average of the differences between measurements obtained from the two compared assays) and 95% limits of agreement between methods. Cohen's kappa coefficient (k) was used to assess inter-methods agreement for qualitative (categorical) items. To assess the diagnostic performance of 25OHD in cardiac death, the receiver-operating characteristic (ROC) curve analysis was conducted and the area under the curve (AUC), sensitivity, specificity and the best cut off of 25OHD using the highest Youden index were calculated. Cox regression analysis was conducted to examine the hazard ratio (HR) and 95% CI for the association between death and baseline 25OHD. P values <0.05 were considered statistically significant. The statistical package Stata (13.0) was used for analysis.

# **Results**

# **HF** cohort

The study group included 247 patients, 203 (82.2%) males and 44 (18.8%) females, with a mean age of  $65\pm13$  years. All patients had stable HF disease, mostly in the NYHA I and II class (80.2%, n=198) with mean EF of  $33\pm8\%$ .



This work is licensed under a Creative Commons Attribution 4.0 International License.

http://www.endocrineconnections.org https://doi.org/10.1530/EC-18-0207 © 2018 The authors Published by Bioscientifica Ltd No patients had severe kidney failure (MDRD <15 mL/min); median PTH level was 27.3±18.8pg/mL (interquartile range 15.8-34.1) and mean albumin-adjusted total serum calcium was 8.9±1 mg/dL. Clinical and biochemical characteristics of patients are summarized in Table 1.

In the whole group (n=247) mean 25OHD levels, measured by HPLC-MS-MS were 18.1±9.5 ng/mL (45.2±23.7 nmol/L), VitD insufficiency prevalence according to IOF guidelines (<20 ng/mL or <50 nmol/L) was 61.1% (n=151) and VitD severe deficiency ( $<10 \,\mathrm{ng/mL}$ or <25 nmol/L) was 24.7% (n=61).

Patients with 25OHD levels <20 ng/mL (or 50 nmol/L) were more often obese (BMI<30, P<0.01), hypertensive (P<0.05) and in NYHA class 3 (P<0.01) compared to those with 25OHD levels >20 ng/mL. No difference in gender was found between the two groups (P=0.3).

When we compared HF patients with healthy subjects (n=76), the former had 25OHD levels lower than the latter  $(18.1\pm9.5 \,\text{ng/mL} \text{ or } 45.2\pm23.7 \,\text{nmol/L} \text{ vs } 23.3\pm9.6 \,\text{ng/mL}$ or  $58.2\pm24.0\,\mathrm{nmol/L}$ , P<0.001) and a statistically higher prevalence of VitD insufficiency (61.1% vs 39.5%, *P*<0.001) and deficiency (24.7% vs 6.6%, *P*<0.001)

During the follow-up period, median (63.4–107.3) months, 40 patients (16.2%) died, 26 (10.5%) for HF-related causes. One hundred nine patients (44.1%) were re-hospitalized, 38 (15.4%) underwent major cardiovascular complications and 19 (7.7%) had a sudden worsening of the disease.

In the whole group, there was a significant inverse relationship between baseline 25OHD and risk of HF-related death, with a HR of 0.59 (95% CI 0.37-0.92, P=0.02), but not all-cause mortality (HR 0.85, CI 0.54-1.33, P=0.48). Multivariate Cox regression analysis adjusted for age, gender, BMI and NYHA classes confirmed that 25OHD concentration remained an independent

Table 1 Clinical and biochemical data of patients in the whole group.

| 65±13<br>203 (82.2%)<br>44 (18.8%) |
|------------------------------------|
| ` '                                |
| 44 (18.8%)                         |
|                                    |
| 27±5                               |
|                                    |
| 200 (80.1%)                        |
| 47 (19.9%)                         |
| 33±8                               |
| 8.9 ± 1                            |
| 27.3 ± 18.8 (15.8–34.1)            |
|                                    |

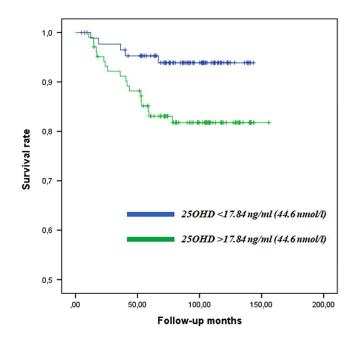
<sup>\*</sup>Values are expresses as mean ± s.d.; \*\*Values are expresses as mean ± s.d. and interquartile range.

© 2018 The authors http://www.endocrineconnections.org Published by Bioscientifica Ltd predictor of HF-related death (HR=0.62, 0.39-0.99, P=0.03)

At ROC analyses, the AUC values based on 25OHD in HF-related death was 0.66. The best cut-off of 25OHD was set at the highest Youden index and was equal to 17.8 ng/mL (44.6 nmol/L), closed to the clinical threshold, with a sensitivity of 51% and specificity of 79%. Moreover, there was also a significant inverse relationship between baseline 25OHD and the worsening of the failure, after adjustment for age, gender, BMI and NYHA classes (HR=0.42, 95% CI 0.22–0.81, P=0.010). Kaplan–Meier survival analyses according to 25OHD levels <17.8 ng/mL (44.6 nmol/L) showed that VitD insufficiency was associated with reduced survival in HF patients (log rank P = 0.017) (Fig. 1).

# Comparison between the two methods of 250HD measurement

In 151 patients, of whose blood sample was available, 25OHD levels were assessed either with LIAISON assay or by HPLC-MS-MS, from two aliquots of the same stored plasma sample. No patient was taking any kind of VitD supplementation. HPLC-MS-MS was considered the gold standard and was compared with the true value from DEQAS: 90% of results were within ±25% of the DEQASassessed values (Table 2).



Kaplan-Meier survival analyses according to 25OHD levels <17.8 ng/mL (44.6 nmol/L) showed that VitD insufficiency was associated with reduced survival in HF patients (log rank P=0.017).



This work is licensed under a Creative Commons Attribution 4.0 International License

**Table 2** 25OHD measured by DEQAS and by PerkinElmer HPLC-MS/MS from our laboratory.

| Sample 250HD | DEQAS (nmol/L) | PerkinElmer<br>MS-MS Vitamin<br>D Assay (nmol/L) | Variability (%) |
|--------------|----------------|--|-----------------|
| 1            | 45.2           | 35.9   | +25.7           |
| 2            | 67.5           | 79.6   | -15.1           |
| 3            | 103.7          | 109.9  | -5.7            |
| 4            | 32.1           | 32.5   | -1.2            |
| 5            | 102.9          | 119.1  | -13.6           |
| 6            | 39.6           | 39.4   | +0.5            |
| 7            | 83.9           | 93.6   | -10.4           |
| 8            | 22.5           | 23.5   | -4.3            |
| 9            | 107.8          | 128.3  | -16             |
| 10           | 55.6           | 68.7   | -19             |
|              |                |  |                 |

Serum 25OHD mean values were under the IOF suggested cut-off with both methods; however, when we compared the two methods, 25OHD values measured with LIAISON were statistically lower than values measured with HPLC-MS-MS ( $18.9\pm9.4\,\mathrm{ng/mL}$  or  $47.2\pm23.5\,\mathrm{nmol/L}$  vs  $19.6\pm9.4\,\mathrm{ng/mL}$  or  $49.0\pm23.5\,\mathrm{nmol/L}$ , P=0.03). When we compared the two methods using clinical thresholds of 25OHD equal to  $20\,\mathrm{ng/mL}$  (or  $50\,\mathrm{nmol/L}$ ), there was a good agreement between two methods (Cohen's kappa coefficient 0.70), but the prevalence of VitD insufficiency was significantly higher using LIAISON than using HPLC-MS-MS (58.3%, n=88 vs 55.6%, n=84, P<0.001).

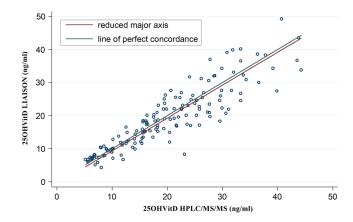
Concordance correlation coefficient of 25OHD measured by LIAISON and HPLC-MS-MS method was 0.896 and the slope between the two methods was 0.995. Figure 2 (SD line) showed that LIAISON method underestimates the 25OHD levels compared to HPLC-MS-MS.

Moreover, the inter-assay bias was evaluated by Bland–Altman plot, showed in Fig. 3. LIAISON assay showed a mean relative bias of -0.739% with 95% of limits of agreement (-9.00 to +7.52%), when compared to the HPLC-MS-MS method.

# **Discussion**

In this study, we demonstrated the high prevalence of hypovitaminosis D in a large cohort of patients with mild-to-moderate HF and the association between 25OHD levels and HF prognosis. Moreover, we showed that 25OHD levels are significantly lower in these populations, compared with matched control group. In our study, 25OHD levels were measured with HPLC-MS-MS, which was demonstrated to be the gold standard for VitD status assessment.

http://www.endocrineconnections.org https://doi.org/10.1530/EC-18-0207 © 2018 The authors Published by Bioscientifica Ltd



**Figure 2**Concordance correlation coefficient of 25OHD measured by LIAISON and HPLC-MS-MS method was 0.896 and the slope between the two methods was 0.995. In the figure (SD line), it is shown that LIAISON method underestimates the 25OHD levels compared to HPLC-MS-MS.

Convincing literature data have been collected about VitD role in pathogenesis and clinical course of cardiovascular diseases, particularly HF. A recent paper from D'Amore *et al.* summarized evidence about the possible mechanisms of VitD effects in HF and clinical studies (16). VitD could exert an antiproliferative role on cardiomyocytes, probably due to the suppression of natriuretic peptide, and it was shown to modulate extracellular matrix turnover in myocardial tissue (17, 18). Moreover, VitD acts as negative regulator of reninangiotensin-aldosterone system, as was demonstrated in knockout mice (19) and in clinical studies (20). Finally, VitD has a direct non-genomic effect on calcium channel control in cardiomyocytes (21).

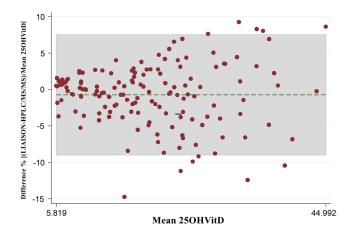


Figure 3
Bland–Altman plots shows percentage difference between 25OHD quantification with LIAISON and HPLC-MS-MS. Green line represents bias of –0.739% (means of paired difference) with 95% of limits of agreement (–9.00 to +7.52%).



This work is licensed under a Creative Commons Attribution 4.0 International License.



Subsequently, clinical cross-sectional and casecontrol studies have reported association between VitD deficiency and HF: the first study was from Shane et al. in 101 patients with severe HF in whom a high prevalence of severe VitD deficiency was demonstrated (26% of patients with 25OHD <37.5 nmol/L or 15 ng/mL) (22). In 2003, Zitterman demonstrated 25OHD levels significantly lower in a population of 54 patients with severe HF, compared to a control group (23). Our study confirms data from previous study, with the novelty that we evaluated a more common HF population with mild-to-moderate HF (prevalent NYHA class I and II), in a larger population and with HPLC-MS-MS 25OHD measurement. Even in the study of Liu et al. in which a moderate HF population was described, the prevalence of patients in NYHA class III was higher compared with our study (52% vs 20%), with mean 25OHD levels of 14.6 ng/mL or 36.6 nmol/L (20). Finally, a very large study was conducted by Gotsman et al. on 3009 patients with HF, compared with a control group in 2012, but 25OHD levels were measured with immunoassay (10). Our study confirmed a prevalence of severe VitD deficiency (<10 ng/mL or 25 nmol/L) of 24%, similar to that of Gotsman study (28%) and confirmed Gotsman evidence that 25OHD levels are lower in HF patients compared to healthy people (36.9 nmol/L IR 23.2-55.9 vs 40.7 nmol/L IR 26.7-56.9). Other data in literature corroborate this association between 25OHD and HF: in a recent community-based cohort study from Busselton Health Survey, 3946 subjects were followed up for 20 years: there was a significant inverse relationship between 25OHD levels and the risk to develop HF with HR of 0.82 (95% CI, 07-0.95), after adjustments for confounding variables (24).

Some previous studies demonstrated that low 25OHD levels were predictor of poor prognosis and survival in general population (25, 26). However, we showed that this is true in our HF patients' cohort. We confirmed data of Liu and Gotsman that 25OHD is a significant and independent predictor of reduced survival in HF patients (10, 20), but we showed that 25OHD levels are specifically associated with mortality due to HF causes and not with general mortality, after adjustment for confounding variables. In our previous study, we demonstrated a significant inverse correlation between 25OHD levels and MECKI score, even when adjusted in a multivariate model (11) MECKI score is a score of mortality risk validated in HF patients, and it is obtained from six variables independently related to prognosis. These variables include peak VO2% at cardiopulmonary test (CPET), that is reduced in HF patients with poor prognosis and that was correlated with 25OHD levels (9, 11, 27). A possible explanation of the influence of VitD on the outcome of HF patients could be its influence on cardiopulmonary function, and on peripheral skeletal muscle efficiency, with subsequent poor performance at CPET.

In this paper, we also investigated a methodological issue, with the aim to confirm that HPLC-MS-MS should be considered the gold standard for 25OHD measurement.

Indeed, the rising knowledge about VitD's paracrine and endocrine role, beyond calcium homeostasis, has highlighted the need for universal and standardized analytical method for VitD measurement.

Many analytical methods are available for VitD quantification, including competitive protein-binding immunoassays assays, enzyme-linked (ELISAs), radioimmunoassay (RIAs), chemiluminescence assays, high-performance liquid chromatography with UV (HPLC-UV), gas chromatography spectrometry and more recently liquid chromatography mass spectrometry (LC-MS-MS) (28). LIAISON assay is one of the most widely used and it is a fully automated chemiluminescence assay, which utilize an antibody with the same affinity for 25OHD2 and 25OHD3 and no apparent cross-reactivity for 3epi25OHD metabolite. Moreover, the current version, DiaSorin LIAISON Total, used in this study, was verified to have improved sensitivity and precision (29, 30) and it is approved from FDA.

On the other hand, since 1970, when high-performance liquid chromatography techniques have been introduced, they have been continuously improved and today the most recent LC-MS-MS assay is frequently referred as the 'gold standard' method (31, 32, 33). This is due to the high sensitivity, reproducibility and accuracy, and this latter also being influenced by the capability to discriminate 25OHD2 and 25OHD3, as well as their epimeric forms. Moreover, it offers the possibility to measure different VitD metabolites at the same moment (30). In 2009, the use of LC-MSMS was advised by the Nutritional Health and Nutrition Examination Survey (34) in USA and from the UK Food Standard Agency (FSA) in their National Diet and Nutrition Survey (35).

In our study, LIAISON method and HPLC-MS-MS showed a strong correlation, as already reported in other studies (12, 28). LIAISON showed a negative bias (-0.74%) compared to HPLC-MS-MS, even if it was a uniform bias across the range of doses tested. This trend was already described; however, the negative bias in our series was smaller than that in the previous studies. Indeed, in Moon *et al.* study, ADVIA Centaur and LIAISON methods were tested against LC-MS-MS and showed a negative bias of



This work is licensed under a Creative Commons Attribution 4.0 International License.

963



respectively -4.3% and -13.5% (28). To further confirm the reliability of LC-MS-MS, others studies already tested LIAISON and LC-MS-MS against those that were proposed to be the 'standard' protocols for VitD measurement. Moon et al. used standard reference material (SRM) 972 provided from United States National Institute of Standards and Technology and showed that LC-MS-MS gave the NIST closest results (compared to LIAISON and other methods) (28). Moreover, also The Vitamin D External Quality Assessment Scheme in 2011 confirmed a negative bias for LIAISON against LC-MS-MS of about -4.5% (29).

The negative bias in the correlation analyses, together with the significantly relevant difference in prevalence of VitD deficiency with the two different methods, arises some considerations about the correct clinical evaluation of hypovitaminosis D.

Indeed, in our series, VitD insufficiency (25OHD <50 nmol/L or 20 ng/mL) and deficiency (25OHD <25 nmol/L or 10 ng/mL) prevalence was significantly lower using HPLC-MS-MS, compared to that using LIAISON: that means that probably some patients would be considered as deficient, if tested with chemiluminescence assay and treated with VitD high-dose supplements. The same concern arose in a recent multicenter study in which VitD samples coming from 11 population series were reanalyzed at the University College Cork using the same protocol of LC-MS-MS, certified by CDC' Vitamin D Standardization Certification Program. They found a discrepancy in the prevalence of hypovitaminosis D in some of the reanalyzed samples: in particular, in the German population, the percentage of patients with 25OHD levels <30nmol/L was 25.9% with the original method (chemiluminescence) and 15.2% after LC-MS-MS reanalyses (36). Therefore, data from our and other studies, light up the importance of standardization in VitD measurement, for the clinical management and therapy. The most important standardization programs in the world use HPLC-MS-MS, trusting the modern HPLC-MS-MS technology. An additional strength of this study was that we directly tested our HPLC-MS-MS results (the PerkinElmer MS-MS VitD Assay) with DEQAS supplied materials.

Our study has some limitations: we have not the survival follow-up of the control group to compare the effect of VitD on mortality between HF and control group.

In conclusion, this large study on patients with mildto-moderate HF showed that 25OHD levels are low in this population and significant lower than in healthy people. Moreover, 25OHD is a significant and independent predictor of HF-related mortality.

http://www.endocrineconnections.org https://doi.org/10.1530/EC-18-0207

© 2018 The authors Published by Bioscientifica Ltd

#### **Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

# **Funding**

This research did not receive any specific grant from any funding agency in the public, commercial or not-for-profit sector.

#### **Author contribution statement**

Federica Saponaro is directly responsible for experiments, data collection, manuscript draft; Alessandro Saba, Claudio Passino, Claudio Marcocci and Riccardo Zucchi are responsible for supervision of experiments and draft writing, Frascarelli Sabina, Prontera Concetta, Clerico Aldo are responsible for 25OHD measurements, Maria Rita Sessa, Antonella Marvulli, Filomena Cetani, Elena Pardi, Simona Borsari are responsible for healthy controls samples collection, Marco Scalese is responsible for statistical analysis.

## References

- 1 Jessup M & Brozena S. Heart failure. New England Journal of Medicine 2003 **348** 2007–2018. (https://doi.org/10.1016/j.jchf.2012.10.002)
- 2 Bleumink GS, Knetsch AM, Sturkenboom MC, Straus SM, Hofman A, Deckers JW, Witteman JC & Stricker BH. Quantifying the heart failure epidemic: prevalence, incidence rate, lifetime risk and prognosis of heart failure - The Rotterdam Study. European Heart Journal 2004 25 1614-1619. (https://doi.org/10.1016/j.ehj.2004.06.038)
- 3 Wang TJ1, Pencina MJ, Booth SL, Jacques PF, Ingelsson E, Lanier K, Benjamin EJ, D'Agostino RB, Wolf M & Vasan RS. Vitamin D deficiency and risk of cardiovascular disease. Circulation 2008 117 503–511. (https://doi.org/10.1161/CIRCULATIONAHA.107.706127)
- 4 Pittas AG, Lau J, Hu FB & Dawson-Hughes B. The role of vitamin D and calcium in type 2 diabetes. A systematic review and metaanalysis. Journal of Clinical Endocrinology and Metabolism 2007 92 2017-2029. (https://doi.org/10.1210/jc.2007-0298)
- 5 Prosser DE & Jones G. Enzymes involved in the activation and inactivation of vitamin D. Trends in Biochemical Sciences 2004 29 664-673. (https://doi.org/10.1016/j.tibs.2004.10.005)
- 6 Holick MF. Vitamin D: extraskeletal Health. Rheumatic Diseases Clinics of North America 2012 38 141–160. (https://doi.org/10.1016/j. rdc.2012.03.013)
- 7 Li YC. Vitamin D regulation of the renin-angiotensin system. Journal of Cellular Biochemistry 2003 88 327-331. (https://doi.org/10.1002/ icb.10343)
- 8 Rahman A, Hershey S, Ahmed S, Nibbelink K & Simpson RU. Heart extracellular matrix gene expression profile in the vitamin D receptor knockout mice. Journal of Steroid Biochemistry and Molecular Biology 2007 103 416-419. (https://doi.org/10.1016/j.jsbmb.2006.12.081)
- 9 Boxer RS, Kenny AM, Cheruvu VK, Vest M, Fiutem JJ & Piña II. Serum 25-hydroxyvitamin D concentration is associated with functional capacity in older adults with heart failure. American Heart Journal 2010 160 893-899. (https://doi.org/10.1016/j. ahi.2010.08.004)
- 10 Gotsman I, Shauer A, Zwas DR, Hellman Y, Keren A, Lotan C & Admon D. Vitamin D deficiency is a predictor of reduced survival in patients with heart failure; Vitamin D supplementation improves outcome. European Journal of Heart Failure 2012 14 357-366. (https:// doi.org/10.1093/eurjhf/hfr175)
- 11 Saponaro F, Marcocci C, Zucchi R, Prontera C, Clerico A, Scalese M, Frascarelli S, Saba A & Passino C. Hypovitaminosis D in patients with heart failure: effects on functional capacity and patients' survival. Endocrine 2017 58 574-581. (https://doi.org/10.1007/s12020-017-1282-9)



This work is licensed under a Creative Commons Attribution 4.0 International License

F Saponaro et al.



- 12 Hsu SA, Soldo J & Gupta M. Evaluation of two automated immunoassays for 25-OH vitamin D: comparison against LC-MS/ MS. Journal of Steroid Biochemistry and Molecular Biology 2013 136 139–145. (https://doi.org/10.1016/j.jsbmb.2012.11.005)
- 13 Tai SS-C, Bedner M & Phinney KW. Development of a candidate reference measurement procedure for the determination of 25-hydroxyvitamin D3 and 25-hydroxyvitamin D2 in human serum using isotope-dilution liquid chromatography-tandem mass spectrometry. Analytical Chemistry 2010 82 1942–1948. (https://doi. org/10.1021/ac9026862)
- 14 Federico G, Genoni A, Puggioni A, Saba A, Gallo D, Randazzo E, Salvatoni A & Toniolo A. Vitamin D status, enterovirus infection, and type 1 diabetes in Italian children/adolescents. Pediatric Diabetes 2018 19 923-929. (https://doi.org/10.1111/pedi.12673)
- 15 CLSI. EP07-A2 Interference Testing in Clinical Chemistry; Approved Guideline, 2nd ed. CLSI: Wayne, PA, USA 2005.
- 16 Amore CD, Marsico F, Parente A, Paolillo S, De Martino F & Gargiulo P. Nutrition, metabolism & cardiovascular diseases vitamin D deficiency and clinical outcome in patients with chronic heart failure: a review. Nutrition, Metabolism and Cardiovascular Diseases 2017 **27** 837–849. (https://doi.org/10.1016/j.numecd.2017.07.009)
- 17 Weber KT, Weglicki WB & Simpson RU. Macro- and micronutrient dyshomeostasis in the adverse structural remodelling of myocardium. Cardiovascular Research 2009 81 500-508. (https://doi. org/10.1093/cvr/cvn261)
- 18 Demer LL & Tintut Y. Vascular calcification: pathobiology of a multifaceted disease. Circulation 2008 117 2938-2948. (https://doi. org/10.1161/CIRCULATIONAHA.107.743161)
- 19 Bouillon R, Carmeliet G, Verlinden L, Van Etten E, Verstuyf A, Luderer HF, Lieben L, Mathieu C & Demay M. Vitamin D and human health: lessons from vitamin D receptor null mice. Endocrine Reviews 2008 **29** 726–776. (https://doi.org/10.1210/er.2008-0004)
- 20 Liu LC, Voors AA, van Veldhuisen DJ, van der Veer E, Belonje AM, Szymanski MK, Silljé HH, van Gilst WH, Jaarsma T & de Boer RA. Vitamin D status and outcomes in heart failure patients. European Journal of Heart Failure 2011 13 619-625. (https://doi.org/10.1093/ eurjhf/hfr032)
- 21 Green JJ, Robinson DA, Wilson GE, Simpson RU & Westfall MV. Calcitriol modulation of cardiac contractile performance via protein kinase C. Journal of Molecular and Cellular Cardiology 2006 41 350-359. (https://doi.org/10.1016/j.yjmcc.2006.05.019)
- 22 Shane E, Mancini D, Aaronson K, Silverberg SJ, Seibel MJ, Addesso V & McMahon DJ. Bone mass, vitamin D deficiency, and hyperparathyroidism in congestive heart failure. American Journal of Medicine 1997 103 197-207. (https://doi.org/10.1016/S0002-9343(97)00142-3)
- 23 Zittermann A, Schleithoff SS, Tenderich G, Berthold HK, Körfer R & Stehle P. Low vitamin D status: a contributing factor in the pathogenesis of congestive heart failure? Journal of the American College of Cardiology 2003 41 105-112. (https://doi.org/10.1016/ \$0735-1097(02)02624-4)
- 24 Zhu K, Knuiman M, Divitini M, Hung J, Lim EM, Cooke BR & Walsh IP. Serum 25-hydroxyvitamin D as a predictor of mortality and cardiovascular events: a 20-year study of a community-based cohort. Clinical Endocrinology 2018 88 154-163. (https://doi.org/10.1111/ cen.13485)

- 25 Pilz S, Marz W, Wellnitz B, Seelhorst U, Fahrleitner-Pammer A, Dimai HP, Bohem BO & Dobnig H. Association of vitamin D deficiency with heart failure and sudden cardiac death in a large cross-sectional study of patients referred for coronary angiography. Journal of Clinical Endocrinology and Metabolism 2008 93 3927-3935. (https://doi.org/10.1210/jc.2008-0784)
- 26 Melamed ML, Michos ED, Post W & Astor B. 25-hydroxyvitamin D levels and the risk of mortality in the general population. Archives of Internal Medicine 2008 168 1629-1637. (https://doi.org/10.1001/ archinte.168.15.1629)
- 27 Kaul A, Gläser S, Hannemann A, Schäper C, Nauck M, Felix SB, Bollmann T, Ewert R & Friedrich N. Vitamin D is associated with cardiopulmonary exercise capacity: results of two independent cohorts of healthy adults. British Journal of Nutrition 2015 25 1-9. (https://doi.org/10.1017/S000711451500464X)
- 28 Moon HW, Cho JH, Hur M, Song J, Oh GY, Park CM, Yun JM & Kim JQ. Comparison of four current 25-hydroxyvitamin D assays. Clinical Biochemistry 2012 45 326-330. (https://doi.org/10.1016/j. clinbiochem.2011.12.025)
- 29 Carter GD. Accuracy of 25-hydroxyvitamin D assays: confronting the issues. Current Drug Targets 2011 12 19-28. (https://doi. org/10.2174/138945011793591608)
- 30 Müller MJ & Volmer DA. Mass spectrometric profiling of Vitamin D metabolites beyond 25-hydroxyVitamin D. Clinical Chemistry 2015 61 1033-1048. (https://doi.org/10.1373/ clinchem.2015.241430)
- 31 El-Khoury JM, Reineks EZ & Wang S. Progress of liquid chromatography-mass spectrometry in measurement of vitamin D metabolites and analogues. Clinical Biochemistry 2011 44 66-76. (https://doi.org/10.1016/j.clinbiochem.2010.05.007)
- 32 Hollis BW. Measuring 25-hydroxyvitamin D in a clinical environment: challenges and needs. American Journal of Clinical Nutrition 2008 88 507S-510S. (https://doi.org/10.1093/ aicn/88.2.507S)
- 33 Carter GD. 25-Hydroxyvitamin D assays: the quest for accuracy. Clinical Chemistry 2009 **55** 1300–1302. (https://doi.org/10.1373/ clinchem.2009.125906)
- 34 Yetley EA, Pfeiffer CM, Schleicher RL, Phinney KW, Lacher DA, Christakos S, Eckfeldt JH, Fleet JC, Howard G, Hoofnagle AN, et al. NHANES monitoring of serum 25-hydroxyvitamin D: a roundtable summary. Journal of Nutrition 2010 140 2030S-2045S. (https://doi. org/10.3945/jn.110.121483)
- 35 de la Hunty A, Wallace AM, Gibson S, Viljakainen H, Lamberg-Allardt C & Ashwell M. UK Food Standards Agency Workshop Consensus Report: the choice of method for measuring 25-hydroxyvitamin D to estimate vitamin D status for the UK National Diet and Nutrition Survey. British Journal of Nutrition 2010 104 612-619. (https://doi.org/10.1017/ S000711451000214X)
- 36 Cashman KD, Kiely M, Kinsella M, Durazo-Arvizu RA, Tian L, Zhang Y, Lucey A, Flynn A, Gibney MJ, Vesper HW, et al. Evaluation of Vitamin D Standardization Program protocols for standardizing serum 25-hydroxyvitamin D data: a case study of the program's potential for national nutrition and health surveys. American Journal of Clinical Nutrition 2013 97 1235-1242. (https://doi.org/10.3945/ ajcn.112.057182.Knowledge)

Received in final form 12 July 2018 Accepted 24 July 2018 Accepted Preprint published online 26 July 2018



This work is licensed under a Creative Commons Attribution 4.0 International License