Elsevier Editorial System(tm) for Ophthalmology Manuscript Draft

Manuscript Number:

Title: Pathophysiology and pharmacological targets of diabetic macular edema: an updated review

Article Type: Translational Science Reviews

Keywords: Diabetic macular edema, diabetic retinopathy, VEGF, angiogenesis, inflammation

Corresponding Author: Prof. Romano Danesi, MD, PhD

Corresponding Author's Institution: University of Pisa

First Author: Stefano Fogli, MD, PhD

Order of Authors: Stefano Fogli , MD, PhD; Selene Mogavero, BSc, PhD; Colin Gerard Egan, BSC, PhD; Romano Danesi, MD, PhD

Abstract: Diabetic macular edema (DME) is a serious condition that can cause blindness in diabetic patients suffering from diabetic retinopathy (DR). Although vascular endothelial growth factor (VEGF) is known to play a role in the development of DME, the pathological processes leading to the onset of the disease are highly complex and the exact sequence in which they occur is still not completely understood. Angiogenesis and inflammation have been shown to be involved in the pathogenesis of this disease; however, whether angiogenesis following VEGF over-expression is a cause or a consequence of inflammation remains to be clarified. Here, we provide an overview of the current data available in the literature focusing on VEGF, angiogenesis, inflammation, DR and DME. Our analysis suggests that angiogenesis and inflammation act interdependently during the development of DME and that VEGF is a critical player in the molecular crosstalk occurring between these two pathways. Consequently, anti-VEGF therapies hold potential for the treatment of DME.

Angiogenesis and inflammation act interdependently during the development of diabetic macular edema and vascular endothelial growth factor is a critical player in the molecular crosstalk occurring between these two pathways.

Pathophysiology and pharmacological targets of diabetic macular edema: an 1 updated review 2 3 4 Stefano Fogli MD, PhD¹, Selene Mogavero BSc, PhD², Colin Gerard Egan BSc, PhD², Romano 5 Danesi MD, PhD¹ 6 7 8 ¹Department of pharmacology and chemotherapy, "Università di Pisa", Pisa, Italy ²Primula Multimedia S.r.L., Pisa, Italy 9 10 11 12 **Corresponding author/address for reprints:** 13 Romano Danesi, MD, PhD 14 Via Savi, 10 15 56126, Pisa 16 Tel: +39 050 992632 17 Fax: +39 050 2218758 18 Email: romano.danesi@med.unipi.it 19 20 21 **Financial support:** 22 Financial support for medical editorial assistance was provided by Novartis Pharma, Italy. The sponsor had no role in the design or conduct of this review. 23 24 25 **Conflict of interest:** 26 No conflicting relationship exists for any author. 27 28 29 **Runnning head:** 30 Pathophysiology and pharmacology targets of diabetic macular edema.

31

32 Abstract

33

34 Diabetic macular edema (DME) is a serious condition that can cause blindness in diabetic patients 35 suffering from diabetic retinopathy (DR). Although vascular endothelial growth factor (VEGF) is 36 known to play a role in the development of DME, the pathological processes leading to the onset of the disease are highly complex and the exact sequence in which they occur is still not completely 37 38 understood. Angiogenesis and inflammation have been shown to be involved in the pathogenesis of 39 this disease; however, whether angiogenesis following VEGF over-expression is a cause or a 40 consequence of inflammation remains to be clarified. Here, we provide an overview of the current 41 data available in the literature focusing on VEGF, angiogenesis, inflammation, DR and DME. Our 42 analysis suggests that angiogenesis and inflammation act interdependently during the development 43 of DME and that VEGF is a critical player in the molecular crosstalk occurring between these two 44 pathways. Consequently, anti-VEGF therapies hold potential for the treatment of DME.

45 46

47 Key words: Diabetic macular edema, diabetic retinopathy, VEGF, angiogenesis, inflammation

48 Introduction

49	Diabetic patients often suffer from diabetic retinopathy (DR) leading to diabetic macular edema
50	(DME), the most common cause of visual loss in this set of patients. ^{1,2} Pathogenesis of this
51	condition is complex and involves several physiological alterations.
52	The blood retinal barrier (BRB) plays a key role here, as its disruption leads to several pathological
53	conditions of the eye such as age-related macular degeneration (ARMD), retinal vein occlusions
54	(RVO) and other chronic retinal diseases. ³
55	In order to understand how the BRB breakdown is involved we need to take a step back and
56	understand what occurs in diabetes.
50	
57	In diabetic patients, hyperglycemia is the triggering factor for tissue alterations such as damages to
58	the capillary endothelial cells in the retina. ⁴ This occurs through several pathways (Figure 1):
59	• the increased polyol pathway where increased glucose concentration leads to hyperglycemic
60	oxidative stress ⁵ ;
61	• the increased formation of advanced glycation end-products (AGEs), which alters
62	intracellular and transmembrane proteins such as integrins and integrin receptors, thus
63	disturbing crucial interactions with proteins of the basal lamina ^{6,7} ;
64	• the activation of protein kinase C (PKC) isoforms, which leads to an increased production of
65	extracellular matrix and cytokines, enhanced contractility, permeability, and vascular cell
66	proliferation, activation of cytosolic phospholipase A2, inhibition Na+-K+-ATPase, all
67	leading to abnormal retinal hemodynamics. ⁸

One hypothesis is that the complex tissue alterations lead to cellular hypoxia.⁹ In response to
hypoxia, vascular endothelial growth factor (VEGF) production is induced, a key promoter of
angiogenesis, abnormal vascular permeability, and, eventually, inflammatory reaction.¹⁰ One of the
most important consequences of the formation of AGEs is also the induction of VEGF.^{11,13}

Being VEGF a promoter of the inflammatory process, leading to hypoxia, and being hypoxia
responsible for the increased expression of VEGF, it becomes clear that this is a self-fueling
exponential loop always leading to increased tissue damage.

75 Negative consequences of increased VEGF expression due to diabetic hyperglycemia are described in several anatomical locations other than the retina, such as the kidney^{14,15} or the brain.¹⁶ In the 76 77 eye, increased VEGF expression leads to pathologic transformation of the retinal vasculature, including permeability, remodeling and neovascularization.¹⁷ Several VEGF isoforms exist, 78 VEGF₁₆₅ being not only the predominant form in the diabetic eye, but, among the VEGF isoforms, 79 also the most potent inducer of leukostasis and BRB breakdown, as shown in an animal model.¹⁸ 80 81 VEGF binds to its receptors on the vascular endothelium and activates the mitogen-activated protein kinase (MAPK) thus triggering endothelial cell proliferation.¹⁰ VEGF is also a positive 82 regulator of angiogenesis by stimulating endothelial cells to degrade their basement membrane and 83 to migrate by releasing matrix metalloproteinases (MMPs) and plasminogen activators (PAs) and by 84 increasing the expression of integrins.¹⁹ Proliferation and migration of endothelial cells is followed 85 by synthesis of basement membranes for the newly formed capillaries.¹⁰ Not surprisingly, similar 86 mechanisms are found in tumor development.²⁰ 87

88 It therefore becomes clear that VEGF plays a predominant role in the pathogenesis of DME, serving 89 as an attractive target for therapy. Nevertheless, there still is substantial uncertainty on the temporal 90 development of retinal alterations, and the main question that arises from the extensive literature on

this topic, is: what comes first? Is angiogenesis leading to inflammation or the contrary? What is the
exact mechanism leading to inflammation and, is it subsequent to angiogenesis? Is it angiogenesis
that, due to the production of mediators, such as VEGF, activates the production of nitric oxide and
all that could potentially be correlated to tissue damage, increased vascular permeability,
endothelial cell proliferation, vascular occlusion ischemic cell death and therefore inflammation, or
is it inflammation that triggers the expression of VEGF and subsequently leads to angiogenesis,
hyperpermeability and so on?

98 The purpose of this review is therefore to examine all available literature, which points toward one

99 explanation or the other, to eventually arrive at a conclusion to this highly debated topic. A

100 thorough summary of all studies analyzed in the next sections comparing various aspects of each

101 study is provided (**Supplementary Table 1**).

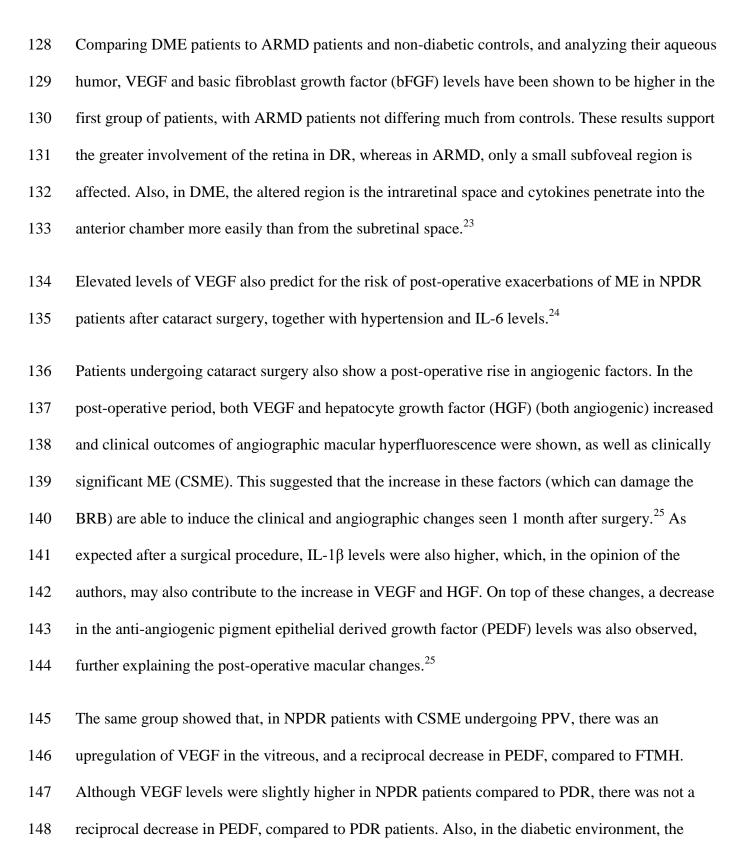
103 Studies supporting a predominant role of angiogenesis on the pathogenesis of diabetic

104 retinopathy

105 There are several studies supporting the idea of angiogenesis being the main reason for DME. An elegant study by Shimada and coworkers evaluated VEGF concentrations from different sites in the 106 107 vitreous. By taking samples from the pre-macular region, the peripheral cortical vitreous and the 108 mid vitreous they showed that VEGF levels are higher in the pre-macular vitreous compared to the 109 other two sites. Also, VEGF correlated with foveal thickness (FT), and consequently with DME 110 severity. This study demonstrates that there is a diffusion of VEGF from the macular region to the 111 periphery and from the posterior to the anterior globe, forming a concentration gradient. Also, 112 VEGF was associated with the presence and severity of DME. But as the authors state in the 113 discussion, "...while these findings demonstrate that VEGF levels in the vitreous are associated with the presence and severity of DME, they do not prove cause and effect. The role of VEGF in the 114 115 production of DME can only be proven by interventional approaches, and hopefully a conclusion may be drawn when current clinical trials of anti-VEGF agents for DME are completed...".²¹ 116

117 The sequence of phenomena has been tried to be verified in a study involving NPDR, PDR and full 118 thickness macular hole (FTMH) patients (as controls). The study shows that, as disease progresses 119 from NPDR to PDR, with capillary loss and retinal ischemia, inflammation increases, since IL-1β 120 concentrations are almost undetectable in NPDR and controls, but raised in PDR patients. 121 Analogously, the interleukin-1 receptor antagonist (IL-1Ra), a member of the IL-1 family that binds 122 to IL-1 receptors but does not induce any intracellular response and prevent IL-1 mediated 123 inflammation, was significantly higher in the control vitreous compared to the diabetic vitreous, 124 meaning that, as disease progresses, proinflammatory cytokines remain unresponsive. Also, retinal 125 microcirculation changes have been noted even before the onset of clinical disease, with endothelin-

1 lower in NPDR compared to PDR, reflecting the high blood flow in NPDR as compared to the
lower blood flow in PDR.²²



149 soluble VEGF receptor (sVEGFR)-1, an anti-angiogenic growth factor, was less concentrated. The authors suggested that in PDR VEGF, though lower than in NPDR, is still capable of producing the 150 151 angiogenesis observed in PDR since both sVEGFR-1 and PEDF levels are low. The full angiogenic potential in NPDR is limited by the sufficiently high levels of PEDF.²⁶ The authors also propose 152 153 that structural and molecular optical coherence tomography (OCT) macular profiles may explain 154 different responses to PPV in DME: when a posterior hyaloid traction is present, macular volume decreases after PPV independent of VEGF concentration, suggesting that raised TGF-B1 stimulates 155 156 a fibrotic response in the posterior hyaloid providing the mechanism for generating tractional forces which cause DME. When a combined diffuse macular thickening and an elevated VEGF level is 157 present, both decrease after PPV indicating that VEGF may be important in the etiology in this 158 group.²⁶ 159

Similar results were later obtained by Javanmard and coworkers, who demonstrated that, although no difference was detectable in aqueous VEGF levels between NPDR patients and normoglycemic controls, sVEGFR-1 levels were significantly decreased in the test subjects versus control.²⁷ The ratio VEGF/sVEGFR-1 was positively correlated with FT. They suggested that the decreased chelating effects of sVEGFR-1 could allow VEGF to induce permeability, so it is the imbalance between VEGF and sVEGFR-1 that determines the fate of DME.²⁷

Asato and coworkers, on the other hand, did not find any difference in sVEGFR-1 levels among
different eye diseases, including idiopatic macular hole (MH), branched RVO (BRVO), central
RVO (CRVO), DME and PDR patients. However, they did note that sVEGFR-1 correlated with age
and that in active PDR sVEGFR-1 levels were lower compared to quiescent PDR, suggesting that
this might be the reason why PDR tends to be more aggressive in youth.²⁸

171 Anti-permeability factors have also been involved in this pathological mechanism, which does not 172 seem to have a simple explanation. Angiopoietin-1 (ANG-1) may act as a anti-permeability factor and ANG-2 antagonizes ANG-1.²⁹ The predominance of ANG-2 in NPDR with CSME could 173 promote an increased permeability combined with the elevated levels of VEGF, facilitating the 174 BRB breakdown.³⁰ 175 Angiotensin II (AII) is yet another factor related to the increase in vascular permeability in DME, 176 together with VEGF.³¹ Vitreous concentrations of AII were increased in patients with DME 177 178 compared to non-diabetic patients and VEGF was increased also compared to diabetics without 179 retinopathy. Also, AII and VEGF correlated with each other and were higher in hyperfluorescent 180 DME compared to hypofluorescent, hinting towards a correlation with disease severity.

181 Figure 2 shows a schematic representation of angiogenesis events followed by inflammation.

183 Studies supporting predominant role of inflammation on the pathogenesis of diabetic

184 retinopathy

A recent study by Umazume and coworkers demonstrated that soluble CD14 (sCD14) may act as a
key regulator of DME, since this mediator has been found to be elevated in DME patients compared
to controls.³² A correlation between sCD14 and interleukin-8 (IL-8) or monocytochemotactic
protein-1 (MCP-1) had also been found in the vitreous fluid of patients with proliferative DR
(PDR).³³ It is therefore possible that sCD14 is involved in the upregulated expression of IL-8 and
MCP-1 in DME patients.³²

Cytokines that repeatedly have been found elevated in DR/DME are interleukin-6 (IL-6) and IL-8. 191 Sonoda and coworkers found that, in patients with type 2 diabetes mellitus and DR undergoing pars 192 plana vitrectomy (PPV), IL-6 was the factor most significantly associated with the presence of a 193 194 serous retinal detachment (SRD). They therefore suggested that VEGF cannot be the only factor 195 responsible for the pathogenesis of DR and speculated that the presence of IL-6 increases the 196 inflammatory reaction in the outer retina resulting in a further disruption of the external limiting membrane.³⁴ It is therefore not surprising that the condition of SRD responds well to corticosteroid 197 therapy³⁵, but the fact that there is a strong correlation between SRD and IL-6 levels only means 198 199 that inflammation facilitates retinal detachment. Furthermore, it does not necessarily mean that 200 inflammation is involved in the pathogenesis of DME, but it may be an effect secondary to the 201 underlying angiogenetic process.

The possibility that VEGF can promote inflammation-induced damage in DR triggered by the two most fount cytokines, IL-6 and IL-8, is supported by several studies. Koskela et al showed that the increased cytokine concentrations in the vitreous of PDR patients were due to intra-ocular changes rather than to BRB breakdown, which had been damaged by DR.³⁶ The origin of the vitreous

206 inflammatory factors IL-6 and IL-8 was the retina or other ocular tissues. A theory that emerges is 207 that there might be a common pathway involved in the inflammation process in vitreoretinal 208 diseases. In DME, indeed, again IL-6 and IL-8, together with MCP-1, have been suggested to be promoting vascular permeability causing the pathology, and that ischemia and VEGF further 209 promote DME to develop into PDR.³⁷ Although they argue that the concentrations of inflammatory 210 211 soluble factors might not necessarily reflect a pathogenic process, in this study it is strongly believed that the high correlation between the three factors indicates that common pathways are 212 involved in the pathogenesis of various vitreoretinal disorders.³⁷ But how can they exclude that 213 214 VEGF was not the initial promoter of inflammation following angiogenesis? Based on this study, 215 this cannot be excluded; indeed, one of the doubts that the authors have is that a substantial amount 216 of VEGF can be initially produced by the sudden profound retinal ischemia, which in turn induces the major three factors, i.e. IL-6, IL-8 and MCP-1, afterward.³⁷ 217

These same 3 cytokines, in addition to induced protein-10 (IP-10), were found to be elevated in a subsequent study including non-PDR (NPDR) and PDR patients.³⁸ The study suggested that the simultaneous measurement of several factors in the same study samples may reveal the relative contribution of each factor to the pathogenesis of DR. VEGF had a major role in PDR mediating ocular angiogenesis. But, as for many similar studies, the authors eventually concluded that further investigations are required to define the precise roles of these factors in the pathogenesis of DR and DME.³⁸

Funk and coworkers observed slightly different results, finding, in the aqueous humor of DME patients at an advanced stage of disease, only MCP-1 and IL-8 significantly elevated compared to controls, and IL-6 and VEGF only slightly, but not significantly, higher. Their conclusion was that the inflammatory markers MCP-1 and IL-8 might have a role in the pathogenesis of DME.³⁹ Bevacizumab therapy, in these patients, was not correlated with changes in clinical disease activity,
and other growth factors or inflammatory cytokines did not change over time, explaining the
negative biologic response. One major limitation of the study, though, was the small sample size
(10 DME patients and 10 controls undergoing cataract surgery).³⁹

233 A different approach was adopted by Shimura and coworkers where patients with bilateral PDR requiring PPV were treated with pan-retinal photocoagulation (PRP) in one eye and not in the other. 234 235 PRP induced the worsening of macular edema (ME) and this was linked with pro-inflammatory 236 cytokines such as IL-6 and RANTES (Regulated upon Activation Normal T-cell Expressed and Secreted), but not with VEGF and stromal derived factor-1 (SDF-1).⁴⁰ They speculated that a 237 238 possible reason was that the vitreous level of VEGF in PDR had been saturated before PRP, so no 239 increase could have been measured. Interestingly, in eyes not undergoing PRP (controls), "spontaneous" ME appeared, and therefore the status of ME in the control eyes was correlated with 240 241 vitreous levels of VEGF. Another possible explanation the authors gave was that inflammatory 242 cytokines, compared with VEGF, had a major role in the pathogenesis of DME. For sure, the pathogenesis of PRP-induced ME was likely to be different from that of "spontaneous" DME.⁴⁰ 243 In PDR patients who had ME, IL-6 was found to be higher compared to the same type of patients 244 without ME.⁴¹ An interesting result of this study was also that IL-6 was not correlated with age, 245 duration of diabetes, vitreous hemorrhage, PRP, type of therapy, hyperglycemia or renal function. 246 247 This is another study supporting a major role of IL-6 in the development of ME in DR patients. An 248 interesting approach that the authors suggested for further studies is the determination of mRNA 249 levels of both IL-6 and IL-6 receptors in the vitreous, which may be important in understanding the 250 temporal association of stimuli such as hypoxia, hyperglycemia, and growth factors (e.g., VEGF) 251 with the induction of IL-6 synthesis, as well as in analyzing molecular responses to potential antiinflammatory treatment strategies.⁴¹ 252

- 253 To our knowledge, the only study that found elevated IL-6 levels in plasma, compared to others that
- consistently detected this increase in the vitreous or aqueous humor in the eye, was performed by
- 255 Shimizu and coworkers.¹² They observed that plasma levels of IL-6 correlated with the severity of
- 256 ME, along with the presence of posterior vitreous detachment, while plasma levels of VEGF,
- 257 transforming growth factor- β 1 (TGF- β 1), and tumor necrosis factor- α (TNF- α) did not correlate
- 258 with ME.¹²
- 259 Figure 3 shows a schematic representation of inflammatory events followed by angiogenesis.
- 260

261 Studies supporting both angiogenesis and inflammation as causative of diabetic retinopathy

262 VEGF contribution to the pathogenesis of PDR has been confirmed by the "ex-adiuvantibus" results that anti-VEGF therapy is efficacious in the treatment of DR. A recent study by Costagliola and 263 coworkers showed that, together with VEGF, also adiponectin (APN) is upregulated in PDR 264 265 compared to non-diabetic controls. They also showed that the anti-VEGF molecule bevacizumab induced a decrease in both VEGF and APN, decreasing FT and improving best-corrected visual 266 acuity (BCVA). But they argued that, since anti-VEGF treatment is not associated with total 267 268 regression of retinal neovascularization secondary to PDR, it might not neutralize other inflammatory molecules involved in the cascade of the BRB breakdown, such as insulin-like growth 269 270 factor-1 (IGF-1), ANG, SDF-1, bFGF-2, HGF, TNF, IL-6, erythropoietin (EPO) and Pigment 271 epithelium-derived factor (PEDF), which are those identified as novel factors in the DR pathogenesis.42 272

273 Two studies published in 2012, one by Lee et al. and another by Jonas et al., and one study from 274 2011 by Suzuki et al., can perhaps be considered the most comprehensive analyses of inflammatory and angiogenic factors in DME.⁴³⁻⁴⁵ The first study analyzed the aqueous humor of DME patients 275 and compared it to that of BRVO-ME and of normal controls. A total of fourteen different 276 277 inflammatory and angiogenic cytokines were analyzed and among these, they showed that DME, IL-8, MCP-1, platelet derived growth factor (PDGF)-AA and VEGF levels were higher and IL-13 278 279 lower compared to controls. Compared to BRVO-ME, DME patients had higher IL-6 and MCP-1 280 levels. IL-8 correlated positively and interferon (IFN)-y negatively with DME severity. In BRVO-281 ME, IL-8 positively correlated with ME severity, as for DME, and with retinal ischemia. The 282 authors concluded that the inflammatory reaction in DME is very active, certainly to a greater 283 extent than in BRVO-ME. Also, the relatively gradual course of the disease could result in slow 284 upregulation of VEGF and vascular remodeling, or a fibrotic process could continuously occur

285 through expression of MCP-1. The authors eloquently describe the same concerns we share: first of all, they point out that it is not appropriate to assume that a particular cytokine plays a role in 286 287 pathogenesis based simply upon measurement of elevated levels in the aqueous. The particular 288 cytokine is released as a result of the disease process, and it could not be the cause of the disease 289 process. Second, since they did not compare those concentrations between DR or BRVO with and 290 without ME, it seems to be difficult to consider that the cytokines which had aqueous 291 concentrations significantly higher than those in controls, may play a role in the development of 292 ME. Third, they could not control all possible confounding variables, such as time from onset, which can affect cytokine levels in the eye.⁴⁵ 293

294 The second study compared aqueous humor levels of 34 different molecules among cytokines in 295 patients with diffuse DME and controls undergoing cataract surgery. On top of elevated levels of VEGF, DME eyes showed an increase in many different cytokines including epidermal growth 296 297 factor (EGF), HGF, IL-1a2, IL-6, IL-8, IFN-γ-IP10, MCP-1, vascular cell adhesion molecule 298 (VCAM), monokine induced by IFN-γ (MIG), MMP-1, MMP-9, PA inhibitor (PAI)-1, placenta 299 growth factor (PIGF), and TGF-B, most of which were associated with retinal macula thickness 300 (RMT). Intracellular adhesion molecule (ICAM)-1 was the cytokine most associated to DME and 301 its severity and VEGF levels correlated with many other cytokine levels. Caution in concluding that 302 "elevated concentrations of molecules in the eyes with DME were causally related to DR and may 303 thus be therapeutic targets" has also been proposed by the authors. They propose that an explanation 304 could also be given by retinal leakage due to an insufficient BRB, leakage from the ciliary body 305 directly into the aqueous humor in the case that the concentrations of these cytokines were systemically elevated in the blood, or a local production or hyperproduction of the cytokines in the 306 diseased retina.44 307

308 The third study was performed on patients with DR, with CRVO and controls with idiopatic 309 epiretinal membrane and MH. In this study the authors presented results from the simultaneous 310 identification of 27 different cytokines and chemokines. The elevated molecules in DME in this study were IL-1Ra, IL-6, IL-8, IL-10, IL-13, IP-10, MCP-1, macrophage inflammatory protein-18 311 312 (MIP-1 β), PDGF and VEGF. So, on top of VEGF, the authors suggest that other cytokines and 313 chemokines may be involved in the pathogenesis of DR and CRVO, and that they are correlated with VEGF levels in the vitreous. The most significantly correlated cytokines to VEGF were IL-10, 314 315 IL-13 and PDGF, suggesting that not only inflammation as well as ischemia is active in the vitreous 316 body of the retina of DR patients, but also that inflammation may activate an intrinsic defense 317 mechanism. The logical conclusion drawn by these authors is that treatment options should simultaneously target inflammation and ischemia, according to the stage of the disease.⁴³ 318 319 In a study performed on bilateral DME patients and cataract surgery controls, the DME patients 320 being treated with an intravitreal corticosteroid (triamcinolone acetonide, IVTA) in one eye and an 321 intravitreal anti-VEGF molecule (bevacizumab, IVBe) in the other, the authors show that the pathogenesis of DME is not only related to VEGF, but many cytokines may be involved. IL-8, IP-322 10, MCP-1 and VEGF were significantly higher in DME patients versus controls, and IVTA 323 324 significantly reduced IL-6, IP-10, MCP-1 PDGF-AA and VEGF, more than IVBe, which only 325 reduced VEGF, but to a larger extent than IVTA. Interestingly, no significant difference in IL-6 aqueous levels was observed between the DME and control groups prior to drug administration.⁴⁶ 326 327 Another factor that might be associated with DR, and could be used as a biomarker, is soluble IL-6 328 receptor (sIL-6R) which was found to be elevated in the vitreous of both proliferative and pre-329 proliferative DR, compared to non diabetic controls. Its levels also correlated with levels of VEGE.⁴⁷ 330

331 Two studies from the same group, slightly differing from each other, showed that both VEGF and 332 IL-6 are elevated in DME and correlated with disease severity. Sampling was performed in aqueous humor and plasma in one study ⁴⁸, and in vitreous humor and plasma in the other.⁴⁹ IL-6 and VEGF 333 correlated with each other. The suggestion therefore was that both VEGF and IL-6 are produced 334 335 together in the intraocular tissues and are involved in the pathogenesis of DME, and it could be either in concert or IL-6 production via VEGF.^{48,49} They actually proposed 3 possibilities: both 336 VEGF and IL-6 may indirectly cause an increase of vascular permeability; IL-6 may indirectly 337 338 cause an increase of vascular permeability via upregulation of VEGF; VEGF alone may cause 339 vascular permeability to increase, with the elevated vitreous level of IL-6 being related to hyperglycemia and not having an influence on vascular permeability.⁴⁹ 340 Figure 4 shows a schematic representation of how angiogenesis and inflammation are part of a 341

342 network of events ultimately leading to tissue damage.

344 **Other studies**

Besides the clinical studies performed on various types of patient with various degrees of
retinopathy, in vitro studies provide useful information in order to elucidate the sequentiality of
phenomena.

348 Cohen and coworkers showed that treatment of various cell lines with IL-6 for 6–48 h results in a 349 significant induction of VEGF mRNA. The induced transcription is mediated by specific DNA 350 motifs located on the putative promoter region of VEGF as well as by specific elements located in 351 the 5'-UTR (untranslated region). IL-6 exerts its biological effects through association with specific 352 cell surface receptors resulting in the activation of specific transcription factors that interact with 353 two types of cis-acting DNA control elements mediating IL-6 response. So they conclude that 354 induction of IL-6 by hypoxia may promote the expression of VEGF that eventually leads to angiogenesis.⁵⁰ They also showed that other cytokines like IFN- β and TNF- α can induce the 355 transcription of VEGF mRNA. Being IFN-β expressed during inflammation, rheumatoid arthritis, 356 357 and wound healing, the authors think it is probable that expression of IFN- β in response to these 358 disorders might be one of the signals that triggers the angiogenic process through the induction of VEGF expression.⁵⁰ 359

The first direct demonstration that VEGF can increase vascular permeability in the eye at clinically relevant concentrations and activate PKC isoforms in the retina was provided by Aiello and coworkers in 1997.⁵¹ In this study, intravitreal VEGF administration to adult rats rapidly increased retinal vascular permeability. PKC mediation of VEGF-stimulated retinal vascular permeability in vivo was supported by multiple findings, including >98% suppression of the vasopermeability response using PKC inhibitors, mimicking of the vasopermeability response using PKC agonists, and direct activation of retinal PKC activity by intravitreal injection of VEGF.⁵¹

Another ex vivo study was performed on primary porcine retinal pigment epithelium (RPE) cells 367 and the human RPE cell line ARPE-19, in order to study the mechanisms responsible for VEGF-368 369 mediated changes in RPE permeability. The administration of VEGF to both cell types resulted in a 370 30-50% reduction in trans-epithelial resistance (TER) within 5 h of treatment, and this was only 371 measurable following apical administration. They showed that VEGF-R2 receptors were 372 responsible for the mediation of the VEGF effect, and that these receptors were localized to the apical surface. So they conclude that VEGF initiates RPE permeability.⁵² 373 We have seen that ANG combined with VEGF is implicated in the increased permeability in the eve 374 of NPRD patients with CSME.³⁰ Confirmation comes from an *ex vivo* study performed on porcine 375 retinal endothelial cells (PREC). This study shows that ANG-2 and VEGF have synergistic effects 376 377 on the increase of permeability, the combination of both having 5 times more inducing potential than VEGF alone, which is more potent than ANG-2 in inducing permeability. They were also able 378 379 to show that the increase in permeability goes along with changes in tight junction integrity.⁵³

We have already mentioned that PPV performed on NPDR patients with ME reduces ME severity.²⁶ In a rabbit model, PPV increased VEGF clearance by 400%, after injection of human VEGF₁₆₅ in the rabbits' eyes. The authors suggest that ME improvements after PPV could be explained by the decrease in vitreous VEGF levels.⁵⁴

In two studies by Deissler *et al.* (2011 and 2013), immortalized bovine REC (iBREC) cells have been used.^{55,56} The first study wanted to test the hypothesis of whether bFGF and IGF-1 as single factors or in combination with VEGF₁₆₅ influence permeability and tight junctions and if these effects could be restored by inhibition of VEGF. An interesting result of the study was that bFGF and IGF-1 alone did not influence cell permeability measured by TER, but they had a synergistic effect with VEGF, most likely to be caused by an enhanced secretion of VEGF. The inhibition of

- VEGF by ranibizumab could completely reverse the decrease in TER. So, these results support the
 major contribution of VEGF to the change in permeability.⁵⁵
- 392 The second study investigated the effects of other members of the VEGF family, such as VEGF₁₂₁,
- 393 PIGF and viral VEGF-E, which activate different sets of VEGF receptors, on barrier function. They
- 394 strongly supported the role of VEGF-A isoforms since even in the presence of all growth factors,
- 395 TER and tight junction composition could be restored to normal in the presence of ranibizumab,
- 396 which only targets VEGF₁₆₅. They also showed that VEGFR-2, probably together with NRP-1, is
- 397 involved in the process of REC barrier impairment. Nevertheless the authors conclude that the
- involvement of other factors should not be ruled out.⁵⁶

400 Conclusion

401 Data derived from the literature support the notion that angiogenesis and inflammation are

402 interdependent processes that may interact synergistically to create the pathogenetic framework of

403 DME. Pre-clinical and translational clinical studies performed by using a combination of high-

404 throughput gene expression and proteomic technologies are needed to provide new insights into the

405 multilevel highly-regulated signaling network involved in this type of disease.

406

407 Clinical implications

408 In the present Review, we suggest the presence of a molecular crosstalk between pro-angiogenic

409 and pro-inflammatory pathways that occurs through the production of growth factors,

410 chemokines/cytokines, proteolytic enzymes, prostaglandins, and nitric oxide. It is worth

411 highlighting that among the different molecules investigated, VEGF may act as an angiogenic

412 stimulator and as a pro-inflammatory mediator and it is therefore an important link between

413 angiogenesis and the inflammatory process in this type of disease.

Based on these pieces of evidence, anti-VEGF therapies can selectively ameliorate DME symptomsand, at least partially, reverse its fundamental pathology.

References

1. Tranos PG, Wickremasinghe SS, Stangos NT, et al. Macular edema. Surv Ophthalmol 2004;49:470–90.

2. Bhagat N, Grigorian RA, Tutela A, Zarbin MA. Diabetic macular edema: pathogenesis and treatment. Surv Ophthalmol 2009;54:1–32.

3. Klaassen I, Van Noorden CJF, Schlingemann RO. Molecular basis of the inner blood-retinal barrier and its breakdown in diabetic macular edema and other pathological conditions. Prog Retin Eye Res 2013;34:19–48.

4. Brownlee M. The pathobiology of diabetic complications: a unifying mechanism. Diabetes 2005;54:1615–25.

5. Gabbay KH, Merola LO, Field RA. Sorbitol pathway: presence in nerve and cord with substrate accumulation in diabetes. Science 1966;151:209–10.

6. McLellan AC, Thornalley PJ, Benn J, Sonksen PH. Glyoxalase system in clinical diabetes mellitus and correlation with diabetic complications. Clin Sci (Lond) 1994;87:21–9.

Glenn JV, Stitt AW. The role of advanced glycation end products in retinal ageing and disease.
 Biochim Biophys Acta 2009;1790:1109–16.

8. Koya D, King GL. Protein kinase C activation and the development of diabetic complications. Diabetes 1998;47:859–66.

9. Ehrlich R, Harris A, Ciulla TA, et al. Diabetic macular oedema: physical, physiological and molecular factors contribute to this pathological process. Acta Ophthalmol 2010;88:279–91.

10. Gupta N, Mansoor S, Sharma A, et al. Diabetic retinopathy and VEGF. Open Ophthalmol J 2013;7:4–10.

11. Treins C, Giorgetti-Peraldi S, Murdaca J, Van Obberghen E. Regulation of vascular endothelial growth factor expression by advanced glycation end products. J Biol Chem 2001;276:43836–41.

12. Shimizu E, Funatsu H, Yamashita H, et al. Plasma level of interleukin-6 is an indicator for predicting diabetic macular edema. Jpn J Ophthalmol 2002;46:78–83.

13. Canning P, Glenn JV, Hsu DK, et al. Inhibition of advanced glycation and absence of galectin-3 prevent blood-retinal barrier dysfunction during short-term diabetes. Exp Diabetes Res 2007;2007:51837.

14. Satirapoj B. Review on pathophysiology and treatment of diabetic kidney disease. J Med Assoc Thai 2010;93 Suppl 6:S228–41.

15. Tufro A, Veron D. VEGF and podocytes in diabetic nephropathy. Semin Nephrol 2012;32:385–93.

16. Shimizu F, Sano Y, Tominaga O, et al. Advanced glycation end-products disrupt the bloodbrain barrier by stimulating the release of transforming growth factor- β by pericytes and vascular endothelial growth factor and matrix metalloproteinase-2 by endothelial cells in vitro. Neurobiol Aging 2013;34:1902–12.

17. Miller JW, Le Couter J, Strauss EC, Ferrara N. Vascular endothelial growth factor a in intraocular vascular disease. Ophthalmology 2013;120:106–14.

18. Ishida S, Usui T, Yamashiro K, et al. VEGF164 is proinflammatory in the diabetic retina. Invest Ophthalmol Vis Sci 2003;44:2155–62.

19. Witmer AN, Vrensen GFJM, Van Noorden CJF, Schlingemann RO. Vascular endothelial growth factors and angiogenesis in eye disease. Prog Retin Eye Res 2003;22:1–29.

20. Bremnes RM, Camps C, Sirera R. Angiogenesis in non-small cell lung cancer: the prognostic impact of neoangiogenesis and the cytokines VEGF and bFGF in tumours and blood. Lung Cancer 2006;51:143–58.

21. Shimada H, Akaza E, Yuzawa M, Kawashima M. Concentration gradient of vascular endothelial growth factor in the vitreous of eyes with diabetic macular edema. Invest Ophthalmol Vis Sci 2009;50:2953–55.

22. Patel JI, Saleh GM, Hykin PG, et al. Concentration of haemodynamic and inflammatory related cytokines in diabetic retinopathy. Eye (Lond) 2008;22:223–28.

23. Jonas JB, Neumaier M. Vascular endothelial growth factor and basic fibroblast growth factor in exudative age-related macular degeneration and diffuse diabetic macular edema. Ophthalmic Res 2007;39:139–42.

24. Funatsu H, Yamashita H, Noma H, et al. Prediction of macular edema exacerbation after phacoemulsification in patients with nonproliferative diabetic retinopathy. J Cataract Refract Surg 2002;28:1355.

25. Patel JI, Hykin PG, Cree IA. Diabetic cataract removal: postoperative progression of maculopathy--growth factor and clinical analysis. Br J Ophthalmol 2006;90:697–701.

26. Patel JI, Tombran-Tink J, Hykin PG, et al. Vitreous and aqueous concentrations of proangiogenic, antiangiogenic factors and other cytokines in diabetic retinopathy patients with macular edema: Implications for structural differences in macular profiles. Exp Eye Res 2006;82:798–806.

27. Javanmard SH, Hasanpour Z, Abbaspoor Z, et al. Aqueous concentrations of VEGF and soluble VEGF receptor-1 in diabetic retinopathy patients. J Res Med Sci 2012;17:1124–7.

28. Asato R, Kita T, Kawahara S, et al. Vitreous levels of soluble vascular endothelial growth factor receptor (VEGFR)-1 in eyes with vitreoretinal diseases. Br J Ophthalmol 2011;95:1745–48.

29. Maisonpierre PC, Suri C, Jones PF, et al. Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. Science 1997;277:55–60.

30. Patel JI, Hykin PG, Gregor ZJ, et al. Angiopoietin concentrations in diabetic retinopathy. Br J Ophthalmol 2005;89:480–83.

31. Funatsu H, Yamashita H, Ikeda T, et al. Angiotensin II and vascular endothelial growth factor in the vitreous fluid of patients with diabetic macular edema and other retinal disorders. Am J Ophthalmol 2002;133:537–43.

32. Umazume K, Usui Y, Wakabayashi Y, et al. Effects of soluble CD14 and cytokine levels on diabetic macular edema and visual acuity. Retina 2013;33:1020–5.

33. Hernández C, Ortega F, García-Ramírez M, et al. Lipopolysaccharide-binding protein and soluble CD14 in the vitreous fluid of patients with proliferative diabetic retinopathy. Retina 2010;30:345–52.

34. Sonoda S, Sakamoto T, Yamashita T, et al. Retinal morphologic changes and concentrations of cytokines in eyes with diabetic macular edema. Retina 2014;34:741–8.

35. Shukla D, Behera UC, Chakraborty S, et al. Serous macular detachment as a predictor of resolution of macular edema with intravitreal triamcinolone injection. Ophthalmic Surg Lasers Imaging 2009;40:115–9.

36. Koskela UE, Kuusisto SM, Nissinen AE, et al. High vitreous concentration of IL-6 and IL-8, but not of adhesion molecules in relation to plasma concentrations in proliferative diabetic retinopathy. Ophthalmic Res 2013;49:108–14.

37. Yoshimura T, Sonoda K, Sugahara M, et al. Comprehensive analysis of inflammatory immune mediators in vitreoretinal diseases. PloS One 2009;4:e8158.

38. Oh IK, Kim S-W, Oh J, et al. Inflammatory and angiogenic factors in the aqueous humor and the relationship to diabetic retinopathy. Curr Eye Res 2010;35:1116–27.

39. Funk M, Schmidinger G, Maar N, et al. Angiogenic and inflammatory markers in the intraocular fluid of eyes with diabetic macular edema and influence of therapy with bevacizumab. Retina 2010;30:1412–9.

40. Shimura M, Yasuda K, Nakazawa T, et al. Panretinal photocoagulation induces proinflammatory cytokines and macular thickening in high-risk proliferative diabetic retinopathy. Graefes Arch Clin Exp Ophthalmol 2009;247:1617–24.

41. Mocan MC, Kadayifcilar S, Eldem B. Elevated intravitreal interleukin-6 levels in patients with proliferative diabetic retinopathy. Can J Ophthalmol 2006;41:747–52.

42. Costagliola C, Daniele A, dell' Omo R, et al. Aqueous humor levels of vascular endothelial growth factor and adiponectin in patients with type 2 diabetes before and after intravitreal bevacizumab injection. Exp Eye Res 2013;110:50–4.

43. Suzuki Y, Nakazawa M, Suzuki K, et al. Expression profiles of cytokines and chemokines in vitreous fluid in diabetic retinopathy and central retinal vein occlusion. Jpn J Ophthalmol 2011;55:256–63.

44. Jonas JB, Jonas RA, Neumaier M, Findeisen P. Cytokine concentration in aqueous humor of eyes with diabetic macular edema. Retina 2012;32:2150–7.

45. Lee WJ, Kang MH, Seong M, Cho HY. Comparison of aqueous concentrations of angiogenic and inflammatory cytokines in diabetic macular oedema and macular oedema due to branch retinal vein occlusion. Br J Ophthalmol 2012;96:1426–30.

46. Sohn HJ, Han DH, Kim IT, et al. Changes in aqueous concentrations of various cytokines after intravitreal triamcinolone versus bevacizumab for diabetic macular edema. Am J Ophthalmol 2011;152:686–94.

47. Kawashima M, Shoji J, Nakajima M, et al. Soluble IL-6 receptor in vitreous fluid of patients with proliferative diabetic retinopathy. Jpn J Ophthalmol 2007;51:100–4.

48. Funatsu H, Yamashita H, Noma H, et al. Increased levels of vascular endothelial growth factor and interleukin-6 in the aqueous humor of diabetics with macular edema. Am J Ophthalmol 2002;133:70–7.

49. Funatsu H, Yamashita H, Ikeda T, et al. Vitreous levels of interleukin-6 and vascular endothelial growth factor are related to diabetic macular edema. Ophthalmology 2003;110:1690–6.

50. Cohen T, Nahari D, Cerem LW, et al. Interleukin 6 induces the expression of vascular endothelial growth factor. J Biol Chem 1996;271:736–41.

51. Aiello LP, Bursell SE, Clermont A, et al. Vascular endothelial growth factor-induced retinal permeability is mediated by protein kinase C in vivo and suppressed by an orally effective beta-isoform-selective inhibitor. Diabetes 1997;46:1473–80.

52. Ablonczy Z, Crosson CE. VEGF modulation of retinal pigment epithelium resistance. Exp Eye Res 2007;85:762–71.

53. Peters S, Cree IA, Alexander R, et al. Angiopoietin modulation of vascular endothelial growth factor: Effects on retinal endothelial cell permeability. Cytokine 2007;40:144–50.

54. Lee AB, Laredo J, Neville R. Embryological background of truncular venous malformation in the extracranial venous pathways as the cause of chronic cerebro spinal venous insufficiency. Int Angiol 2010;29:95–108.

55. Deissler HL, Deissler H, Lang GE. Inhibition of vascular endothelial growth factor (VEGF) is sufficient to completely restore barrier malfunction induced by growth factors in microvascular retinal endothelial cells. Br J Ophthalmol 2011;95:1151–6.

56. Deissler HL, Deissler H, Lang GK, Lang GE. VEGF but not PIGF disturbs the barrier of retinal endothelial cells. Exp Eye Res 2013;115C:162–71.

Figure legends

- Figure 1. Physiopathological mechanisms of diabetic retinopathy
- Figure 2. Angiogenesis events followed by inflammation
- Figure 3. Inflammatory events followed by angiogenesis
- Figure 4. Combined mechanisms of angiogenesis and inflammation

Tables

Supplementary Table 1. Summary overview of studies evaluating levels of inflammation and angiogenesis mediators in diabetic retinopathy patients

Study (first name and year)	Type of patients (n)	Disease stages/severity evaluated	Molecule(s) investigated	Sampling site	Main results	Conclusions of authors			
Clinical stud	Clinical studies supporting predominant role of angiogenesis on the pathogenesis of diabetic retinopathy								
Javanmard 2012 ²⁷	 NPDR (n=27) Normoglycemic controls (n=33) 	FT	• VEGF • sVEGFR-1	Aqueous humor	 No difference in VEGF aqueous levels between subjects and controls NPDR had lower sVEGFR-1 vs controls Positive correlation between VEGF/sVEGFR-1 concentration and FT 	 Decreased chelating effects of sVEGFR-1 may allow VEGF to activate the proangiogenic endothelial cell state and induce permeability Imbalance of VEGF and sVEGFR-1 may determine the fate of DME 			
Asato 2011 ²⁸	 MH (n=30) BRVO (n=37) CRVO (n=27) DME (n=42) PDR (n=51) All treated by vitrectomy 	Yes	sVEGFR-1	Vitreous humor	 sVEGFR-1 not significantly different among different eye diseases SVEGFR-1 correlated with age In active PDR, sVEGFR-1 lower vs quiescent PDR 	 High sVEGFR-1 decreases risk of angiogenesis The increase in sVEGFR-1 concentration with age might explain why PDR in youth tends to be more aggressive 			
Shimada 2009 ²¹	 DME w/o PVD nor treated by PRP (n=71) MH (n=10) 	FT	VEGF	Vitreous humor	 VEGF higher in premacular vitreous vs peripheral cortical vitreous and mid vitreous FT correlated with VEGF In controls VEGF was below detection limit 	 Diffusion of VEGF from macular region to periphery and from posterior to anterior globe VEGF is associated with presence and 			

Patel 2008 ²²	• NPDR (n=15) • PDR (n=5) • FTMH (n=5)	Yes	 ET-1 Prostacyclin NO IL-1β IL-1 Ra 	Vitreous humor	 No difference of NO and prostacyclin in different groups ET-1 lower in NPDR vs PDR and FTMH ET-1 correlated with FT and macular volume in NPDR with ME IL-1β detected in PDR Diabetics had lower IL-1 Ra 	severity of DME but this does not imply a cause/effect relationship As disease progresses with capillary loss and retinal ischemia (PDR) inflammation increases Retinal microcirculation undergoes changes even before the onset of clinical disease ET-1 inversely correlates with blood flow, high in NPDR and low in PDR
Jonas 2007 ²³	 ARMD (n=35) DME (n=21) Controls (n=24) 		• VEGF • bFGF	Aqueous humor	 VEGF and bFGF higher in diabetics vs ARMD and controls Controls and ARMD did not differ much More marked differences for VEGF 	 In DR more retina is involved and more tissue is affected In ARMD, only a small subfoveal region is affected
Patel 2006 ²⁵	 PDR/NPDR (n=7) undergoing uneventful phacoemulsification with intraocular lens implant (cataract surgery) 	Yes	 VEGF HGF IL-1β PEDF 	Aqueous humor	 VEGF(165) increased from 68pg/ml to 723pg/ml 1 day after surgery and decreased to 179pg/ml at 1 month HGF steadily increased over the month IL-1β and PEDF had acute rise on day 1 and decreased again 	 Cataract surgery causes altered concentrations of angiogenic and antiangiogenic growth factors worsening diabetic maculopathy
Patel 2006 ²⁶	 NPDR with CSME undergoing PPV (n=20) FTMH (n=8) PDR (n=22) 	Clinical assessment including OCT	 VEGF-A PEDF HGF MMP-9 sVEGFR-1 TGF-β1 	 Baseline vitreous humor Baseline aqueous humor Post operative aqueous humor 	 VEGF-A higher in NPDR vs FTMH and PDR PEDF higher in FTMH vs NPDR and PDR PEDF in NPDR higher vs PDR HGF, sVEGFR-1 and TGF-β1 differed in NPDR vs PDR and controls 	 Upregulation of VEGF in the vitreous of diabetics with a reciprocal decrease in PEDF Structural and molecular OCT macular profiles may explain

						varying response to PPV in diffuse CSME
Patel 2005 ³⁰	 NPDR and CSME (n=17) PDR (n=10) MH (n=5) All undergoing PPV 		ANG-1 and -2	Vitreous humor	 Median ANG-1 was low in MH (17pg/ml), very high in NPDR with CSME (2002pg/ml) and 186pg/ml in PDR Median ANG-2 was very high in NPDR with CSME (4000pg/ml) and undetectable in MH and PDR 	 ANG-2 is an antagonist of ANG-1 which may act as anti- permeability factor Predominance of ANG- 2 may facilitate VEGF induced retinal vascular permeability in CSME
Funatsu 2002 ²⁴	NPDR undergoing cataract surgery (n=104)	Postoperative exacerbation of ME	 VEGF IL-6 proteins 	Aqueous humor	 Hypertension, VEGF, IL-6 and protein correlated with exacerbation of ME Increase in VEGF increased ME after surgery 	 VEGF predicts risk for postoperative exacerbation of ME
Funatsu 2002 ³¹	 DME (n=20) Diabetics w/o retinopathy (n=6) Non diabetics (n=14) 	Severity assessed by fluorescence	• VEGF • All	Vitreous humor	 VEGF higher in DME vs others All higher in DME vs non diabetics All correlated with VEGF All and VEGF higher in hyperfluorescent DME vs hypofluorescent 	All and VEGF are related to the increase of vascular permeability in DME
Clinical stud	ies supporting predomina	ant role of inflamma	ation on the pathogene	sis of diabetic retinopa	ithy	
Umazume 2013 ³²	 DME (n=14) PPV non diabetic controls (n=24) All undergoing cataract surgery 	No	 sCD14 IL-8 IFN-IP-10 MCP-1 MIG VEGF 	Vitreous humor Aqueous humor Serum	 All factors elevated in vitreous of DME eyes sCD14 and VEGF in vitreous and aqueous fluids higher in DME vs controls Vitreous and aqueous levels of sCD14 correlated in DME eyes Vitreous sCD14 correlated with VEGF, IL-8, MCP-1, and preoperative visual acuity 	sCD14 may act as a key regulator of VEGF production and have a role in DR pathology
Sonoda 2014 ³⁴	T2DM with DR and undergoing PPV (n=52)	 SRD retinal cystic changes retinal swelling 	• VEGF • IL-6, -8	Vitreous humor	 IL-6 associated to SRD retinal cystic changes and retinal swellings not associated with the concentrations of intravitreal cytokines 	 IL-6 can increase the inflammatory reaction in the outer retina resulting in a further disruption of the ELM VEGF is not the only factor
Koskela 2013 ³⁶	 PDR (n=38) Non diabetic controls with macular hole or 	All patients had advanced disease	 sE-selectin sICAM-1, -3 sPECAM-3 	Vitreous humorPlasma	 IL-6 and IL-8 higher in vitreous vs plasma Vitreous IL-10, sPECAM-1, sE-selectin, sICAM-1 and sVCAM-1 higher in PDR vs 	 Local inflammation in PDR triggered by IL-6 and IL-8

Oh 2010 ³⁸	idiopathic epiretinal membrane (n=16) • All undergoing PPV • DR (n=50) • Non diabetics (n=28)	Severity of diabetic retinopathy assessed by OCT	sP-selectin sVCAM-1 IL-1β, -2, -4, -5, -6, -8, - 10, -12p70 TNF-α, -β IFN-γ IL-1β TNF-α MCP-1 IP-10	Aqueous humor	 controls Adhesion molecule concentrations in vitreous in PDR were less than 10% of those in plasma II-10 lower in vitreous vs plasma vitreous IL-10/IL-8 ratio was lower in PDR vs controls MCP-1, IP-10, IL-8, and VEGF higher in DR vs non diabetics MCP-1 and IP-10 correlated with severity of DR 	 VEGF might promote inflammation-induced damage in DR triggered by IL-6 and IL-8 Chemokines may play a role in pathogenesis of DR VEGF mediates mainly
			• IL-6, -8 • VEGF		 IL-6 correlates with macular thickness 	angiogenesis in PDR
Funk 2010 ³⁹	 DME (n=10) cataract surgery controls (n=10) 	All patients had an advanced stage of DR	 IL-4, -6, -8, -10 ICAM-1 IFN-γ MCP-1 TNF-α EGF FGF-2 PDGF-AB, -BB VEGF 	Aqueous humor	 MCP-1 and IL-8 higher in DME vs controls IL-6 and VEGF higher in DME but not significant 	 MCP-1 and IL-8 might have a role in pathogenesis of DME
Yoshimura 2009 ³⁷	 DME (n=92) PDR (n=147) BRVO (n=30) CRVO (n=13) RRD (n=63) MH or ERM as controls (n=83) 	Yes	20 soluble factors (9 cytokines, 6 chemokines, and 5 growth factors)	Vitreous humor	 IL-6, -8 and MCP-1 elevated in all groups of vitreoretinal diseases vs control VEGF elevated in PDR and CRVO 	 There may be a common pathway involved in the inflammation process in vitreoretinal diseases IL-6, IL-8 and MCP-1 promote vascular permeability causing DME Ischemia and VEGF further promote DME development to PDR
Shimura 2009 ⁴⁰	Bilateral PDR requiring PPV (n=14)	FT	• VEGF • SDF-1 • IL-6	Vitreous humor	 IL-6 and RANTES in PRP pretreated eyes were higher vs controls Macular thickness correlated with VEGF 	 PRP induced macular edema was caused by inflammation

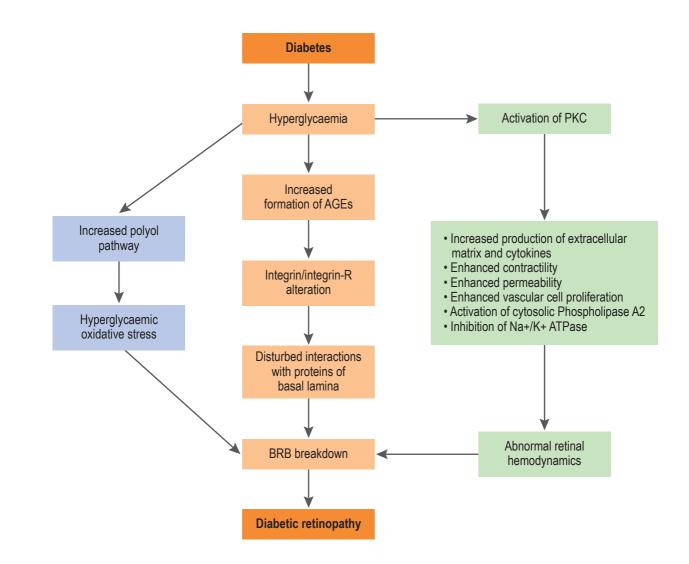
Mocan 2006 ⁴¹	 PDR (n=8) Non diabetics undergoing vitrectomy (n=8) 	With or without ME	• RANTES	Vitreous humor Serum	 and IL-6 IL-6 higher in PDR vs controls (vitreous) Serum IL-6 undetectable PDR with ME had higher IL-6 vs non ME No correlation between IL-6 and age, duration of DM or vitreous hemorrage, PRP, type of therapy, hyperglycemia, renal 	 VEGF levels could have been saturated before PRP Status of macular edema in control eyes correlated with VEGF Inflammatory cytokines may be more responsible than VEGF in causing DME IL-6 may have a role in PDR and is produced intraocularly
Shimizu 2002 ¹²	• Mild DR (n=159)	no macular edema focal edema diffuse edema cystoid edema.	 VEGF IL-6 TGF-β₁ TNF-α Lipoprotein(a) - plasma VonWillebrand factor - serum Thrombomodulin - serum 	Plasma Serum	 PRP, type of therapy, hypergivcemia, renal function Plasma IL-6 and PVD correlated with severity of ME 	IL-6 in plasma and PVD can be predictors of ME
Clinical studi	es supporting both angio	genesis and inflamr	nation causative of diabeti	retinonathy		
Costagliola 2013 ⁴²	 PDR with ME receiving bevacizumab (n=20) Non diabetics undergoing cataract surgery (n=20) 	BCVA and FT	• VEGF • APN	Aqueous humor	 APN and VEGF higher in PDR vs controls After IVBe APN and VEGF decreased significantly IVBe decreased FT and improved BCVA 	 Efficacy of anti-VEGF treatment indicates that VEGF contributes to pathogenesis of PDR anti-VEGF may not achieve neutralization of other inflammatory molecules involved in the cascade of the breakdown of BRB
Lee 2012 ⁴⁵	• DME (n=18)	• CSMT	IL-2, -5, -6, -8, -12p70, -	Aqueous	IL-8, MCP-1, PDF-AA and VEGF higher and	 Role of inflammation

	 BRVO-ME (n=12) Normal controls (n=16) 	• TMV	13 MCP-1 MIP-1α PDGF-AA TGF-α IFN-γ EGF FGF2 VEGF	humor	 IL-13 lower in DME vs control IL-8 and VEGF higher in BRVO-ME vs control IL-6 and MCP-1 higher in DME vs BRVO-ME IL-8 positively and IFN-γ negatively correlated to DME severity In BRVO-ME, IL-8 positively correlated with ME severity and retinal ischemia 	in BRVO-ME less influential than in DME Ischemic insult may be central in BRVO-ME
Jonas 2012 ⁴⁴	 Diffuse DME (n=23) Controls undergoing cataract surgery (n=22) 	RMT	 TGF-α, -β EGF FGF-β HGF IFN-α, -β, -γ IL-1a2, -1b, -2, -3, -4, -5, -6, -8, -10, -12p40, -12p70 IFN-γ-IP10 ICAM-1 MCP-1, -3 MMIF MIG MMP-1, -9 PAI-1 PIGF PDGF-BB SCDF-1 TRAIL VCAM VEGF 	Aqueous humor	 DME patients vs controls had higher EGF, HGF, ICAM-1, IL-1a2, -6, -8, IFN-γ-IP10, MCP-1, MIG, MMP-1, -9, PAI-1,PIGF, TGF-β, VCAM, VEGF RMT was associated with concentrations of EGF, ICAM-1, IL-3, -6, -8, MCP-1, MIG, MMP-9, TGF-β, PIGF, VCAM, VEGF VEGF correlated with PIGF, PAI-1, ICAM-1, MIG, MCP-1, VCAM, IL-6, -8, EGF, MMIF 	 Many cytokines correlate with DME and its severity ICAM-1 was the most associated No causality can be inferred
Suzuki 2011 ⁴³	 DR (n=76) CRVO (n=10) ERM/MH (n=23) 	No	27 different cytokines and chemokines	Vitreous humor	 In DR, IL-6, -8, -10, -13, IP-10, MCP-1, MIP- 1β, PDGF and VEGF were higher than in controls In CRVO, IL-1β, -2, -5, -8, -9, -10, -12, -13, eotaxin, G-CSF, IFN-γ, IP-10, MCP-1, MIP- 1β, TNF-α and VEGF were higher than in controls IL-2, -9, -12, MCP-1 and IFN-γ higher in 	 On top of inflammatory cytokines and neutrotrophic factors like VEGF, IL-10 and - 13 may be involved in pathogenesis of DR and CRVO

					CRVO vs DR IL-10 and -13 correlated to VEGF in DR PDGF was inversely correlated to VEGF	 Cytokines and chemokines may be correlated to VEGF in vitreous Inflammatory reaction may be more active in CRVO vs DR
Sohn 2011 ⁴⁶	 Bilateral DME (n=11, 22 eyes) Cataract surgery (n=6) DME patients received triamcinolone in 1 eye and bevacizumab in the other 	No	 IL-6, -8 IP-10 MCP-1 PDGF-AA VEGF 	Aqueous humor	 IL-8, IP-10, MCP-1, VEGF higher in DME vs control IVTA reduced FT more than IVBe IVTA reduced IL-6, IP-10, MCP-1, PDFG-AA, VEGF IVBe reduced VEGF more than IVTA 	 Pathogenesis of DME not only related to VEGF Many cytokines may be involved in DME pathogenesis
Kawashima 2007 ⁴⁷	 PDR (n=28) PPDR/DME (n=7) MH/ERM (n=10) 	Yes	• sIL-6R • VEGF	• Vitreous humor • Serum	 sIL-6R higher in PDR and PPDR vs non diabetics (vitreous) sIL-6R higher in PDR vs control (serum) sIL-6R correlated with VEGF in vitreous in PDR 	 sIL-6R can be a biomarker of PDR
Funatsu 2003 ⁴⁹	 DME w/o PVD (n=26) Non diabetic ocular disease (n=12) 	Hyperfluorescent DME (more severe)	• IL-6 • VEGF	 Vitreous humor Plasma 	 Vitreous IL-6 and VEGF higher in DME vs control IL-6 correlated with VEGF IL-6 and VEGF higher in hyperfluorescent DME vs minimally fluorescent DME 	IL-6 together and/or via VEGF may promote an increase of vascular permeability in DME subjects w/o PVD
Funatsu 2002 ⁴⁸	• DME (n=54)	ETDRS scale	• VEGF • IL-6	 Aqueous humor Plasma 	 Aqueous VEGF and IL-6 correlated with severity of DME and with aqueous protein concentration Aqueous levels of VEGF and IL-6 higher than plasma levels VEGF levels correlated with IL-6 levels Status of the posterior vitreous correlated with severity of ME 	Both VEGF and IL-6 are produced together in the intraocular tissues and are involved in the pathogenesis of ME

All, angiotensin II; ANG, angiopoietin; APN, adiponectin; ARMD, age related macular degeneration; b, basic; BCVA, best-corrected visual acuity; BRB, blood retinal barrier; BRVO, branch retinal vein occlusion; CRVO, central retinal vein occlusion; CSME, clinically significant macular edema; CSMT, central subfield macular thickness; DM, diabetes mellitus; DME, diabetic macular edema; DR, diabetic retinopathy; EGF, epidermal growth factor; ELM; external limiting membrane; ERM, idiopatic epiretinal membrane; ET, endothelin; ETDRS, Early Treatment Diabetic Retinopathy Study; FGF, fibroblast growth factor; FT, foveal

thickness; FTMH, full thickness macular hole; G-CSF, granulocyte colony-stimulating factor; HGF, human growth factor; ICAM, intercellular adhesion molecule; IFN, interferon; IL, interleukin; IP, induced protein; IVBe, intravitreal bevacizumab; IVTA, intravitreal triamcinolone acetonide; MCP, monocytochemotactic protein; ME, macular edema; MH, idiopatic macular hole; MIG, monokine induced by IFN-γ; MIP, macrophage inflammatory protein; MMIF, macrophage migration inhibitory factor; MMP, matrix metalloproteinase; n, number of subjects; NO, nitric oxide; NPDR, non proliferative diabetic retinopathy; OCT, optical coherence tomography; PAI, plasminogen activator inhibitor; PDGF, platelet-derived growth factor; PDR, proliferative diabetic retinopathy; PECAM, plateletendothelial cell adhesion molecule; PEDF, pigment epithelial derived growth factor; PIGF, placenta growth factor; PPDR, pre-proliferative diabetic retinopathy; PPV, pars plana vitrectomy; s, soluble; PRP, pan-retinal photocoagulation; PVD, posterior vitreous detachment; R, receptor; Ra, receptor antagonist; RANTES, regulated upon activation normal T-cell expressed and secreted; RMT, retinal macula thickness; RRD, rhegmatogenous retinal detachment; s, soluble; SCDF, stromal cell-derived factor; SDF, stromal derived factor; SRD, serous retinal detachments; T2DM, type 2 diabetes mellitus; TGF, transforming growth factor; TMV, total macular volume; TNF, tumor necrosis factor; TRAIL, TNF-α-related apoptosis inducing ligand; VCAM, vascular cell adhesion molecule; VEGF, vascular endothelial growth factor; vs, versus; w/o, without.



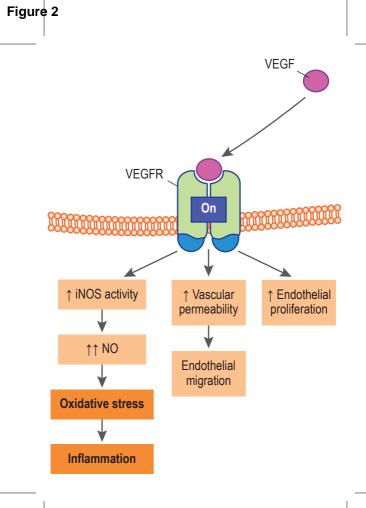


Figure 3

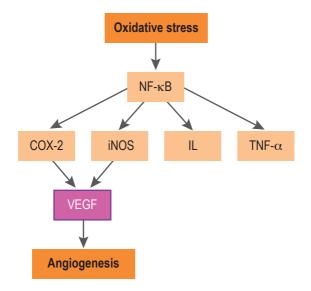
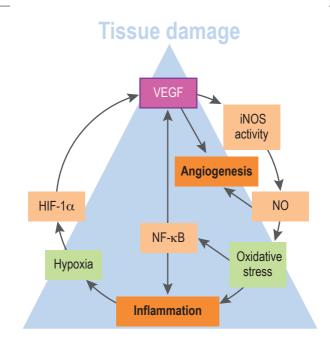


Figure 4



*Conflict of Interest Form (ICMJE COI) Click here to download Conflict of Interest Form (ICMJE COI): coi_disclosure DANESI.pdf *Conflict of Interest Form (ICMJE COI) Click here to download Conflict of Interest Form (ICMJE COI): coi_disclosure EGAN.pdf *Conflict of Interest Form (ICMJE COI) Click here to download Conflict of Interest Form (ICMJE COI): coi_disclosure Fogli.pdf *Conflict of Interest Form (ICMJE COI) Click here to download Conflict of Interest Form (ICMJE COI): coi_disclosure_MOGAVERO.pdf

Azienda Ospedaliero - Universitaria Pisana

Comitato etico di Area Vasta Nord-Ovest per la Sperimentazione Clinica

Regione 090 - Azienda 901

Pisa, 12/6/2014

To whom it may concern,

The Ethics Committee for Clinical Trials does not require submission for approval of review manuscripts, provided they do not report on original, unpublished research data on humans.

Romano Danesi Chairman, Ethics Committee

Presidenza c/o U.O. Farmacologia clinica Universitaria: tel.: 050-992632 - fax: 050-2218758 - email: romano.danesi@unipi.it Segreteria amministrativa c/o U.O. Affari Generali AOUP - Coordinatore: Sig.ra Franca Cossu (email: f.cossu@ao-pisa.toscana.it - tel.: 050-996392 - fax: 050-996293) Segreteria scientifica c/o U.O. Farmacologia clinica Universitaria AOUP - Coordinatore: Dott. Diego Carignani (email: d.carignani@ao-pisa.toscana.it - tel.: 050-993439/992262 - fax: 050-2218758)