

Crop Protection
Manuscript Draft

Manuscript Number: CROPRO-D-17-00281

Title: Occurrence of viruses and viroids in ornamental citrus mother plants in Tuscany (Central Italy)

Article Type: Short Communication

Section/Category: Crop pathogens such as fungi, oomycetes, bacteria, viruses, other microbes and nematodes

Keywords: CTV, viroids, citrus, nursery

Abstract: In Italy, Citrus tristeza closterovirus (CTV) was found several times in the last decades and Plant Protection Services (PPSs) are involved in monitoring plans and surveillance activity. Beside well-monitored orchards linked to citrus industry, the occurrence of virus or virus-like disease in ornamental nurseries may represent an under notice risk factor. A CTV monitoring program was modified with the aim to include other viruses (Citrus variegation virus, CVV; Citrus psorosis virus, CPsV) and viroids (Citrus exocortis viroid, CEVd; Hop stunt viroid, HSVd; Citrus bent leaf viroid, CBLVd; Citrus dwarfing viroid, CDVd; Citrus bark cracking viroid, CBCVd). Ornamental mother plants were monitored for four-years throughout application of cost-saving multiplex RT-qPCR protocols. CTV incidence was 1.6-13.5%, with an average distribution of 11.9%. CVV and CPsV average incidence was 6.3% and 2.7%, respectively. Higher CTV, CVV and CPsV incidences were observed in *C. x paradisi*, *C. grandis* and *C. x clementina*. The most widespread viroid is CEVd (32.9%), frequently observed in *C. x limonia* and *C. limon*. HSVd (10.5%) and CDVd (7.1%) were mostly found in *C. x limonia*. Lower infection rates were observed for CBLVd (2.0%) and CBCVd (1.4%). However, nurseries' response to PPS' virus alert seems partially effective, because CTV incidence was reduced in nurseries re-checked after first detection, but the pathogen was not eradicated from two nurseries out of three. Furthermore, occurrence of viroids was reduced in just one nursery. Concluding, multiplex diagnosis of viruses and viroids in ornamental citrus nurseries should offer useful alert to nurseries, due to repeated survey of harmful viruses and high widespread of viroids.

1 Plant health management for ornamental nurseries

2 **Occurrence of viruses and viroids in ornamental citrus mother plants in**
3 **Tuscany (Central Italy)**

4

5 Domenico Rizzo¹, Alberto Materazzi², Luciana Stefani¹, Alessandra Panattoni², Roberto Pierro²,
6 Luigi De Bellis³ and Andrea Luvisi³

7

8 1 Servizio Fitosanitario Regionale, Regione Toscana, Via Ciliegiole 98, 51100 Pistoia, Italy

9 2 Department of Agriculture, Food and Environment, University of Pisa, Via del Borghetto, 80, 56124 Pisa,
10 Italy

11 3 Department of Biological and Environmental Sciences and Technologies, University of Salento, via
12 Prov.le Monteroni, 73100 Lecce, Italy

13

14 Correspondence

15 Andrea Luvisi, Department of Biological and Environmental Sciences and Technologies, University of
16 Salento, Lecce, Italy. E-mail: andrea.luvisi@unisalento.it

17

18 **Keywords**

19 CTV, viroids, citrus, nursery

20

21 **Abstract**

22 In Italy, Citrus tristeza closterovirus (CTV) was found several times in the last decades and Plant
23 Protection Services (PPSs) are involved in monitoring plans and surveillance activity. Beside well-
24 monitored orchards linked to citrus industry, the occurrence of virus or virus-like disease in
25 ornamental nurseries may represent an under notice risk factor. A CTV monitoring program was
26 modified with the aim to include other viruses (Citrus variegation virus, CVV; Citrus psorosis virus,

27 CPsV) and viroids (Citrus exocortis viroid, CEVd; Hop stunt viroid, HSVd; Citrus bent leaf viroid,
28 CBLVd; Citrus dwarfing viroid, CDVd; Citrus bark cracking viroid, CBCVd). Ornamental mother
29 plants were monitored for four-years throughout application of cost-saving multiplex RT-qPCR
30 protocols. CTV incidence was 1.6-13.5%, with an average distribution of 11.9%. CVV and CPsV
31 average incidence was 6.3% and 2.7%, respectively. Higher CTV, CVV and CPsV incidences were
32 observed in *C. x paradisi*, *C. grandis* and *C. x clementina*. The most widespread viroid is CEVd
33 (32.9%), frequently observed in *C. x limonia* and *C. limon*. HSVd (10.5%) and CDVd (7.1%) were
34 mostly found in *C. x limonia*. Lower infection rates were observed for CBLVd (2.0%) and CBCVd
35 (1.4%). However, nurseries' response to PPS' virus alert seems partially effective, because CTV
36 incidence was reduced in nurseries re-checked after first detection, but the pathogen was not
37 eradicated from two nurseries out of three. Furthermore, occurrence of viroids was reduced in just
38 one nursery. Concluding, multiplex diagnosis of viruses and viroids in ornamental citrus nurseries
39 should offer useful alert to nurseries, due to repeated survey of harmful viruses and high widespread
40 of viroids.

41


42 **Introduction**

43 Citrus tristeza virus (CTV) is widespread throughout tropical citrus-growing areas but concerns are
44 raising in Europe, where strict quarantine measures are necessary to avoid the introduction of
45 CTV into countries where the virus is not present. Measures to control CTV damage include
46 quarantine and budwood certification programmes and elimination of infected trees (Moreno et al.,
47 2008), deeply involving Plant Protection Services (PPSs) for health check and post-diagnosis
48 procedures.

49 In Italy, where the virus was found several times in the last decades (Djelouah et al., 2009; Davino
50 et al., 2013), PPSs are involved in monitoring plants and surveillance activity. While citrus trees are
51 mainly grown in southern Italy and are related to citrus industry, several ornamental citrus plants

52 are cultivated in areas not thoroughly involved in citrus industry, such as Central or North Italy. In
53 these areas, the occurrence of citrus diseases may be under notice due to limited cultivation but
54 virus or virus-like infections of mother plants of ornamental citrus could lead to severe widespread
55 of diseases in importing countries within the European Union (EU).

56 For PPSs, the main component of costs for diagnostic molecular test (i.e. PCR, RT-PCR, qPCR) are
57 sample collection (due to staff and travel costs), sample preparation and extraction of nucleic acids.
58 These time-consuming tasks increase staff costs if carried out manually, whereas the use of
59 semiautomatic grinders or automated DNA/RNA extraction systems increase equipment costs.
60 Thus, once sample was collected and prepared, the marginal cost of multiple pathogen recognition
61 is reduced: this is the main reason of using multiplex detection protocols when available. Besides
62 CTV, other viruses cause concern in citrus cultivation, such as Citrus variegation virus (CVV) or
63 Citrus psorosis virus (CPsV) (Gonsalves and Garnsey 1975; da Graça et al. 1991; Martín et al.
64 2004; Velázquez et al., 2016) which monitoring by PPS should support nursery activities and
65 improve trust policies among stakeholders. The opportunity to investigate the presence of various
66 viruses is underlined by development of multiplex reverse transcription quantitative PCR (RT-
67 qPCR) protocols (Loconsole et al., 2010; Osman et al., 2015).

68 Citrus are also natural hosts of several viroid species (Flores et al., 2005; Ding, 2009), wh  role
69 on symptoms expression was investigated (Murcia et al., 2015). Exocortis and cachexia are severe
70 diseases caused by Citrus exocortis viroid (CEVd) and Hop stunt viroid (HSVd), respectively.
71 While viroids such as Citrus bent leaf viroid (CBLVd), Citrus dwarfing viroid (CDVd), or Citrus
72 bark cracking viroid (CBCVd) may cause little effects on fruits, the infection may reduce height and
73 canopy volume (Bani Hashemian et al., 2010; Rizza et al., 2011; Murcia et al., 2015). Significant
74 effects can be also observed on rootstocks (Polizzi et al., 1991). Commonly, their control is based
75 on preventive measures as availability of viroid-free budwood as a source of propagation material
76 follow by adequate indexing procedures (Eiras et al., 2009). Even if studies are more oriented to

77 fruit trees, these alteration may also play a significant role in ornamental citrus and many molecular
78 diagnostic techniques for viroids are available (Luigi and Faggioli, 2013; Gucek et al., 2017).

79 In this paper we investigated the impact of viruses and viroids in ornamental nursery in Tuscany
80 (Central Italy) using multiplex RT-qPCR protocols. Thus, in order to provide enhanced service to
81 farmers, the CTV monitoring program was modified with the aim to include emerging but yet not
82 regulated pathogen such as viroids and evaluate farmers' response to PPS pest alerts.

83

84 **Materials and Methods**

85


86 Leaf samples were collected in 2012-2015 from ornamental mother plants of *Citrus* spp. (19
87 species), *Fortunella* spp. (6 species), *Microcitrus* spp. (3 species), *Poncirus trifoliata* and hybrids
88 (23 *Citrus* spp. hybrids, *C. aurantifolia* x *F. margarita*, *C. x sinensis* x *P. trifoliata* x *C. x*
89 *paradisiaca*, *Citrange Morton*, *Eremocitrus glauca* x *C. x sinensis*, *F. margarita* x *C. x clementina*,
90 *M. australasica* x *F. margarita*). Plants (124 in 2012, 228 in 2013, 193 in 2014 and 169 in 2015)
91 were grown in open field conditions in 15 nurseries located in two Tuscan districts (Central Italy).

92 In each nursery, sampling was representative of each lot of grown plants.

93 Occurrence of virus and viroids was also analyzed in three nurseries in which CTV was detected. In
94 these nurseries, CTV-infected plants were destroyed within 6 months from diagnosis and farmers
95 were informed about whole health status of tested mother plants. Two year later since first PPS'
96 alert to CTV infection, different lots of mother plants were checked for viruses and viroids,
97 comparing results of the two health check.

98 Total RNAs (TNAs) from citrus tissues were extracted from 0.2 g of leaf petioles after
99 homogenization with the Mixer Mill MM 400 (Retsch, Germany), following Foissac et al. (2001).

100 TNAs were then eluted in 150 µl of RNase free water and their concentration was determined using
101 a UV-vis Spectrophotometer. RT-qPCR reactions were performed in a final volume of 25 µl on the
102 CFX96™ Real time System (Biorad). Loconsole et al. (2010) was followed for TaqMan probes,

103 primers and amplification conditions of CTV, CVV and CPsV. Viroids were assessed following
104 Wang et al.,  2009.

105

106 Results

107

108 Viruses and viroids distribution during 2012-2015 was reported in Table 1. Incidence of virus or
109 viroid infection in analyzed species/hybrids was reported in Table 2.

110 CTV, the main target of monitoring, was widespread in the monitored area. Even if CTV was not so
111 frequent during the first year of monitoring (1.6%), the pathogen was retrieved in the following
112 years, reaching more than 13% of infection rates in the third year. In 2012-2015, the virus was
113 found in more than 21% of tested plants of *C. x paradisi*, but high infection rates were also
114 observed in *C. deliciosa* (18.2%), *C. limon* (12.8%) or *C. x Limonimedica Florentina* (12.5%).
115 However, the virus was found in six species/hybrids out of more than 25 tested. Distribution of
116 CVV was 3.6-10.5% but it is more widespread among species, involving 10 different
117 species/hybrids. The virus was particularly frequent (>25% of infected plants) in *C. grandis*, *C.*
118 *bergamia* and *C. aurantifolia*. Conversely, CPsV, which infection rates in 2012-2015 were globally
119 below 5%, was limited to four species/hybrids, with a quite high frequency in *C. x clementina*
120 (17.6%).

121 Mixed infections of viruses were quite uncommon, involving just more than 8% of infected plants
122 (Table 3). However, CTV was found in mixed infection with both CVV and CPsV. Triple mixed
123 infection was not observed.

124 All investigated viroids were found during survey. CEVd was the most widespread viroids, which
125 incidence was over 30% in three different years of monitoring (average infection of 32.9%). The
126 pathogen was detected in almost all analyzed species/hybrids, with more than half of plants infected
127 for *C. limon* and *C. x limonia*. Another widespread viroid was HSVd, which presence was
128 confirmed in 10.5% of analyzed plants. This viroid, highly widespread among species, was

129 frequently found in *C. x limonia* (42.9%), *C. x Limonimedica Florentina* (37.5%) and *C. x lumia*
130 (33.3%). CDVd was found in more than 7% of analyzed plants and infections were observed in 15
131 different species/hybrids. High infection rate was observed in *C. x limonia*, *C. x lumia* and *F.*
132 *margarita* x *C. x clementina*. Distribution of CBLVd and CBCVd was lower than other viroids,
133 2.0% and 1.4%, respectively. CBLVd was observed in *C. limon* and two hybrids, while CBCVd
134 infected mainly *C. x Limonimedica Florentina*.

135 Even if single infection of viroid was the most frequent status for tested plants, mixed infections
136 were also quite common, with almost 40% of incidence (Table 3). The most frequent combinations
137 of mixed infections involved the most frequent viroids, CEVd, HSVd and CDVd. This finding was
138 underlined by the high frequency of CEVd/HSVd/CDVd mixed infections, which incidence was
139 higher than 10%.

140 In relation to species/hybrids, a worrisome health status was observed in *C. limon* which was the
141 host for all tested viruses or viroids. Furthermore, *C. medica* and *C. x sinensis* seems common hosts
142 for all viruses and most of viroids.

143 Occurrence of CTV was analyzed in nurseries subjected to repetitive health check (Table 5).
144 Nurseries' response to PPS alert seems partially effective, because CTV incidence was reduced in
145 each farm, even if the pathogen was not eradicated from two nurseries out of three. A similar
146 behavior in nursery response was observed for CVV and CPsV (Table 5). Conversely, occurrence
147 of viroids was lower for in one nursery (N1) while increase in the other two farms.

148

149 **Discussion**

150

151 In Spain, citrus cultivars free of virus and virus-like pathogens became available thanks to
152 implementation of a citrus improvement program, and they cover today more than 70% of the
153 citrus-growing regions of the country (Bani Hasheimian et al., 2010), confirming the role of
154 quarantine, certification programmes and elimination of infected trees in protecting citrus

155 production. Based on the current legislation, the finding of CTV in a nursery would implicate the
156 mandatory destruction of the entire lot of plants. Thus, repetitive findings of CTV during 2012-
157 2015 underline a worrisome health status of ornamental citrus mother plants in Tuscany, even if
158 CTV was less widespread than in other producing regions (Abbas et al., 2015). Our observation
159 indicates as CTV incidence was reduced in nurseries re-checked two years after first detection, but
160 the pathogen was not eradicated from two nurseries out of three. Furthermore, the health status
161 seems threaten not only by CTV, but by two further viruses and a plethora of viroids. Conversely
162 from viruses, which may be strongly regulated (as CTV) or which diseases are well known by
163 nurseryman, viroids may represents an unnoticed - but harmful - menace.

164 High widespread of citrus viroids was previously assessed in commercial fruit trees in Greece
165 (Barbarossa et al., 2007), Uruguay (Pagliano et al., 2013) and low performing orchards with high
166 incidence of viroids were also observed in Spain (Bani Hasheimian et al., 2010). However, few data
167 is available for ornamental plants. The pospiviroid status of ornamental plants was assessed in Italy
168 for solanaceous ornamental genera, but beside high incidence of Potato spindle tuber viroid
169 (PSTVd), their incidence was low (Luigi et al., 2011). In Tuscany, CEVd and HSVd occurred in 45
170 and 9% of plants in citrus orchards during a survey carried out about ten years ago (Ragozzino et
171 al., 2005), while higher incidence was absorbed in the close region of Lazio (Central Italy)
172 (Ragozzino et al., 2005).

173 Our findings indicate as viroids are commonly widespread in Tuscan ornamental citrus tree
174 nurseries, reaching almost 50% of incidence for CEVd. Furthermore, our findings indicate high
175 frequency of mixed infections of viroids in ornamental citrus trees. Interactions among viroids co-
176 infecting the same tree could affect symptom expression and field performance (Vernière et al.,
177 2006). However, long-term field assays also revealed that viroid-induced effects might be
178 attenuated or enhanced when trees were exposed to mixed viroid infections (Vernière et al., 2006,
179 Vidalakis et al., 2010). Since the primary mode of transmission of most viroids is through
180 mechanical means, consideration must be given to the potential spread of the viroid by equipment

181 used in farm operations (Barbosa et al., 2005; Eastwell and Nelson, 2007). As reviewed by
182 Kovalskaya and Hammond (2014) current effective control methods for viroid diseases include
183 detection and eradication, and cultural controls, as well as CTV protection programme.

184 Concluding, multiplex diagnosis of virus and viroids in Tuscan ornamental citrus nurseries should
185 offer useful alert to nurseries (not just in relation to quarantine pests) due to repeated survey of
186 harmful viruses and high widespread of viroids.

187

188 **Conflict of Interest**

189

190 The authors declare that the research was conducted in the absence of any commercial or financial
191 relationships that could be construed as a potential conflict of interest.

192

193 **References**

194

195 Abbas M, Khan MM, Mughal SM, Ji P. (2015). Comparison of infection of Citrus tristeza
196 closterovirus in Kinnow mandarin (*Citrus reticulata*) and Mosambi sweet orange (*Citrus sinensis*)
197 in Pakistan. *Crop Prot.* 78:146-150

198

199 Bani Hashemian SM, Murcia N, Trenor I, Duran-Vila N. (2010) Low performance of citrus trees
200 grafted on Carrizo citrange is associated with viroid infection. *J Plant Pathol* 92:511-517.

201

202 Barbosa CJ, Pina JA, Pérez-Panadés J, Bernad L, Serra P, Navarro L, Duran-Vila N. (2005)
203 Mechanical transmission of citrus viroids. *Plant Dis* 89:749-754.

204

205 Barbarossa L, Loconsole G, Vovlas C. (2007) Virus and virus-like diseases of citrus in Epirus. *J*
206 *Plant Pathol* 89:273-276.

207

208 da Graça JV, Lee RF, Moreno P, Civerolo EL, Derrick KS. (1991) Comparison of isolates of Citrus
209 ringspot, psorosis, and other viruslike agents of citrus. *Plant Dis* 75:613–616.

210

211 Davino S, Willemsen A, Panno S, Davino M, Catara A, Elena SF, Rubio L. (2013) Emergence and
212 Phylodynamics of Citrus tristeza virus in Sicily, Italy. *Plos ONE* 8(6) e66700.

213

214 Ding B. (2009) The biology of viroid–host interactions. *Ann Rev Phytopathol* 47:105–131.

215

216 Djelouah K, Valentini F, D'Onghia AM. (2009) Historical review of Citrus tristeza virus in Italy. In:
217 D'Onghia AM, Djelouah K, Roistacher CN (eds) Citrus tristeza virus and Toxoptera citricidus:
218 a serious threat to the Mediterranean citrus industry. Bari, CIHEAM, pp 59-62.

219

220 Eastwell KC, Nelson ME. (2007) Occurrence of viroids in commercial hop (*Humulus lupulus* L.)
221 production areas of Washington State. Internet resource:
222 <https://www.plantmanagementnetwork.org/pub/php/research/2007/hop/> (verified January 13, 2017)

223

224 Eiras M, Silva SR, Stuchi ES, Targon MLPN, Carvalho SA. (2009) Viroids in Citrus. *Trop Plant*
225 *Pathol* 34:275-296.

226

227 Flores R, Hernández C, Martínez de Alba AE, Darós JA, Di Serio F. (2005) Viroids and viroid–host
228 interactions. *Annu Rev Phytopathol* 43:117–139.

229

230 Foissac X, Svanella-Dumas L, Gentit P, Dulucq MJ, Marais A, Candresse T. (2001) Polyvalent
231 detection of fruit tree tricho, capillo and foveaviruses by nested RTPCR using degenerated and
232 inosine containing primers (PDO RT-PCR). *Acta Hort* 550:37–40.

233

234 Gonsalves D, Garnsey SM. (1975) Functional equivalence of an RNA component and coat protein
235 for infectivity of Citrus leaf rugose virus. *Virology* 1:23–31.

236

237 Gucek T, Trdan S, Jakse J, Javornik B, Matousek J, Radisek S. (2017). Diagnostic techniques for
238 viroids. *Plant Pathol.* 66:339-358.

239

240 Kovalskaya N, Hammond RW. (2014) Molecular biology of viroid–host interactions and disease
241 control strategies. *Plant Sci.* 228:48-60.

242

243 Loconsole G, Saponari M, Savino V. (2010) Development of real-time PCR based assays for
244 simultaneous and improved detection of citrus viruses. *Eur J Plant Pathol* 128:251–259.

245

246 Luigi M, Luison D, Tomassoli L, Faggioli F. (2011) Natural spread and molecular analysis of
247 pospiviroids infecting ornamentals in Italy. *J Plant Pathol* 93:491-495.

248

249 Luigi M, Faggioli F. (2013) Development of a quantitative real-time RT-PCR (qRT-PCR) for the
250 detection of hop stunt viroid. *Eur J Plant Pathol* 137:231-235.

251

252 Martín S, Alioto D, Milne RG, Garnsey SM, García ML, Grau O, Guerri J, Moreno P. (2004)
253 Detection of Citrus psorosis virus by ELISA, molecular hybridization, RT-PCR and immunosorbent
254 electron microscopy and its association with citrus psorosis disease. *Eur J Plant Pathol* 110:747–
255 757.

256

257 Moreno, P., Ambrós, S., Albiach-Martí, M.R., Guerri, J., Peña, L., 2008. Citrus tristeza virus: A
258 pathogen that changed the course of the citrus industry. *Mol Plant Pathol* 9:251-268

259

260 Murcia N, Bani Hashemian SM, Serra P. (2015) Citrus Viroids: symptom expression and
261 performance of Washington Navel sweet orange trees grafted on Carrizo citrange. *Plant Dis*
262 99:125-136.

263

264 Osman F, Hodzic E, Kwon SJ, Wang J, Vidalakis G. (2015) Development and validation of a
265 multiplex reverse transcription quantitative PCR (RT-qPCR) assay for the rapid detection of Citrus
266 tristeza virus, Citrus psorosis virus, and Citrus leaf blotch virus. *J Virol Methods* 220:64-75.

267

268 Pagliano G, Umaña R, Pritsch C, Rivas F, Duran-Vila N. (2013) Occurrence, prevalence and
269 distribution of Citrus viroids in Uruguay. *J Plant Pathol* 95:631-635.

270

271 Polizzi G, Albanese G, Azzaro A, Davino M, Catara A. (1991) Field evaluation of dwarfing effect
272 of two combinations of citrus viroids on different citrus species. In: Brlansky RH, Lee RF, Timmer
273 LW (eds) Proceedings of the 12th Conference of the International Organization of Citrus
274 Virologists (IOCV), Riverside, CA, USA, pp 230-233.

275

276 Ragozzino E, Faggioli F, Barba M. (2005). Distribution of citrus exocortis viroid and hop stunt
277 viroid in citrus orchards of central Italy as revealed by one-tube one-step RT-PCR. *Phytopathol.*
278 *Mediterr.* 44:322–326

279

280 Rizza S, La Rosa R, Tessitori M, Albanese G, Catara A. (2011) Symptom expression and growth of
281 citrus varieties on Citrange rootstocks inoculated with different Citrus viroids. *J Plant Pathol* 93:53.

282

283 Velázquez K, Alba L, Zarza O, Vives MC, Pina JA, Juárez J, Navarro L, Moreno P, Guerri J.
284 (2016) The response of different genotypes of citrus and relatives to Citrus psorosis virus
285 inoculation. *Eur J Plant Pathol* 144:73-81.
286

287 Vernière C, Perrier X, Dubois C, Dubois A, Botella L, Chabrier C, Bové JM, Duran-Vila N. (2004)
288 Citrus viroids: symptom expression and effect on vegetative growth and yield of clementine trees
289 grafted on trifoliolate orange. *Plant Dis* 88:1189-1197.
290

291 Vidalakis G, Pagliaccia D, Bash JA, Semancik JS. (2010) Effects of mixtures of citrus viroids as
292 transmissible small nuclear RNA on tree dwarfing and commercial scion performance on Carrizo
293 citrange rootstock. *Ann Appl Biol* 157:415-423.
294

295 Wang X, Zhou C, Tang K, Zhou Y, Li Z. (2009) A rapid one-step multiplex RT-PCR assay for the
296 simultaneous detection of five citrus viroids in China. *Eur J Plant Pathol* 124:175–180.

Highlights (for review)

- CTV infections were repeatedly observed in ornamental nurseries
- CTV eradication was not always accomplished in 2 years of monitoring
- High virus incidences were observed in *C. x paradisi*, *C. grandis* and *C. x clementina*
- The most widespread viroid is CEVd frequently observed in *C. x limonia* and *C. limon.*

Table 1 Distribution of viruses and viroids in Tuscan nurseries during four years of monitoring. Infected samples out of analyzed samples are reported.

Viruses	I	II	III	IV	Total
CTV	2/124	14/228	63/193	6/169	85/714
CVV	7/124	24/228	7/193	7/169	45/714
CPsV	0/124	10/228	6/193	3/169	19/714
Viroids	I	II	III	IV	Total
CEVd	6/124	84/228	89/193	56/169	235/714
HSVd	5/124	34/228	36/193	0/169	75/714
CBCVd	0/124	10/228	0/193	0/169	10/714
CBLVd	0/124	3/228	11/193	0/169	14/714
CDVd	5/124	24/228	22/193	0/169	51/714

Table 2 Incidence (%) of Citrus tristeza closterovirus (CTV), Citrus variegation virus (CVV), Citrus psorosis virus (CPsV), Citrus exocortis viroid (CEVd), Hop stunt viroid (HSVd), Citrus bent leaf viroid (CBLVd), Citrus dwarfing viroid (CDVd) and Citrus bark cracking viroid (CBCVd) in mother plants of ornamental citrus, according to species (consistency of three or more plants).

Mother plant	CTV	CVV	CPsV	CEVd	HSVd	CBCVd	CBLVd	CDVd
<i>C. aurantifolia</i>	-	28.6	-	28.6	14.3	-	-	28.6
<i>C. aurantifolia</i> x <i>F. margarita</i>	-	-	-	-	-	-	-	-
<i>C. bergamia</i>	-	33.3	-	33.3	-	-	-	-
<i>C. deliciosa</i>	18.2	-	-	9.1	13.6	-	-	4.5
<i>C. deliciosa</i> x <i>C. x paradisi</i>	-	-	-	5.3	-	-	-	-
<i>C. grandis</i>	-	50.0	-	25.0	-	-	-	-
<i>C. hystrix</i>	-	-	-	-	-	-	-	-
<i>C. limetta</i>	-	-	-	28.6	14.3	-	-	14.3
<i>C. limon</i>	12.8	9.0	5.6	56.4	17.0	4.3	6.9	11.2
<i>C. macrophylla</i>	-	-	-	25.0	-	-	-	12.5
<i>C. medica</i>	2.5	15.0	2.5	37.5	17.5	-	-	12.5
<i>C. mitis</i>	-	-	-	15.4	-	-	-	-
<i>C. myrtifolia</i>	-	14.3	-	14.3	-	-	-	-
<i>C. unshiu</i>	-	-	-	-	-	-	-	-
<i>C. volkameriana</i>	-	12.5	-	-	12.5	-	-	-
<i>C. x aurantium</i>	-	-	-	33.3	5.6	-	5.6	-
<i>C. x clementina</i>	-	-	17.6	29.4	11.8	-	5.9	11.8
<i>C. x limonia</i>	-	-	-	57.1	42.9	-	-	57.1
<i>C. x Limonimedica Florentina</i>	12.5	-	-	25.0	37.5	12.5	-	-
<i>C. x lumia</i>	-	-	-	33.3	33.3	-	-	33.3
<i>C. x paradisi</i>	21.7	13.0	-	17.4	13.0	-	-	8.7
<i>C. x paradisi</i> x <i>C. x sinensis</i>	-	-	-	-	-	-	-	-
<i>C. x meyeri</i>	-	-	-	20.0	-	-	-	-
<i>C. x sinensis</i>	8.2	4.1	1.4	19.2	9.6	-	-	8.2
<i>F. margarita</i>	-	20.0	-	20.0	-	-	-	20.0

<i>F. margarita</i>	x	<i>C. x</i>	-	-	-	-	-	-	-	33.3
<i>clementina</i>										
<i>M. australasica</i>			-	-	-	-	25.0	-	-	25.0
Others			-	-	-	0.2	0.3	-	-	0.1

Table 3 Incidence (%) of single or mixed infection of virus.

Infection	Incidence (%)
Single infection	91.67
Mixed infection – 2 viruses	8.33
CVV/CPsV	3.13
CTV/CPsV	3.13
CTV/CVV	2.08

Table 4 Incidence (%) of single or mixed infection of viroids.

Infection	Incidence (%)
Single infection	61.86
Mixed infection – 2 viroids	21.40
CEVd/HSVd	11.16
CEVd/CDVd	4.65
CEVd/CBLVd	2.33
HSVd/CDVd	1.86
CBLVd/CDVd	0.47
HSVd/CBLVd	0.47
CEVd/CBCVd	0.47
Mixed infection – 3 viroids	14.88
CEVd/HSVd/CDVd	10.23
CEVd/CBLVd/CDVd	1.86
CEVd/HSVd/CBCVd	1.86
CEVd/HSVd/CBLVd	0.93
Mixed infection – 4 viroids	1.86
CEVd/HSVd/CBLVd/CDVd	1.40
CEVd/CBLVd/CBCVd/CDVd	0.47

Table 5 Occurrence of viruses and viroids in nurseries (N1, N2, N3) re-checked two years after first detection of CTV. T1 = infected samples out of analyzed samples observed during first detection of CTV; T2 = infected samples out of analyzed samples two years after CTV detection.

	N1		N2		N3	
	T1	T2	T1	T2	T1	T2
Viruses						
CTV	12/72	3/38	2/48	3/73	10/10	0/5
CVV	2/72	0/38	4/48	3/73	0/10	1/5
CPsV	3/72	0/38	0/48	1/73	2/10	0/5
Viroids						
CEVd	52/72	2/38	0/48	30/73	5/10	5/5
HSVd	26/72	0/38	1/48	0/73	0/10	0/5
CBCVd	0/72	0/38	0/48	0/73	1/10	0/5
CBLVd	7/72	0/38	0/48	0/73	0/10	2/5
CDVd	10/72	0/38	0/48	0/73	0/10	3/5