

Accepted Manuscript

Phytoremediated marine sediments as suitable peat-free growing media for production of Red Robin photinia (*Photinia x fraseri*)



Paola Mattei, Alessandro Gnesini, Cristina Gonnelli, Chiara Marraccini, Grazia Masciandaro, Cristina Macci, Serena Doni, Renato Iannelli, Stefano Lucchetti, Francesco P. Nicese, Giancarlo Renella

PII: S0045-6535(18)30384-9

DOI: 10.1016/j.chemosphere.2018.02.172

Reference: CHEM 20930

To appear in: *Chemosphere*

Received Date: 27 October 2017

Revised Date: 22 December 2017

Accepted Date: 26 February 2018

Please cite this article as: Paola Mattei, Alessandro Gnesini, Cristina Gonnelli, Chiara Marraccini, Grazia Masciandaro, Cristina Macci, Serena Doni, Renato Iannelli, Stefano Lucchetti, Francesco P. Nicese, Giancarlo Renella, Phytoremediated marine sediments as suitable peat-free growing media for production of Red Robin photinia (*Photinia x fraseri*), *Chemosphere* (2018), doi: 10.1016/j.chemosphere.2018.02.172

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1 **Phytoremediated marine sediments as suitable peat-free growing media for production of Red**
2 **Robin photinia (*Photinia x fraseri*)**

3 Paola Mattei¹, Alessandro Gnesini¹, Cristina Gonnelli², Chiara Marraccini², Grazia Masciandaro³,
4 Cristina Macci³, Serena Doni³, Renato Iannelli⁴, Stefano Lucchetti⁵, Francesco P. Nicese¹,
5 Giancarlo Renella¹

6 ¹Department of Agrifood Production and Environmental Sciences, University of Florence, P.le delle
7 Cascine 18, 50144 – Florence, Italy

8 ²Department of Biology, University of Florence, v. Micheli 1, 50121 – Florence, Italy

9 ³National Council of Research, Institute for Ecosystems Study (CNR-ISE), via. Moruzzi 1, – 56124
10 Pisa, Italy.

11 ⁴University of Pisa, Department of Energy, Systems, Territory and Construction Engineering, v.
12 Gabba 22, – 56122, Pisa, Italy

13 ⁵Agri Vivai s.r.l., v. Casalina, 118G, 51100 – Pistoia, Italy

14

15

16

17

18

19

20

21

22 **Corresponding author:**

23 Giancarlo Renella, Department of Agrifood Production and Environmental Sciences, University of
24 Florence, P.le delle Cascine 18, 50144 – Florence, Italy, e-mail Giancarlo.renella@unifi.it

25

26 **Abstract**

27 Sediments dredged by an industrial port, slightly contaminated by heavy metals and petroleum
28 hydrocarbons, were phytoremediated and used as peat-free growing media for the red robin photinia
29 (*Photinia x fraseri* L.). Plants were grown on sediment only (S), sediment mixed with composted
30 pruning residues (S+PR), sediment fertilized with controlled release fertilizers (S+F) and peat-based
31 growing media as control (C). Plant elongation and dry weight, leaf contents of chlorophyll,
32 malondialdehyde (MDA), macronutrients and heavy metals were determined at the end of one
33 growing season. Environmental impact related to the use of sediment-based as compared to peat-
34 based growing media was assessed by the Life Cycle Analysis (LCA). Sediment-based growing
35 media presented significantly higher bulk density, pH and electrical conductivity values, lower C
36 and N contents, and significantly higher total and available P. Red robin photinia grown on S+F
37 growing media showed morphological and chemical parameters similar to those of control plants
38 (C), whereas plants grown on S and S+PR showed lower growth. Leaf concentration of nutrients
39 and heavy metals varied depending on the considered element and growing media, but were all
40 within the common values for ornamental plants, whereas the highest MDA concentrations were
41 found in plants grown on traditional growing media. The LCA indicated the use of sediments as
42 growing media reduced the C footprint of ornamental plant production and the contribute of
43 growing media to the environmental impact per produced plant. We concluded that sediments
44 phytoremediation and use in plant nursery is a practical alternative re-use option for dredged
45 sediments.

46

47

48 **Keywords:** Dredged sediments; Phytoremediation; Ornamental Plant; Peat; LCA; Sustainable plant
49 production

50

51 **Introduction**

52 High grade ornamental plant production in containers requires growing media that generally contain
53 peat and palm fiber and coir pith, mixed with pumice or perlite to improve porosity and stability,
54 and fertilized to reach adequate levels of nutrients. It was estimated that in the European Union
55 (EU) about 40 million m³ of growing media are produced every year (Altmann et al., 2008), and
56 peat is currently still the prime raw material for professional growing media (Schmilewski 2009;
57 Ceglie et al., 2015). Palm coir fiber and pith are by-products of the coconut processing that are
58 increasingly used as ingredients of growing media thank to its suitable pH value (between 5 and 7),
59 good physical stability and water retention properties, and is widely available on the market. In
60 recent years increasing conflicts of peat extraction and use with EU environmental Directives such
61 as Environmental Impact Assessment Directive (92/43/EC), Habitats Directive (Natura 2000), Birds
62 Directive (Directive 2009/147/EC), are emerging. This urges the plant nursery sector to search for
63 alternative peat-free growing media and soil improvers. Also pumice and perlite, other key
64 ingredient of peat-based growing media, are non-renewable resource and high impact materials,
65 respectively, and their use must be progressively reduced in the future. Palm coir fiber- and pith-
66 based growing media also have high environmental impact related to high initial salinity and
67 transportation, as they produced mainly along the South-East India and Sri-Lanka coasts, and
68 require long shipment distances for EU producers.

69 Regardless of the large research efforts to reduce peat in growing media (e.g. Abad et al., 2001;
70 Garcia Gomez et al., 2002), by replacing them with various plant residues and organic wastes
71 (Garcia Gomez et al., 2002; Hernandez Apaolaza et al., 2005), the materials tested so far have
72 shown limited potentials mainly due to variability of the original materials, adverse physico-
73 chemical properties and sanitary problems, particularly in the use of composted biosolids (Abad et
74 al., 2001; De Lucia et al., 2013).

75 Sediments from the coastal, riverine and lacustrine environments of developed Countries are
76 regularly dredged to guarantee the free navigation and docking activities, to prevent flooding
77 events, and control water contamination caused by the release of pollutant accumulated into bottom
78 sediments. Management of dredged sediments is a major environmental issue in worldwide, as they
79 are generally considered as waste materials, although the major international Conventions (e.g.
80 Barcelona Convention, 1995) have suggested to reuse the dredged sediments to the maximum
81 extent. The amount of dredged sediments in Western Europe is in the order of 100-200 million
82 tonnes per year (Bortone et al., 2004). In Europe, currently none of the EU environmental
83 Directives deal specifically with dredged material, but three main Directives have a direct or
84 indirect impact on sediment management (Figure 1): the Water Framework Directive (2000/60/EC),
85 the Waste Framework Directive (75/442/EEC, 91/156/EEC), and the Habitat Directive
86 (92/43/EEC). The recent changes in the EU and national legislation on sediment management, have
87 progressively supported the re-use of unpolluted or remediation and re-use of dredged sediments. In
88 fact, Europe must face large-scale remediation of sediments in many river basins impacted by
89 historical contamination, and there is growing concern to fulfill the European Water Framework
90 Directive (SedNet, 2003).

91 Ornamental plant production is among the most remunerative agricultural sectors, with an estimate
92 annual gross production in the EU of ca. 20 millions of Euros, and a slightly increasing demand in
93 the last 20 years (European Commission, 2015). Most ornamental plants are produced in containers,
94 and demand for growing media will also likely increase in the future. Therefore, the use of
95 alternative peat-free growing media, constituted by locally available low impact materials, is an
96 interesting option to prevent limitations and losses of value for the production of ornamental plants.
97 In a previous study, we showed that phytoremediation conducted using *Paspalum vaginatum*,
98 *Spartium junceum* and *Tamarix gallica* could be a technique for reclaiming dredged marine
99 sediments and allow them for a safe use as growing media for several ornamental plants (Mattei et

100 al., 2017). However, while the phytoremediation degraded the total petroleum-derived
101 hydrocarbons by more than 50%, the total heavy metal concentrations were not significantly
102 decreased (Masciandaro et al., 2014), but stabilized into recalcitrant forms (Doni et al., 2015). We
103 hypothesized that phytoremediated sediments, although containing significantly higher
104 concentrations of heavy metals than peat, could be optimized to allow the production of ornamental
105 plants of comparable grade as plants grown on traditional peat-based growing media, and reduce the
106 environmental impact of plant nursery production, with no increase of metal mobility during the
107 plant growth. We tested our hypothesis growing 'Red Robin' photinia plants (*Photinia x Fraseri*), a
108 plant of prime interest for the Italian ornamental plant market, on either phytoremediated sediments
109 or traditional peat-based growing media. The environmental impact of the photinia production on
110 sediment- or peat-based growing media was evaluated by the Life Cycle Analysis (LCA). The LCA
111 is a widely accepted procedure for assessing the impact of productive process involving various
112 activities, and is considered a key decision support tool, increasingly used for assessing the
113 environmental performances of products throughout their whole life cycle, identifying differences
114 among different systems in terms of resource consumption and environmental impacts (Cellura et
115 al., 2012). Recent applications of the LCA to the plant nursery sector has allowed to assess the
116 relatively high impact of peat-based growing media on the production process of ornamental plants
117 (Lazzerini et al., 2016).

118

119 **Materials and methods**

120 2.1. Sediment properties and experimental set up

121 The used dredged sediments were phytoremediated in a confined facility of the port of Livorno
122 (Central Italy, 43_3302500 N, 10_1703900 E), as described by Masciandaro et al. (2014). Sediment
123 phytoremediation was conducted using swamp couch (*P. vaginatum*) Sw. alone or in combination
124 with Spanish broom (*S. junceum* L.) or French tamarisk *T. gallica*, for 2 years. Phytoremediation

125 improved the physical, chemical and biological fertility of the sediments, but left higher
126 concentrations of heavy metals and total petroleum hydrocarbons as compared to the traditional
127 peat-based growing media (Table 1), although the heavy metals were stabilized in more recalcitrant
128 forms as compared to those of the untreated sediments (Doni et al., 2015).

129 In March 2012, 100 kg of remediated sediments were transported to the Center for Experimental
130 Plants Nursery (Ce.Spe.Vi.) in Pistoia (Tuscany, Central Italy), manually homogenized, and used
131 for growing one-year-old vegetatively propagated plants of the evergreen shrub red robin photinia
132 (*Photinia x fraseri*). Plantlets were transplanted into 2-L pots containing remediated sediment only
133 (S), remediated sediment and composted pruning residues (S+PR) mixed at 1:1 v:v, remediated
134 sediment added with a controlled release fertilizer (S+F) (Osmocote, Everris) at rate of 4.5 g per
135 plant (corresponding to 0.7 g of N), or peat and pumice mixture mixed at 1:1 fertilized with
136 Osmocote at the same rate as above, representing the control treatment (C). Plants were manually
137 watered and nursery managed for the whole growth period. On September 2012, the plants were
138 transplanted into 2l pots, 8 replicates for each type of substrate. Therefore, the experimental set-up
139 shared a total of 32 plants, arranged in a completely randomized design. The experiment was
140 carried out in the Center for Experimental Nursery (CeSpeVi), Pistoia (Tuscany). From September
141 until the end of October 2012, the plants grew in plain-air, irrigated twice per day for 20 minutes by
142 a sprinkler irrigation system with water only. At the end of October 2012, all plants were transferred
143 into not-heated glass greenhouse, where they were manually watered until March 2013.

144

145 2.2. Sediments physico-chemical properties

146 Sediment texture of 39% sand, 45% silt, and 16% clay, Bulk density of the growing media was
147 determined by using cylinders of 100 cm³ filled with the different growing media. The initial dry
148 weight of the growing media was recorded, then all growing media were saturated with water. The
149 saturated samples were and completely dried at 105°C for 24 h, the dry weight was recorded, and

150 the bulk density was calculated as: bulk density ($\text{kg}\cdot\text{m}^{-3}$) = dry weight (kg)/volume (m^3). Total
151 organic C and N in sediments were determined by dry combustion using a NA 1500 CHNS
152 Analyzer (Carlo Erba, Milan, Italy). Inorganic C (IC) was determined as carbonate to Santi et al.
153 (2006). The sediment pH value was measured in 1:2.5 (w:v) aqueous suspensions by pHmeter (GLP
154 22 CRISON, Spain), after 30 min shaking followed by 5 min settling, and the electrical
155 conductivity (EC) was measured by a conductivity meter (COND400, Eutech Instruments, USA) on
156 the same extracts. Pseudototal concentration of Ca, Mg, Cd, Cr, Cu, Ni, Pb and Zn were determined
157 by extraction in aqua regia (McGrath and Cunliffe, 1985). The elemental availability was
158 determined by extractions with 1M NH_4NO_3 using the protocol described by Renella et al. (2004),
159 on both original phytoremediated sediments and peat, and on sediment- and peat-based growing
160 media at the end of the plant growth cycle. Elemental concentrations were measured by flame
161 atomic absorption (Perkin Elmer AAnalyst 200).

162

163 2.3 Measurement of plant growth and stress, chlorophyll content and element concentration

164 Plant growth parameters were determined at the beginning and at the end of the growth period.
165 Plant biomass was determined at the end of the experiment by the difference between the plants
166 fresh weight and weight of plants after drying at 60°C . Chlorophyll A and B concentration was
167 quantified analyzing 2 leaves from each plant per substrate type according to (Jeffrey e Humphrey,
168 1975). Leaves were manually grinded in a mortar in the presence of 10 ml of 90% acetone, and the
169 extract was collected in polypropilene tubes and stored in the fridge for 24 h. Afterward, the
170 extracts were centrifuged for 5 min a $4000 \times g$ and 1ml of supernatant was analyzed using a
171 spectrophotometer (Lambda 35 PerkinElmer) at 664 and 647 nm (Taiti et al., 2016). Absorbance at
172 and 750 nm was also measured to correct the values for eventual turbidity. Absorbance values were
173 used for quantification of chlorophyll A and B, respectively, according to (Jeffrey e Humphrey,
174 1975). Concentration of malondialdheyde (MDA) was determined by the reaction with 2-

175 thiobarbituric acid (TBA), and the concentration of the MDA-TBA was measured by
176 spectrophotometry at 532 nm (Pignattelli et al., 2012).

177 Concentration of Cd, Cr, Cu, Ni, Pb, Zn, Mg and Ca in plants were determined on 8 leaves for each
178 plant and type of growing media. Leaves were dried and milled, and 100 mg were transferred in 25
179 ml beaker for mineralization with 65% HNO₃ and 60% HClO₄, and subjected to temperature
180 increase from 100 to 300°C. After mineralization, all extracts were diluted to 10 ml with deionized
181 water and elemental concentrations were quantified by atomic absorption spectrophotometry
182 (AAAnalyst 200, PerkinElmer).

183

184 2.4 LCA analysis

185 The system boundary was defined by the carbon footprint emissions associated with the production
186 of photinia plants using a 'cradle to farm-gate' approach (Shine et al., 2005). The system included
187 emissions associated with the following processes: phytoremediation and transport of marine
188 sediments (1), production and transportation of plastic and peat yearly used (2), production and
189 application of fertilizers (3), production of crop protection chemicals (4), electricity and fuel used
190 for nursery operations such as fertilizer application, irrigation, plants transport within the nursery)
191 (5). The complete system diagram of an LCA analysis of sediment is reported in Figure 1. The LCA
192 was mainly focused on the impact of the materials used for the growing media. Impact associated to
193 the farm infrastructure (e.g. buildings, greenhouses, irrigation and fertigation systems) and to
194 sediment dredging were not considered, because the plants were all grown in the same farm and the
195 sediment dredging was conducted independently from its subsequent use of as growing media. The
196 Global Warming Potential (GWP) expressed as kg of CO₂ equivalents (kg CO₂ eq.) was used as
197 impact category, defined as the cumulative radiative forcing effect between the present moment and
198 a selected time interval caused by a unit mass of gas emitted in the present time. The Kg CO₂ eq.

199 values were calculated per m³ of growing media and per produced plant using the GaBi software
200 (PE-International, <http://www.gabi-software.com/>), supported by the Ecoinvent 3.3 database.

201 2.5. Data analysis

202 Results are presented as mean of eight independent replicates plants for each type of growing
203 media. Significance of differences between mean values of plant elongation and dry biomass of
204 growing media parameters were assessed by the analysis of variance (ANOVA) followed by the
205 Fischer PLSD test, with $P < 0.05$ as significance threshold value.

206

207 **Results**

208 The main physicochemical characteristics of the different growing media are reported in Table 1.
209 The bulk density was significantly lower in the peat-based growing media (C), whereas followed by
210 S+PR, S and S+F growing media (Table 1). The pH value of the peat-based growing media was
211 acidic whereas the sediment-based growing media had all similar alkaline pH value (Table 1). The
212 EC value was significantly higher in the sediment-based growing media than in the peat-based
213 growing media, with the highest value found for the S growing media (Table 1). Total and organic
214 C contents were significantly higher in the peat-based than in sediment-based growing media,
215 whereas the total N content was significantly higher in the C and S+F than in the S and S+PR
216 growing media (Table 1). Total and available P were significantly higher in the S+F, followed
217 by the S and S+PR growing media, whereas the lowest total and available P concentrations were
218 observed in the C growing media (Table 1). Nutrients and heavy metals availability were relatively
219 low in sediments immediately after phytoremediation and peat prior to their use for preparing
220 growing media (Table 2). In C growing media at the end of the plant growth cycle, Cd, Cr, Cu, Ni
221 and Pb availability was below the detection limit, whereas availability of Ca, Cd, Cu and Zn was
222 significantly lower in the S+PR than in S and S+F growing media (Table 2). The Cr availability was

223 below the detection limit in the original phytoremediated sediment and peat, and in all growing
224 media at the end of the plant growth cycle (Table 2).

225
226 3.2 Plant growth, leaf chlorophyll content, and plant stress indication.

227 Plant elongation at the end of the growth period showed similar trends for all types of growing
228 media, with similar values for S+F and C treatments and significantly lower values for S and S+PR
229 growing media (Figure 1). Biomass was significantly higher for plants grown on C and S+F
230 growing media as compared to plants grown on S and S+PR growing media, either for roots and
231 shoots (Figure 1). The leaf dry weight showed no significant differences between plants regardless
232 of the growing media (Figure 1). Content of chlorophyll a was significantly higher in plants grown
233 on C and S+F growing media, content of chlorophyll b was higher in plants grown on C and S
234 growing media (Figure 1). As a result, total chlorophyll content was significantly higher in plants
235 grown on C and S+F growing media (Figure 1). Malondialdehyde (MDA) concentration was
236 significantly higher values in leaves of plants grown on C, followed by plants grown on S and
237 S+PR, whereas the significantly lower value was detected in leaves of plants grown on S+F
238 growing media (Figure 1).

239
240 3.3 Heavy metal and macronutrient concentrations in plant roots and leaves

241 Among heavy metals, Cu and Ni concentrations showed significantly higher concentrations in
242 leaves of plants grown on S+F, followed by plants grown on S+PR for Cu and grown on S+Pr and S
243 for Ni (Figure 2). Significantly higher concentrations of Pb and Zn were found in leaves of plants
244 grown on peat-based growing media (C), followed by plants grown on S+Pr and S+F for both
245 elements (Figure 2). The lowest Pb and Zn concentrations were detected in leaves of plants grown
246 on the S+Pr growing media (Figure 2). Calcium and Mg concentrations showed the following
247 ranking orders: $C > S+F > S+PR = S$ for Ca, and $S+PR > S = C > S+F$ for Mg (Figure 2).

248

249 3.4 LCA analysis

250 The LCA results showed a positive environmental effect of the use sediment in growing media, as it
251 resulted in values of 12.5 kg CO₂ eq per m³ of phytoremediated sediment and 70.0 kg CO₂ eq per
252 m³ of peat. Calculation of the relative impact of individual plants grown on sediment-based growing
253 media expressed as % of the GWP impact of the plants produced on peat-based growing media (the
254 standard practice), showed that the use of all sediment-based growing media reduced the C footprint
255 (Table 3). In particular, a reduction of C footprint of 24% was obtained using the S+F, of 26% using
256 the S and of 10% using the S+PR growing media, as compared to peat-based growing media. The
257 differences between the various sediment-based growing media were due to the fertilization for the
258 S+F and the use of composted pruning residues for the S+PR growing media (Table 3). The LCA
259 also showed that the growing plants on reclaimed sediment also reduced the incidence of the
260 growing media on the overall GWP per produced plant from 22% for peat-based growing media to
261 9%, 4% and 8% for the S, S+PR and S+F growing media (Table 3).

262

263 **Discussion**

264 The results of our work showed that marine sediments, reclaimed by phytoremediation could be
265 used for growing ornamental plants. The sediment-based growing media presented pH and EC
266 values and bulk density out of the typical range for commercially available growing media (Abad et
267 al., 2001), except for the bulk density and the EC value of the S+PR growing media, due to the
268 incorporation of pruning residues into sediments. Although the growing media physico-chemical
269 parameters are generally considered fundamental for optimal growing media (Schmilewski, 2009),
270 the red-robin photinia plants grew on all sediment-based growing media. However, it was also
271 observed that fertilization was the key factor for obtain plants of comparable size and form
272 comparable to those grown on the peat-based growing media (Stratton et al., 2001). Fertilization is

273 particularly important for plants grown in containers because of the limited amount of substrate
274 available for the roots, which can cause plant deficiency (Huett, 1997; Abad et al., 2001). The lower
275 growth of plants observed on S+PR growing media was also likely due to the lower nutrient content
276 of this growing media, particularly P, as compared to the other sediment-based growing media
277 (Table 1). In fact, the pruning residues, mainly consisting of lingo-cellulosic moiety, did not
278 contribute to the plant mineral nutrition and led to stunted growth (Figure 3). These results are in
279 agreement with pervious finding by Benito et al. (2005) who reported good growth of various
280 ornamental plants on composted pruning residues only when they were mixed with other fertile
281 ingredients in growing media. Possibly, a combination of pruning residues and fertilization may
282 lead to the constitution of suitable growing media.

283 Overall, these results showed that a standard fertilization with a slow-release NPK fertilizer was
284 sufficient to improve the plant growth on the phytoremediated sediment to comparable levels as the
285 peat-based growing media. This was indicated by the similar chlorophyll content in leaves of
286 sediment- and peat-based growing media. Chlorophyll a and b are the main photosynthetic
287 pigments, providing the chemical energy responsible for the biomass production (Filella et al.,
288 1995), and chlorophyll content is generally reduced under nutrient deficiency and plant stress
289 (Merzlyak et al., 1999; Moran et al., 2000). Chlorophyll content is also responsible for the plant
290 chroma, which is an important parameter for grading the ornamental plants. Improvement growth
291 and aspects of ornamental plants in relation to N availability have been reported both for (Mills and
292 Jones, 1996; Hernandez Apaolaza et al., 2005). An important aspect to underline, is that the current
293 Italian legislation (Legislative Decree 75/2010) does not officially admit reclaimed sediments as
294 materials for preparing growing media for plant nursery. The presented results showed that
295 phytoremediated sediments may be proposed as inorganic materials to be included in the Italian
296 legislation regulating the use of plant growing media through in a next legislative revision.

297 Leaf concentrations of Cu, Ni, Pb and Zn were relatively low and comparable with those measured
298 in 1 year old leaves of red robin photinia grown on contaminated a soil (Giorgioni and Quitadamo,
299 2013). The undetectable Cd and Cr concentrations in leaves of all plants could be explained by the
300 low Cd and Cr concentrations in the peat-based growing media and by the alkaline pH value of the
301 sediment-based growing media that could have reduced their phytoavailability by precipitation with
302 carbonates of sediments (Renella et al., 2004). indicated that were not taken up or not translocated
303 to the aerial part of plants grown on both sediments and peat.

304 The significantly higher MDA concentration in leaves of photinia plants grown on the peat-based as
305 compared to sediment-based growing media was unexpected, as MDA concentration in plant
306 increase in response to oxidative stress, and generally indicates the onset of a plant stress phase
307 (Posmyk et al., 2005). The MDA is one of the end products of polyunsaturated fatty acids
308 peroxidation (Upadhyaya and Panda, 2004), induced by an imbalance between oxidative and
309 reductive reactions in favor of the oxidative processes, leading to the production of oxygen radical
310 species, such as superoxide anion ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2) and the hydroxyl- radical ($OH\cdot$)
311 (Sies, 1991). Increased MDA content in plants has been observed under stress induced by light,
312 temperature, drought, hypoxia and exposure to heavy metals (Mishra and Singhal, 1992; Blokhina
313 et al., 2003; de Carvalho, 2008). Bennicelli et al. (1998), studied that the time relationship of TBA-
314 MDA and the superoxide dismutase (SOD) foliar concentration in function of substrate aeration,
315 and reported that plants grown on oxygen deficient growing media produced higher SOD activity
316 and had a lower TBA-MDA concentrations, whereas plants on more aerated growing media had a
317 lower SOD and higher MDA concentrations. It can not be excluded that plants grown on sediment-
318 based growing media adapted to hypoxic conditions by inactivation of active forms of oxygen
319 (Vergara et al., 2012), therefore reducing the peroxidation products. However, it has been reported
320 that the increase of MDA content may be specifically related to oxidative stress (Bennicelli et al.,
321 1998). Non specific reactions of TBA with other non-lipid metabolites (e.g. cellular carbohydrates)

322 or fatty peroxide-derived decomposition products present in plants grown on peat-based growing
323 media can not be excluded (Janero, 1990; Valenzuela, 1991). Phenylpropanoid-type pigments can
324 also contribute to the overestimation of the MDA content in leaves (Stafford, 1994).

325 The LCA showed that the sediment re-cycle as ingredient of growing media for plant nursery can
326 also improve the C footprint of plant production as compared to the use of the traditional peat-based
327 growing media. The reduction of the C footprint related to the use of sediments in growing media
328 was similar between the S and S+F, with the only difference due to the use of the controlled release
329 fertilizer. Differently, plants grown on S+PR growing media had a less positive result (90%
330 compared the control plants), due to the environmental impact associated to the pruning residue
331 composting process. The LCA also confirmed that the growing media contribute less to the GWP of
332 nursery plant nursery production which are predominated by the cultivation practice (Lazzerini et
333 al., 2016), but the use of sediment-based growing media can further reduce its relative contribution
334 (Table 3). Overall, by considering the results of the LCA and the volumes of growing media used in
335 the EU plant nursery districts, the sediment re-use in plant nursery is an option may significantly
336 alleviate the problems of sediment inland management and disposal (Bert et al., 2009). The benefit
337 for the dredging industrial sector may be even larger by reducing that the sediment re-use in the
338 plant nursery further reduce the high environmental impact associated to the sediment storage in
339 confined facilities (Puccini et al., 2013; Bates et al., 2015).

340

341 **Conclusions**

342 Our results demonstrated that phytoremediated marine sediments can be used for growing high
343 grade ornamental plants in container, regardless of the residual organic and heavy metal
344 contamination, due to both the low metal availability and low metal accumulating capacity of the
345 ornamental plants. Standard fertilization was sufficient to produce plants with a comparable plant
346 grade as those grown on traditional peat-based growing media, whereas the mixing of sediments

347 with composted pruning residues improved the bulk density and EC value of the growing media but
348 resulted in lower plant growth and needs further improvement. The use of growing media based on
349 phytoremediated sediments reduced the C footprint of plant production offering practical
350 management alternatives to both plant nursery and dredging industries), particularly by setting short
351 supply chains of sediment dredging, remediation and use. We see no reasons for hindrance in the
352 official admission of phytoremediated sediments in the National legislation regulating the plant
353 growing media as inorganic matrices.

354

355 **Acknowledgements** The experiment was carried out in the Center for Experimental Nursery
356 (CeSpeVi), Pistoia (Tuscany). We wish to thank Dr Paolo Marzialetti for his fundamental advice
357 and assistance in the plant growth.

358

359 **References**

360 Abad, M., Noguera, P., Bures, S. 2001. National inventory of organic wastes for use as growing
361 media for ornamental potted plant production: case study in Spain. *Bioresource Technology*, 77,
362 197-200.

363 Altmann, M. 2008. Socio-Economic Impact of the Peat and Growing Media Industry on
364 Horticulture in the EU. Available at
365 http://www.epagma.eu/sites/default/files/growing%20media/socio_economic_study2008.pdf
366 (accessed on December 2017).

367 Barcelona Convention 1995. Dumping protocol. Available at
368 http://wedocs.unep.org/bitstream/id/53220/consolidated_dumping_eng.pdf (accessed on December
369 2017).

- 370 Bates, M.E., Fox-Lent, C., Seymour, L., Wender, B.A., Linkov, I. 2015. Life cycle assessment for
371 dredged sediment placement strategies. *Science of the Total Environment* 11,309-318. doi:
372 10.1016/j.scitotenv.2014.11.003. Epub 2014 Dec 29.
- 373 Benito, M., Masaguer, A., Antonio, R. D., Moliner, A. 2005. Use of pruning waste compost as a
374 component in soilless growing media. *Bioresource Thecnology*, 96, 597-603.
- 375 Bennicelli, R.P., Stepniewski, W., Zakrzhevsky, D.A., Balakhnina, T.I., Stepniewska, Z., Lipiec, J.
376 1998. The effect of soil aeration on superoxide dismutase activity, malondialdehyde level, pigment
377 content and stomatal diffusive resistance in maize seedlings. *Environmental and Experimental*
378 *Botany* 39, 203-211.
- 379 Bert, V., Seuntjens, P., Dejonghe, W., Lacherez, S., Thuy, H.T., Vandecasteele, B. 2009.
380 Phytoremediation as a management option for contaminated sediments in tidal marshes, flood
381 control areas and dredged sediment landfill sites. *Environmental Science and Pollution Research*
382 *International*, 16, 745-64
- 383 Blokhina, O., Virolainen, E., Fagerstedt, K.V. 2003. Antioxidants, oxidative damage and oxygen
384 deprivation stress: a review. *Annals of Botany* 91, 179-194.
- 385 Bortone, G., Arevalo, E. , Deibel, I., Detzner, H., DePropriis, L., Elskens, F., Giordano, A.,
386 Hakstege, P., Hamer, K., Harmsen, J., Hauge, A., Palumbo, L., VanVeen, J. 2004. Synthesis of the
387 sednet workpackage 4 outcomes. *J. Soils Sediments* 4,225-232.
- 388 Ceglie, F.G., Bustamante, M.A., Ben Amara, M., Tittarelli, F. 2015. The challenge of peat
389 substitution in organic seedling production: Optimization of growing media formulation through
390 mixture design and response surface analysis. *PLoS ONE*10(6), e0128600.
391 <https://doi.org/10.1371/journal.pone.0128600>.
- 392 Cellura, M., Longo, S., Mistretta, M. 2012. Life Cycle Assessment (LCA) of protected crops: an
393 Italian case study. *Journal of cleaner production*, 28, 56-62.

- 394 Cruz de Carvalho, M. H. 2008. Drought stress and reactive oxygen species. *Plant Signaling and*
395 *Behavior* 3:3, 156-165.
- 396 De Lucia, B., Cristiano, G., Vecchietti, L., Rea, E., Russo, G. 2013. Nursery growing media:
397 agronomic and environmental quality assessment of sewage sludge-based compost. *Applied and*
398 *Doni, S., Macci, C., Peruzzi, E., Iannelli, R., Masciandaro, G. 2015. Heavy metal distribution in a*
399 *sediment phytoremediation system at pilot scale. Ecological Engineering* 81, 146-157.
- 400 *Environmental Soil Science, Volume 2013, article id 565139,*
401 <http://dx.doi.org/10.1155/2013/565139>.
- 402 European Commission, 2015. European Commission, DG Agriculture and Rural Development,
403 Civil Dialogue Group, Horticultural Products, Flowers and Ornamental Plants, Working Document
404 02/10/2015. Available at [https://ec.europa.eu/agriculture/sites/agriculture/files/fruit-and-](https://ec.europa.eu/agriculture/sites/agriculture/files/fruit-and-vegetables/product-reports/flowers/statistics-2015_en.pdf)
405 [vegetables/product-reports/flowers/statistics-2015_en.pdf](https://ec.europa.eu/agriculture/sites/agriculture/files/fruit-and-vegetables/product-reports/flowers/statistics-2015_en.pdf) (accessed on December 2017).
- 406 Unit C.2. Olive oil, horticultural products Filella, I., Serrano, I., Serra, J., Peñuelas, J. 1995.
407 Evaluating wheat nitrogen status with canopy reflectance indices and discriminant analysis. *Crop*
408 *Science* 35, 1400–1405
- 409 Garcia-Gomez, A., Bernal, M.P., Roig, A. 2002. Growth of ornamental plants in two composts
410 prepared from agroindustrial wastes. *Bioresource Technology*, 83, 81-87.
- 411 Giorgioni, M.E., Quitadamo, L. 2013. Ornamental Shrub capacity for absorption and accumulation
412 of heavy metals form urban polluted soils. *Acta Horticulture* 990, 501-508.
- 413 Hernandez Apaolaza, L., Gasco, A. M., Gasco, J. M., Guerrero, F. 2005. Reuse of waste materials
414 as growing media for ornamental plants. *Bioresource Technology*, 96, 125 -131.
- 415 Huett, D.O. 1997. Fertiliser use efficiency by containerised nursery plants 2. Nutrient leaching.
416 *Australian Journal of Agricultural Research*, 48, 259-265.
- 417 ISO, International Standard Organization (2006). ISO 14044:2006 Environmental management -
418 Life cycle assessment - Requirements and guidelines.

- 419 Janero, D.R. 1990. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid
420 peroxidation and peroxidative tissue injury. *Free Rad Biol Med* 9, 515-540.
- 421 Jeffrey, S.W., Humphrey, G.F. 1975. New spectrophotometric equations for determining
422 chlorophyll a, b c1 and c2 in higher plants, algae and natural phytoplankton. *Biochemie und*
423 *Physiologie Pflanzen* 167, 191-194.
- 424 Lazzerini, G., Lucchetti, S., Nicese, F.P. 2016. Green House Gases(GHG) emissions from the
425 ornamental plant nursery industry: a Life Cycle Assessment(LCA) approach in a nursery district in
426 central Italy. *Journal of Cleaner Production* 112, 4022-4030.
- 427 Masciandaro, G., Di Biase, A., Macci, C., Peruzzi, E., Iannelli, R., Doni, S. 2014.
428 Phytoremediation of dredged marine sediment: Monitoring of chemical and biochemical processes
429 contributing to sediment reclamation. *Journal of Environmental Management* 134, 166-174.
- 430 Mattei, P., D'Acqui, L.P., Nicese, F.P., Lazzerini, G., Masciandaro, G., Macci, C., Doni, S.,
431 Sarteschi, F., Giagnoni, L., Renella, G. 2017. Use of phytoremediated sediments dredged in
432 maritime port as plant nursery growing media. *Journal of Environmental Management* 186, 225-
433 232.
- 434 McGrath, S.P., Cunliffe, C.H. 1985. A simplified method for the extraction of the metals Fe, Zn,
435 Cu, Ni, Cd, Pb, Cr, Co and Mn from soils and sewage sludges. *Journal of the Science of Food and*
436 *Agriculture* 36, 794-798
- 437 Merzlyak, M.N., Gitelson, A.A., Chivkunova, O.B., Rakitin, V.Y. 1999. Nondestructive optical
438 detection of leaf senescence and fruit ripening. *Physiologia Plantarum* 106, 135–141.
- 439 Mills, H.A., Jones, J.B. 1996. *Plant analysis handbook II. A practical sampling, preparation,*
440 *analysis, and interpretation guide.* MicroMacro Publishing Inc., Athens, GA, USA, pp. 422.
- 441 Mishra, R.K., Singhal, G.S. 1992. Function of Photosynthetic Apparatus of Intact Wheat Leaves
442 under High Light and Heat Stress and Its Relationship with Peroxidation of Thylakoid Lipids. *Plant*
443 *Physiology* 98, 1-6.

- 444 Moran, J.A., Mitchell, A.K., Goodmanson, G., Stockburger, K.A. 2000. Differentiation among
445 effects of nitrogen fertilization treatments on conifer seedlings by foliar reflectance: a comparison
446 of methods. *Tree Physiology* 20, 1113–1120.
- 447 Pignattelli, S., Colzi, I., Buccianti, A., Cecchi, L., Arnetoli, M., Monnanni, R., Gabbrielli, R.,
448 Gonnelli, C. 2012. Exploring element accumulation patterns of a metal excluder plant naturally
449 colonizing a highly contaminated soil. *Journal of Hazardous Materials* 227, 362-369.
- 450 Posmyk, M.M., Bailly, C., Szafranska, K., Janas, K.M., Corbineau, F. 2005. Antioxidant enzymes
451 and isoflavonoids in chilled soybean (*Glycine max* (L.) Merr.) seedlings. *Journal of Plant*
452 *Physiology* 162, 403-412.
- 453 Puccini, M., Seggiani, M, Vitolo, S, Iannelli, R. 2013. Life cycle assessment of remediation
454 alternatives for dredged sediments. *Chemical Engineering Transactions* 35, 781-786.
- 455 Renella, G., Adamo, P., Bianco, M.R., Landi, L., Violante, P., Nannipieri, P. 2004. Availability and
456 speciation of cadmium added to a calcareous soil under various managements. *European Journal of*
457 *Soil Science* 55, 123-133.
- 458 Santi, C.A., Certini, G., D'Acqui, L.P. 2006. Direct determination of organic carbon by dry
459 combustion in soils with carbonates. *Communications in Soil Science and Plant Analysis* 37, 155-
460 162.
- 461 Schmilewski, G. 2009. Growing medium constituents used in the EU. *Acta Horticulture* 819, 33-46.
- 462 PAS 2050:2011 Specification for the assessment of the life cycle greenhouse gas emissions of
463 goods and services.
- 464 SedNet (European Sediment Research Network) 2003. The SedNet strategy paper: The opinion of
465 SedNet on environmentally, socially, and economically viable sediment management. Available at
466 www.sednet.org (accessed on December 2017).

- 467 Shine, K.P., Fuglestedt, J.S., Hailemariam, K., Stuber, N. 2005. Alternatives to the global warming
468 potential for comparing climate impacts of emissions of greenhouse gases. *Climatic Change* 68,
469 281-302.
- 470 Sies, H. 1991. Oxidative stress, oxidants and antioxidants. *Experimental Physiology* 82, 291-295.
- 471 Stafford H. 1994. Anthocyanins and betalains: evolution of mutually exclusive pathways. *Plant*
472 *Science*, 101, 91-98.
- 473 Stratton, M. L., Good, G. L., Barker, A.V. 2001. The effects of nitrogen source and concentration
474 on the growth and mineral composition of privet. *Journal of Plant Nutrition*, 24, 1745-1772.
- 475 Taiti, C., Giorni, E., Colzi, I., Pignattelli, S., Bazihizina, N., Buccianti, A., Luti, S., Pazzagli, L.,
476 Mancuso, S., Gonnelli, C. 2016. Under fungal attack on a metalliferous soil: ROS or not ROS?
477 Insights from *Silene paradoxa* L. growing under copper stress *Environmental Pollution* 210, 282-
478 292
- 479 Upadhyaya, H., Panda, S.K. 2004. Responses of *Camellia senensis* to drought and rehydration.
480 *Biologia Plantarum* 48, 597-600.
- 481 Valenzuela, A. 1991. The biological significance of malondialdehyde determination in the
482 assessment of tissue oxidative stress. *Life Sciences* 48, 301-309.
- 483 Vergara, R., Parada, F., Rubio, S., Perez, F.J. 2012. Hypoxia induces H₂O₂ production and activates
484 antioxidant defence system in grapevine buds through mediation of H₂O₂ and ethylene. *Journal of*
485 *Experimental Botany* 63, 4123-4131.

Figure 1. LCA system diagram of the considered materials and processes in for the experiment of the Red Robin photinia production on sediment- and peat-based growing media.

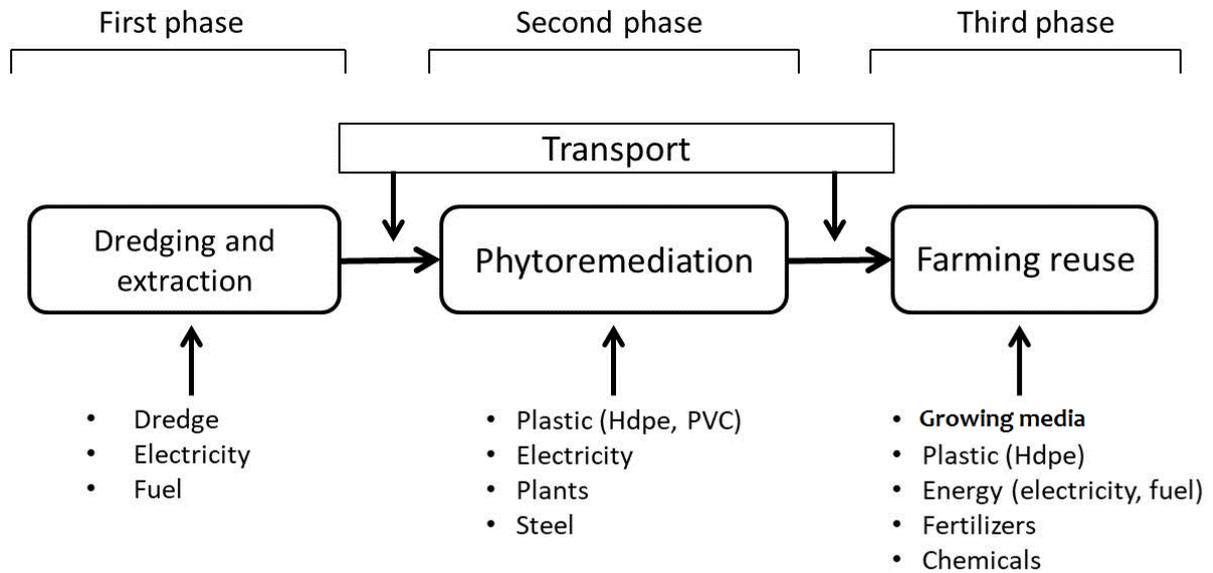


Figure 2. Red robin photinia pant elongation, biomass, leaf dry weight, chlorophyll content and malondialdehyde concentration after growth on sediment- and peat-based growing media. Different superscripts indicate significant differences ($P < 0.05$) among mean values for plant parameters grown on different growing media. Symbols S, S+PR, S+F and C indicate the sediment, sediment plus pruning residues, sediment plus fertilizer and peat-based growing media, respectively.

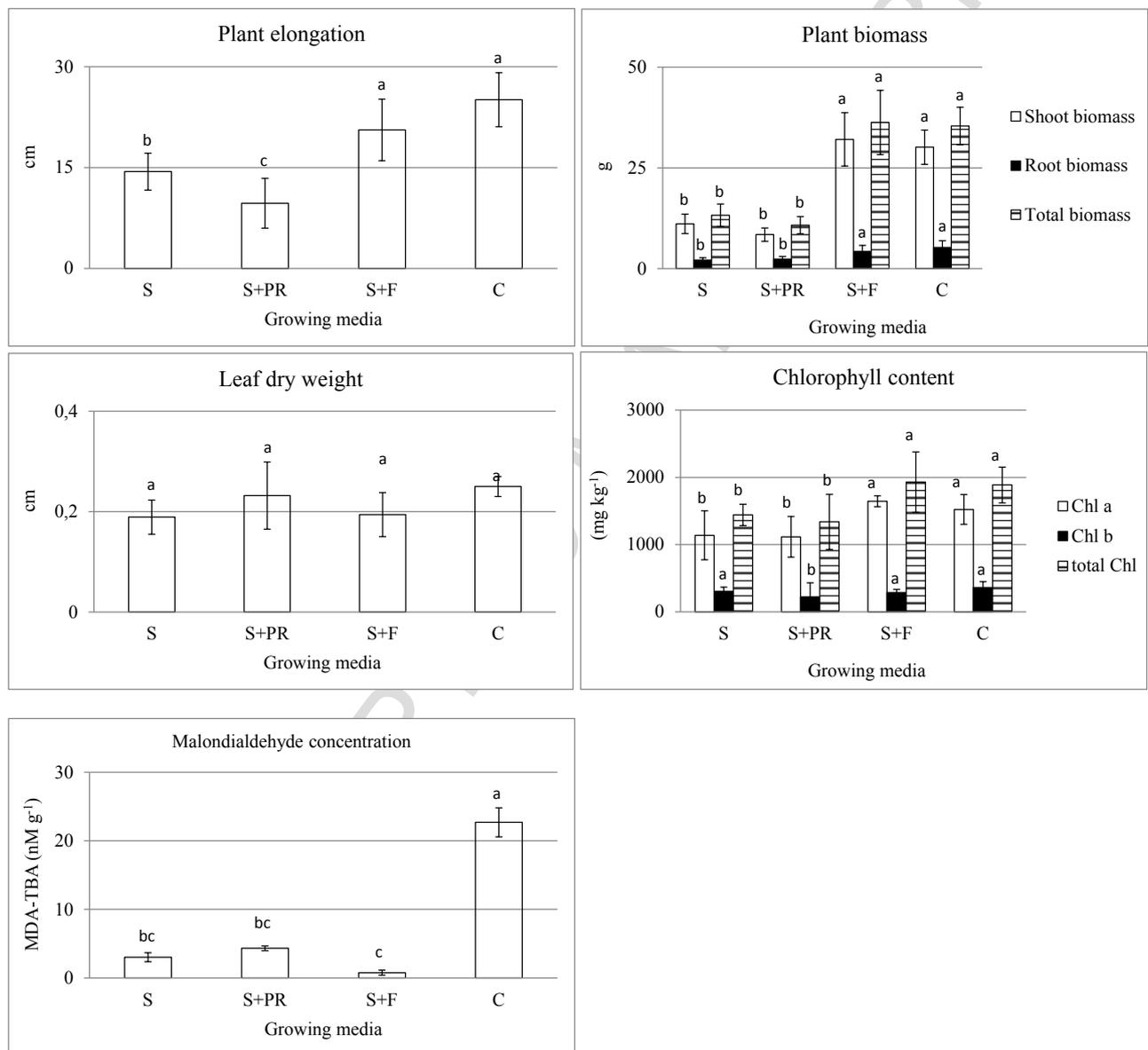
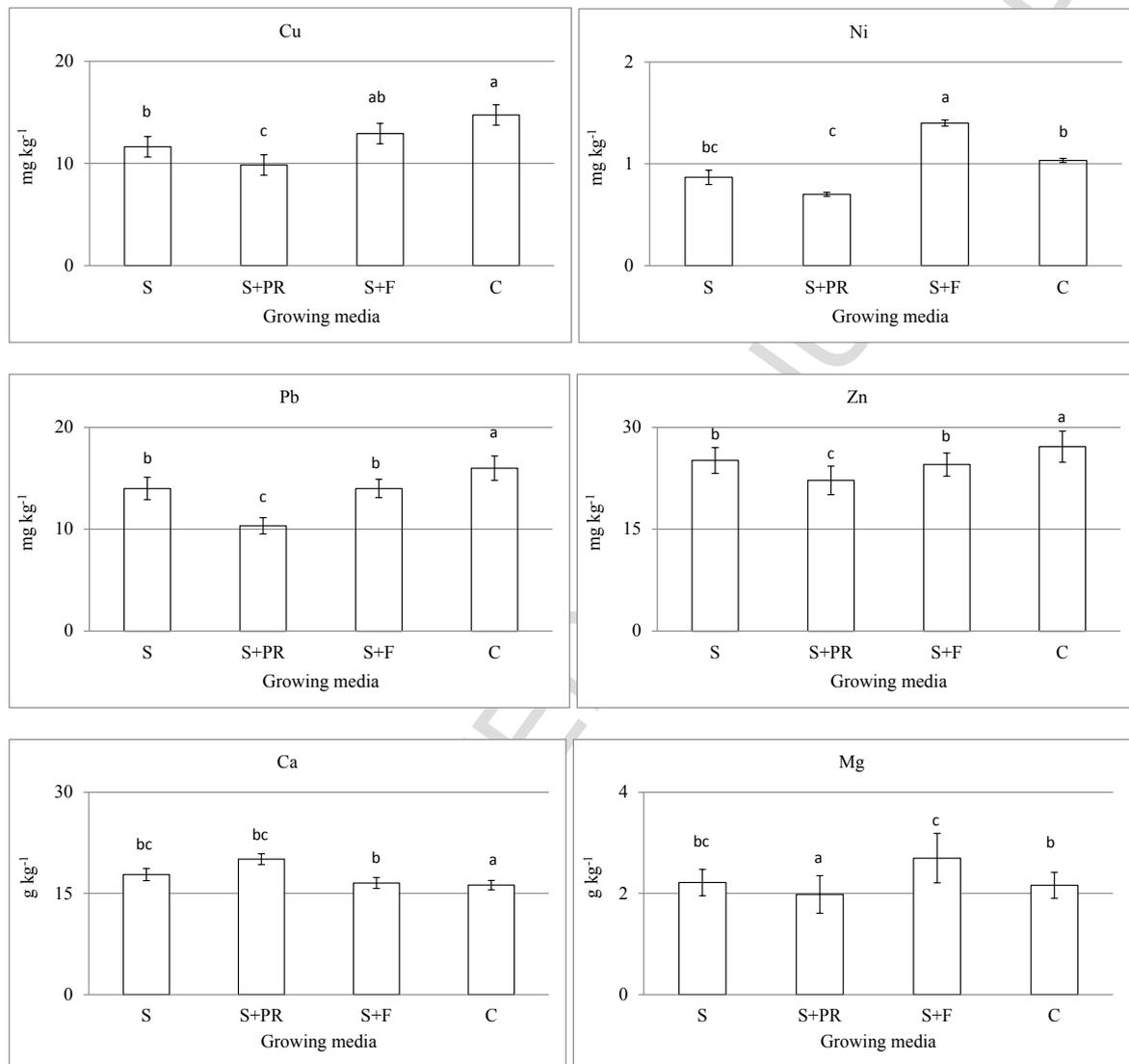


Figure 3. Concentration of heavy metals and macronutrients in the red robin photinia leaves at the end of the plant growth period. Different superscripts indicate significant differences ($P < 0.05$) among mean values for leaves of plants grown on different growing media. Symbols S, S+PR, S+F and C indicate the sediment, sediment plus pruning residues, sediment plus fertilizer and peat-based growing media, respectively.



Highlights

- ▶ Phytoremediated marine were used as peat-free growing media for growing red robin photinia plants
- ▶ Environmental impact related to the use of sediments- common peat-based growing were compared by LCA
- ▶ Plants grown on fertilized sediments were comparable to those grown on peat-based growing media
- ▶ The LCA showed a lower C footprint of plants grown on sediment- than peat-based growing media

Table 1. Main physico-chemical parameters of the sediment- and peat-based growing media. Different superscripts indicate significant differences ($P < 0.05$) among mean values ($n = 8$). Symbols S, S+PR, S+F and C indicate the sediment, sediment plus pruning residues, sediment plus fertilizer and peat-based growing media, respectively.

Physico-chemical parameters	Growing media			
	S	S+PR	S+F	C
Bulk density (kg m ³)	1012±74 ^c	684±59 ^b	982±68 ^c	238±26 ^a
pH _(H2O)	7.88±0.03 ^b	7.69±0.04 ^b	7.78±0.02 ^b	4.55±0.08 ^a
EC (mS cm ⁻¹)	1.08±0.09 ^a	0.87±0.12 ^c	0.92±0.07 ^c	0.38±0.06 ^c
Total C (g kg ⁻¹)	27.3±2.4 ^c	40.8±4.9 ^b	26.2±2.7 ^c	893±24.9 ^a
Organic C (g kg ⁻¹)	17.3±2.3 ^c	31.4±2.8 ^b	16.1±1.3 ^c	524.9±2.2 ^a
Inorganic C (g kg ⁻¹)	11.9±2.0 ^b	13.4±1.8 ^b	9.17±0.33 ^a	ND*
Total N (g kg ⁻¹)	1.22±0.11 ^b	1.53±0.31 ^b	2.67±0.33 ^a	2.50±0.24 ^a
Total P (g kg ⁻¹)	51.3±3.78 ^a	33.6±2.5 ^b	57.4±13.1 ^a	7.96±1.30 ^d
Available P (mg kg ⁻¹)	2.43±0.15 ^b	1.57±0.10 ^c	3.22±0.27 ^a	0.46±0.06 ^d
Ca (g kg ⁻¹)	27.3±3.1 ^a	16.5±2.1 ^b	22.9±1.83 ^a	0.43±0.05 ^c
Mg (g kg ⁻¹)	16.2±1.5 ^a	10.6±1.9 ^b	17.0±2.09 ^a	0.50±0.14 ^c
Cd (mg kg ⁻¹)	1.52±0.27 ^a	0.72±0.10 ^b	1.37±0.21 ^a	0.49±0.09 ^c
Cr (mg kg ⁻¹)	59.3±5.5 ^a	35.7±2.5 ^b	61.6±3.5 ^a	34.1±17 ^b
Cu (mg kg ⁻¹)	82.1±5.6 ^b	59.0±6.0 ^c	82.7±4.1 ^b	126.6±20 ^a
Ni (mg kg ⁻¹)	67.1±4.0 ^a	40.6±4.05 ^b	65.8±5.1 ^a	2.24±0.79 ^c
Pb (mg kg ⁻¹)	65.1±2.9 ^a	39.6±3.5 ^b	64.8±4.5 ^a	9.14±1.3 ^c
Zn (mg kg ⁻¹)	284.6±24 ^a	256.9±50 ^a	286.1±29 ^a	9.15±1.9 ^b

*ND indicates values not determined.

Table 2. Heavy metal availability in phytoremediated sediment and peat before their use for preparation growing media and at the end of the plant growth cycle. Different superscripts indicate significant differences ($P < 0.05$) among mean values ($n = 8$) for both the original materials and at the end of the plant growth cycle. Symbols S, S+PR, S+F and C indicate the sediment, sediment plus pruning residues, sediment plus fertilizer and peat-based growing media, respectively. Values indicated by < 0.01 indicate concentrations below the detection limits.

Elements	Original materials				
	Sediment	Peat	Growing media		
			S+PR	S+F	C
Ca (mg kg ⁻¹)	2.3±0.2 ^a	0.41±0.06 ^b	1.011±0.24 ^b	2.246±0.25 ^a	0.506±0.20 ^c
Mg (mg kg ⁻¹)	0.39±0.05 ^a	0.16±0.02 ^b	0.32±0.07 ^a	0.33±0.04 ^a	0.14±0.03 ^b
Cd (mg kg ⁻¹)	0.04±0.01	< 0.01	0.020±0.002 ^b	0.031±0.005 ^a	< 0.01
Cr (mg kg ⁻¹)	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
Cu (mg kg ⁻¹)	0.04±0.01	< 0.01	0.012±0.002 ^b	0.032±0.005 ^a	< 0.01
Ni (mg kg ⁻¹)	0.02±0.003	< 0.01	< 0.01	0.018±0.001 ^a	< 0.01
Pb (mg kg ⁻¹)	0.01±0.0	< 0.01	< 0.01	< 0.01	< 0.01
Zn (mg kg ⁻¹)	0.05±0.01 ^a	0.03±0.04 ^a	0.028±0.001 ^b	0.040±0.003 ^a	0.024±0.003 ^b

Table 3. GWP values expressed as CO₂ equivalent (CO₂ eq.) of red robin photinia plants produced on sediment-based compared to the same plants produced on peat-based growing media, and GWP values of the tested growing media. Values in the ‘Variation’ column indicate the percentage of reduction of CO₂ eq. as compared to the plants grown on the different growing media. Values in the ‘Incidence’ indicate the relative contribution of growing media to the CO₂ eq. for the plants grown on the different growing media. Symbols S, S+PR, S+F and C indicate the sediment, sediment plus pruning residues, sediment plus fertilizer and peat-based growing media, respectively.

Growing media	Per plant	GWP (Kg CO ₂ eq.)		
		Growing media	Variation (%)	Incidence (%)
S	0.232	0.02	-25.7	9
S+PR	0.282	0.01	-9.6	4
S+F	0.236	0.02	-24.4	8
C	0.312	0.07	-	22