Accepted Manuscript

Title: Targeting STATs in neuroinflammation: the Road Less Traveled!

Authors: Seyed Mohammad Nabavi, Touqeer Ahmed, Maheen Nawaz, Kasi Pandima Devi, Devasahayam Jaya Balan, Valeria Pittalà, Sandro Argüelles-Castilla, Lara Testai, Haroon Khan, Antoni Sureda, Marcos Roberto de Oliveira, Rosa Anna Vacca, Suowen Xu, Bahman Yousefi, Valeria Curti, Maria Daglia, Eduardo Sobarzo-Sánchez, Rosanna Filosa, Seyed Fazel Nabavi, Maryam Majidinia, Ahmad Reza Dehpour, Samira Shirooie



| PII: | \$1043-6618(18)30572-3 |
|----------------|--|
| DOI: | https://doi.org/10.1016/j.phrs.2018.12.004 |
| Reference: | YPHRS 4096 |
| To appear in: | Pharmacological Research |
| Received date: | 23 July 2018 |
| Revised date: | 1 December 2018 |
| Accepted date: | 10 December 2018 |

Please cite this article as: Nabavi SM, Ahmed T, Nawaz M, Devi KP, Balan DJ, Pittalà V, Argüelles-Castilla S, Testai L, Khan H, Sureda A, de Oliveira MR, Vacca RA, Xu S, Yousefi B, Curti V, Daglia M, Sobarzo-Sánchez E, Filosa R, Nabavi SF, Majidinia M, Dehpour AR, Shirooie S, Targeting STATs in neuroinflammation: the Road Less Traveled!, *Pharmacological Research* (2018), https://doi.org/10.1016/j.phrs.2018.12.004

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Targeting STATs in neuroinflammation: the Road Less Traveled!

Seyed Mohammad Nabavi ^{1*}, Touqeer Ahmed ², Maheen Nawaz ², Kasi Pandima Devi ³, Devasahayam Jaya Balan ³, Valeria Pittalà ⁴, Sandro Argüelles-Castilla ⁵, Lara Testai ⁶, Haroon Khan ⁷, Antoni Sureda ^{8*}, Marcos Roberto de Oliveira ⁹, Rosa Anna Vacca ¹⁰, Suowen Xu ¹¹, Bahman Yousefi ^{12, 13}, Valeria Curti ¹⁴, Maria Daglia ¹⁴, Eduardo Sobarzo-Sánchez ^{15, 16}, Rosanna Filosa ¹⁷, Seyed Fazel Nabavi ¹, Maryam Majidinia ¹⁸, Ahmad Reza Dehpour ¹⁹ and Samira Shirooie ²⁰

1. Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran

2. Neurobiology Laboratory, Department of Healthcare Biotechnology, Atta-ur-Rahman School of Applied Biosciences, National University of Sciences and Technology, Sector H-12, Islamabad – 44000, Pakistan

3. Department of Biotechnology, Alagappa University (Science Campus), Karaikudi 630 003, Tamil Nadu, India

4. Department of Drug Sciences Sciences, Viale Andrea Doria 6, 95125 - Catania, Italy

5. Department of Physiology, Faculty of Pharmacy, University of Seville, Seville, Spain

6. Department of Pharmacy, via Bonanno 6 - 56126, Pisa. Italy

7. Department of Pharmacy, Abdul Wali Khan University, Mardan 23200, Pakistan

8. Research Group on Community Nutrition and Oxidative Stress and CIBEROBN (Physiopathology of Obesity and Nutrition), University of Balearic Islands, E-07122 Palma de Mallorca, Spain

9. Department of Chemistry/ICET, Federal University of Mato Grosso (UFMT), Av. Fernando Corrêa da Costa, 2367, Cuiaba, MT, 78060-900, Brazil

10. Institute of Biomembranes, Bioenergetics and Molecular Biotechnologies, National Council of Research, I-70126, Bari, Italy

11. University of Rochester, Aab Cardiovascular Research Institute, Rochester, NY, 14623, USA

12. Molecular Medicine Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

13. Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

14. Department of Drug Science of University of Pavia, Pavia, Italy

15. Laboratory of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Santiago de Compostela, 15782, Spain

16. Instituto de Investigación e Innovación en Salud, Facultad de Ciencias de la Salud, Universidad Central de Chile, Santiago, Chile

17. Second University of Naples, Via L. De Crecchio, 7 80138 Napoli, Italy

18. Solid Tumor Research Center, Urmia University of Medical Sciences, Urmia, Iran

19. Department of Pharmacology, Faculty of Medicine, Tehran University of Medical Sciences, Tehran, Iran

20. Department of Pharmacology, Faculty of Pharmacy, Kermanshah University of Medical Sciences, Kermanshah, Iran

Short title: STAT inhibitors in neuroinflammation

Address for correspondence: SM Nabavi, Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran, email: <u>Nabavi208@gmail.com</u>; A Sureda, Research Group on Community Nutrition and Oxidative Stress (NUCOX) and CIBEROBN (Physiopathology of Obesity and Nutrition CB12/03/ 30038), University of Balearic Islands, Ctra. Valldemossa, Km 7.5, E-07122 Palma de Mallorca, Balearic Islands, Spain, email: <u>antoni.sureda@uib.es</u>

Graphical abstract



Abstract

JAK/STAT transduction pathway is a highly conserved pathway implicated in regulating cellular proliferation, differentiation, survival and apoptosis. Dysregulation of this pathway is involved in the onset of autoimmune, haematological, oncological, metabolic and neurological diseases. Over the last few years, the research of anti-neuroinflammatory agents has gained considerable

attention. The ability to diminish the STAT-induced transcription of inflammatory genes is documented for both natural compounds (such as polyphenols) and chemical drugs. Among polyphenols, quercetin and curcumin directly inhibit STAT, while *Berberis vulgaris* L. and *Sophora alopecuroides* L extracts act indirectly. Also, the Food and Drug Administration has approved several JAK/STAT inhibitors (direct or indirect) for treating inflammatory diseases, indicating STAT can be considered as a therapeutic target for neuroinflammatory pathologies through direct and indirect interaction. Considering the encouraging data obtained so far, clinical trials are warranted to demonstrate the effectiveness and potential use in the clinical practice of STAT inhibitors to treat inflammation-associated neurodegenerative pathologies.

Keywords: STAT; neuroinflammation; inhibitors; therapeutics

1. Introduction

Neuroinflammation is a very complex process comprising all the inflammatory processes that occur within the central nervous system (CNS) and involving multiple cell types and mediators depending on the cause of origin and evolution. The inflammatory response is under tight control by both pro- and anti-inflammatory signals and mediators. In general, transient and acute inflammatory response is beneficial, inducing an adaptive response that helps the host to be defended against pathogens. On the contrary, chronic neuroinflammation is deleterious leading to neuronal dysfunction and tissue damage. Thus, inflammation of the CNS is an essential

process in protecting the brain against insults, but it can also contribute to worse many brain and neurological diseases, including brain injuries, cancer, epilepsies and neurodegenerative diseases mainly Parkinson's or Alzheimer's diseases [1,2]. The inflammatory process in the CNS is mainly mediated by the activation of the microglia, resident macrophages and astrocytes and the release of many mediators including cytokines, chemokines or growth factors [3].

STATs (Signal transducers and activators of transcription) are transcription factors found within the cytoplasm in an inactive or latent state. STAT and Janus kinase (JAK) constitute the ubiquitous and highly conserved JAK/STAT pathway, which is involved in transducing signals from membrane-localized receptors to cell nucleus. When cytokines, hormones and growth factors bind to their own receptor, conformational changes are induced leading to activation of JAKs. Activated JAKs phosphorylate a tyrosine residue in the cytokine receptor, providing a docking site for STATs. Then, STATs bind to the cytokine receptor through SH2 (Src 2 homology) domains, and are rapidly activated upon autophosphorylation on a tyrosine residue existing in the C-terminal domain (CTD). Phosphorylated STATs are released from the receptor, undergo homo-dimerization (and in some cases hetero-dimerization) and translocate into the cell nucleus where they could bind to target genes promoters, thereby engaging diverse elements of the transcriptional apparatus and stimulating gene expression [4-7]. STATs are DNA-binding proteins downstream of the engagement of JAK and is responsible for regulating diverse cellular processes related to cell metabolism, inflammation, apoptosis and immune response [6,8-10]. Moreover, an elevated gene expression of STAT pathway, derived from hyperactivation of the JAK/STAT pathway, is present in many neurological disorders contributing to disease pathogenesis [11]. Therefore, a dysregulation of this pathway is implicated in various diseases, including autoimmune, haematological, oncological, metabolic and neurological diseases [12-15].

Conversely, genetic dysfunctions/mutations of JAK/STAT pathway are implicated in inflammatory diseases, erythrocytosis, leukemias, as well as in the predisposition to obesity or type 2 diabetes mellitus (T2DM) [16]. This scenario highlights the deep implication of JAK/STAT in the cell homeostasis.

Many natural and synthetic substances have the capability to interfere with JAK/STAT pathways. Thus, they could represent a therapeutic option among other treatments available for neuroinflammatory diseases. These include natural compounds chiefly, polyphenols, extracted from plants with potential to develop as nutraceuticals, synthetic drugs and metallic compounds. This review aims to report data from literature regarding the role of STATs in neuroinflammation and the interaction of certain molecules and plant extracts with JAK/STAT pathways being able to become therapeutic option for neurodegenerative pathologies.

2. The STAT family: origin, structure and function

In mammals, the JAK family consists of 4 members: JAK1, JAK2, JAK3, and Tyk2 whereas the STAT family is comprised by seven members to date: STAT1-4, STAT5A-B and STAT6, all of which retain a tyrosine residue near C-terminal target of phosphorylation by JAKs [17,18]. JAK/STAT pathway is schematically represented in figure 1. Depending on the cytokine or growth factor signals, diverse combinations between JAKs and STATs can be produced with an elevated grade of specificity [13]. The seven STAT proteins derive from gene duplication and random genetics changes. In this sense, Souza-Neto et al. using comparative genomics confirmed the conservation of JAK-STAT pathway between insects and mammals [19]. The STAT molecule STAT92E from *Drosophila melanogaster* is homologue to human STAT5 with an identity of 37% [20]. Through evolution the ancestral STAT genes duplicated into A- and B-

type STATs [21]. The D-STAT, Ag-STAT, STATs 5A, 5B and 6 comes under A-type STATs. Among these D-STAT and Ag-STAT are insect STATs found in *D. melanogaster* and *Anopheles gambiae*. The STATs 5A, 5B and 6 are vertebrate STATs. B-type STATs, which include STATs 1, 2, 3 and 4, belong to vertebrate. Since no B-type STAT is observed in insect genomes, it has been predicted that the duplication of STAT genes happened previous to the evolutionary separation between vertebrates and insects [22]. The sequencing of STAT homologues in vertebrate showed the presence of conserved motif of C-terminal serine phosphorylation in STAT1, STAT3 and STAT4, whereas the STAT5s contains a PSP (Pro-Ser-Pro) motif. Except STAT2, all the B-type STATs contain PMSP (Pro-Met-Ser-Pro) motif. STAT2 is found in species like zebrafish or *Xenopus laevis*, which are distantly related to humans. The sequence analysis suggests that after the duplication of STAT gene and appearance of B-type STATs, the PMSP motif was acquired [23]. The available information suggests the STAT gene duplication and its functional diversification occurred to their evolutionary ancestors. This diversification process causes various functional role played by STAT in cell biology.

The STATs genes in human are located on chromosome 2 (STAT 1 and STAT 4), chromosome 12 (STAT 2 and STAT 6), and chromosome 17 (STAT 3, STAT 5a, and STAT 5b) with an average length of 750-850 amino acids and a size range from 90–115 kDa [24]. Although from a functional point of view the STATs proteins show different specific function, they share a common structure with remarkable homologies [25-27]. Six different domains are highly conserved within STATs, including an N-terminal domain (ND), coiled-coil domain (CCD), and the DNA-binding domain (DBD) located in the midst of the protein, the linker domain (LD), the SH2 domain, and, finally, a transcriptional activation domain (TAD) in the CTD. A schematic illustration of STATs structure is depicted in Fig. 2.

The ND is a highly-conserved area containing the amino acidic sequences responsible for protein-protein interactions which mediate oligomerization of STAT [28]. STAT dimers interaction is cooperative, and cease following of the ND deletion [29]. The CCD, with an amino acidic region comprised between 130 and 315, bridges the ND with the DBD. This domain, possessing alfa-helical structures, is generally responsible for interactions with regulatory proteins [30-32]. The DBD determines the recognition of specific DNA sequence elements for each STAT and is generally located in the region comprised between 320-475 amino acids [32,33]. From a structural perspective, it shows similarity to an immunoglobin-like fold with β sheets very close to the observed in NF-kB or p53 [32]. The LD, possessing mostly an alfahelical structure sits between the DNA binding domain and the SH2 domain and it is also implicated in the transcription by interaction with the transcriptional machinery of the cell. The SH2 domain is located in the amino acid region between 600 and 700 [34]. It plays a key role in recognizing phosphorylated tyrosine residues and, therefore, mediates the interplay between STATs and other phosphorylated proteins, such as growth factors, JAKs, and other members of the STATs family to form dimer [35,36]. [37,38]. Differences within the SH2 domains are responsible for selectivity towards different specific receptors. Nearby the SH2 domain is located the key tyrosine residue needed for STAT activation. In addition to Tyr phosphorylation necessary for dimerization and migration into the nucleus, phosphorylation a Ser residue, with the exception of STAT2 and 6, is also required to start transcriptional activity [39,40]. This conserved Ser residue, is located into the TAD in the STATs C-terminal region (amino acidic residues 661-851) [40,41].

STATs are very versatile transcription factors that modulate a number of activities of cellular growth, survival and differentiation [42,43]. STAT members also govern four main effector

responses, in particular STAT1, STAT2 and STAT4 are responsible for antiviral type1 response, STAT3 is responsible for antibacterial and antifungal type 3 response, STAT6 is responsible for anti-helminth type2 response, and STAT5 is important in the initiating inflammation processes [16]. Overall, STATs are crucial for optimal immune expression, otherwise could lead to immunodeficiency or autoimmune disorders [44,45]. Moreover, deregulation of STATs has also been reported to trigger angiogenesis and in turn promote tumors survival [46-50].

STAT1 and STAT2 have shown a crucial role in the host defense in case of hepatitis C virus (HCV) infection and interferon (IFN)-α treatment-induced HCV clearance [51,52]. The HCVinfected hepatocytes lead to the generation of IFN- β , as a result, STAT1 and STAT2 activation occurred through modulation of IFN- α/β receptor and thus cause the upstream regulation of several antiviral proteins in the surrounding uninfected environment that stop further proliferation of infection [53-57]. STAT1 mutations have been associated with a progressive multifocal leukoencephalopathy (PML) produced by the polyomavirus JC virus, which typically occurs only in immune-compromised patients [35]. In addition, the activity and expression of STAT1 is deregulated in both cancer and insulin resistance disorders [36]. STAT3 is considered a proto-oncogene constitutively activated in several human cancers and plays a crucial role in survival of tumor cells, cell proliferation, migration, metastasis and angiogenesis [58]. STAT3 has reported to be required for optimal function of the electron transport chain in a transcriptional-independent manner [59-62]. In STAT3-/- cells it has been evidenced defects in mitochondrial complexes I and II [60]. However, the mechanism by which STAT3 modulates complexes I and II remains to be elucidated. Moreover, STAT3 was reported to support oncogenic transformation that maintains mitochondrial respiratory chain activity and blocks the transition pore of mitochondrial permeability [63]. STAT3 over-expression also showed a direct

association with the development of arthritis [64,65]. STAT4 is distinct from other STAT member family since its expression is limited to T lymphocytes, myeloid cells and spermatozoa [66,67]. STAT4 was first observed to be activated in response to treatment with interleukin-12 (IL-12) of T cells [68] and studied for its role as central protective mediator during immune responses and immune-mediated diseases [69]. STAT5A and 5B share almost 95% of sequence identity and the functions of these proteins appear to be dependent on hormone and tissue in a context-dependent manner [70]. Recently, the participation of STAT5 signaling pathway in modulating maternal and feeding behaviors has been suggested [71]. STAT6 has been reported to be activated by IL-4 and IL-13 and modulates the expression of genes that regulate allergic inflammatory responses mediating the pathogenesis of allergic disorders such as asthma, atopic dermatitis and food allergy [72,73]. Additionally, a chimeric decoy oligodeoxynucleotides significantly down regulated lung inflammation through STAT6 modulation [74].

Finally, an association between diverse polymorphisms in STATs and the development of some inflammatory diseases has been reported. In this sense, a strong gene-gene interaction of STAT-4 T90089C, STAT-6 G2964A, and IFN- γ T874A has been found to cause an up-regulation of asthma in a Chinese population [75]. STATs gene polymorphisms STAT3 C4796793G and STAT5b C6503691T favor the appearance of rheumatic heart diseases and could be an effective biomarker for the prognosis of individuals with high risk of rheumatic heart diseases [76].

Cytokines, growth factors and hormones are the main activators of STATs family. STAT1 has been reported to be activated by IFN- α , IFN- β or IFN- γ and several interleukins (IL-6, IL-7, IL-10 and IL-27) in immune cells [77-82], and this STAT1 activation was reduced by Zn²⁺ chelation [77,78]. Recently, estrogen treatment has been also shown to up-regulate STAT1 mRNA expression in THP-1 and K562 cells [83]. In an animal model of injury-induced inflammation,

STAT2 mRNA levels were up-regulated in astrocytes in a process mediated by NF-κB signaling [84]. In human artery endothelial cells, oxidized low density lipoprotein (oxLDL) treatment was able to increase the STAT1/2 mRNA levels [85]. Cytokines such as IL-6, IL-1β, and TNFa increased STAT3 mRNA levels in pancreatic [86] and lung cells [87,88]. β-catenin/T-cell factor (TCF) pathway has evidenced to increase mRNA STAT3 expression human esophageal squamous carcinoma cells [89]. STAT3 was also increased in ob/ob livers after long-term 17βestradiol treatment [90]. STAT4 activation in mediated by IFN- α/β and by diverse interleukins (IL-2, IL-12, IL-13, and IL-17) [91-93]. STAT4 can also undergo epigenetic regulation by DNA hypermethylation, since higher levels of STAT4 mRNA were found after treatment with demethylating drugs in human primary T cells [94]. STAT5 is activated by prolactin, erythropoietin and growth factors [95]. Cytokines such as IL-2, IL-3, IL-4, IL-5, IL-6 and IL-7 could induce STAT5 A/B expression in different immune cell types [96-101]. STAT6 activation is mediated by IL-4 and IL-13 [102]. In fact, IL-4 facilitates the direct gene-specific demethylation in the differentiation of innate immune cells through a mechanisms that involve activation of STAT6 [103] and IL-13 acts as a central effector cytokine in ulcerative colitis by activating the STAT6 [104].

Recent studies have identified three molecules that negatively regulate JAK/STAT signaling including protein inhibitors of activated stats (PIAS), suppressors of cytokine signaling (SOCS), and protein tyrosine phosphatases (PTPs). PIAS, characterized by a zinc-binding ring-finger domain in the central portion, bind the activated STAT dimers preventing them from binding DNA, or promoting SUMOylation which can modulate STATs phosphorylation and dimerization [105,106]. SOCS proteins contain a central SH2 domain which directly binds to phosphorylated tyrosines of activated JAKs, causing the JAK kinase activity to be blocked,

preventing the recruitment of the STATs [105,107]. PTPs are a group of distinct proteins that dephosphorylate JAKs at the level of tyrosine residues reversing the JAK-STAT activity [108]. miRNA are important gene expression modulators, and several miRNA are found to control the expression of STAT family of transcription factors. In this context, Kohanbash and Okada summarized the reciprocal regulations between STATs and miRNA in the context of diseases [109].

STATs can also interact with other transcription factors (STAT co-activators) modulating gene transcription in an indirect way. Some transcription factor such as orphan nuclear hormone receptor, USF1 (upstream stimulatory factor-1), ZXDC (zinc finger X-linked duplicated family member), ETS (erythroblast transformation-specific), p300/cAMP-responsive element-binding protein (CBP), AP-1 (activator protein-1), and Sp1 (specificity protein 1) potently drive target gene expression of STAT1 [110,111]. The transcriptional activity of STAT3 can be enhanced by its association with HNF-3 (hepatocyte nuclear factor 3), NcoA/SRC1a (steroid receptor coactivator 1), GR (glucocorticoid receptor), Crif1 (CR6-interacting factor 1), Sp1, p300/CBP and c-jun [112,113]. The interaction of STAT5 with p300/CBP, high-mobility group N2 protein (HMGN2), CPAP (centrosomal P4.1-associated protein), Nmi (N-Myc interactor), Sp1, YY1 (Ying Yang-1), and C/EBPβ increase STAT5 target gene expression [114-116]. The transcriptional activation of STAT6 requires the interaction with p160/SRC (NCoA) coactivator family, C/EBP and p100 [117-119].

3. Role of STATs signaling in neuroinflammation

Microglia and astrocytes are the main CNS-resident cells responsible for the immune and inflammatory responses. Activation of these cells induces inflammation and the release of

multiple inflammatory mediators including cytokines and growth factors. In this scenario, the JAK-STAT pathways are implicated in the regulation of the inflammatory response since they are activated by some of these cytokines/growth factors but also by other inflammatory signals such as LPS, gangliosides and thrombin [120,121]. Upon exposure to pro-inflammatory signals (cytokines, cellular or bacterial debris), resident macrophages and microglia become polarized towards a pro-inflammatory M1 phenotype and release inflammatory mediators and chemokines, in a process mediated by STATs [122,123]. Under physiological conditions, when the CNS is damaged, the JAK-STAT pathway plays an important role in the neuronal regeneration process as well as in the formation of glia scars around the injured area. In accordance, it has been observed that after nerve injury, STAT3 expression is induced and activated by phosphorylation in regenerating neurons promoting the regrowth of damaged axons [124]. Moreover, the deletion of SOCS3 in diverse mutant mouse models of optical nerve injury induces the regeneration of injured axons [125]. Diverse cytokines and oxidative stress have been reported to activate mainly the STAT1 and STAT3 isoforms leading to the induction of inflammation-associated and survival-associated gene expression [126,127]. Although CNS inflammation is an important mechanism to protect the brain against injuries, when it occurs excessively or prolonged, inflammation can contribute to the appearance or worsen various diseases that affect the brain. Since the JAK-STATs mediate the inflammatory responses, a dysregulation of these pathways is related to neuroinflammation and neuroglial survival. In addition, depending of the STAT activated and the site of activation the effects on inflammation and cell survival could be not identical.

Although evidences report the participation of STAT3 in brain inflammation, the exact role is not clear and still controversial. The activation of this pathway results in the expression of genes

that sequentially repress the transcription of pro-inflammatory genes contributing to the resolution of the inflammatory process [128]. However, it has been evidenced that STAT3 is capable to promote cell apoptosis and brain damage or favor cell survival. Specifically, diverse studies have evidenced that STAT3 excessive activation contributes to neuronal damage, whereas in other studies the deleterious effects are associated with the disruption of STAT3 signaling [129-131]. On the contrary, the role of STAT1 is clearer as it induces cell death [132]. STAT3 activation also induces the expression of STAT3 itself, thus promoting a positive feedback loop of STAT3 that if not properly regulated can lead to neuronal damage [133]. It was also evidenced that STAT3 is necessary for the expression of miR-155 in cultured Th17 cells, indicating a function in the etiology of inflammatory disorders inducing the expansion of Th17 cells [134]. Mutations in STAT proteins that lead to excessive activation of the pathways are not frequent and the excessive activity is generally triggered by the overproduction of cytokines, and/or deregulation of endogenous negative modulators of the JAK/STAT [10,11]. The negative feedback molecules, especially SOCS1 and 3 that are expressed in immune cells and cells of the CNS, are an important point of interest since they act as suppressors of cytokine signaling, limiting the cellular cytokine response [135]. In accordance, many anti-inflammatory drugs, such as aspirin, exert some of their effects via SOCS proteins [136]. It has been reported that SOCS1 limits chemokine-induced migration of immune cells within the brain and ameliorates the release of chemokines and pro-inflammatory mediators by inhibiting the STAT and the NF-kB activation [137-139]. Similarly, SOCS3 inhibits the signalling of a wide range of immune molecules such as IL-6, IL-10, IFNs, LPS, among others [137,140,141].

It is interesting to mention that de-regulated JAK/STAT pathways are found in neurodegenerative disorders. For example, in a model of Parkinson's disease developed in rats

(by viral mediated overexpression of α -synuclein), the pharmacological inhibition of the JAK/STAT pathway by the JAK inhibitor AZD1480, significantly prevented neuroinflammation and neurodegeneration [142]. The activation of the JAK/STAT pathway in Parkinson's disease is related to the elevated levels of IFN- γ and IL-6 [142]. Multiple sclerosis is a devastating demyelinating disease that curses with chronic inflammation and neuronal degeneration. In patients with this pathology, it has been evidenced that immune cells from subjects suffering from relapsing-remitting multiple sclerosis express [141]more phosphorylated STAT3 during relapse than in remission [143]. Moreover, diverse studies concentrate on the induction of SOCS3, which leads to the inhibition of STAT1 and STAT3 in humans and in experimental autoimmune encephalomyelitis animal model [144,145]. On the contrary, in Alzheimer's disease the levels of phosphorylated STAT3 in hippocampal neurons were reduced in a mice model of Alzheimer's disease (Tg2576) and in Alzheimer's disease patients [146]. In this study, pharmacological inhibition of the JAK2/STAT3 pathway caused memory impairment in Tg2576 mice, whereas colivelin, a derivate of the neuroprotective peptide humanin, totally recovered cognitive function in an Alzheimer's disease model (Tg2576) via activation the JAK2/STAT3 pathway [146]. Also, nicotinic acetylcholine receptors have been reported to reduce AB neurotoxicity by activating the JAK2/STAT3 pathway [147].

In addition, neural damage resulting from infections, trauma and deposition of abnormal proteins like β -amyloid and transactive response (TAR) DNA binding protein, can induce secondary response such as neuroinflammation aggravating the neuronal impairment far more than the primary injury [148].

4. POLYPHENOLS

4.1 Direct inhibitors of STAT

Although diverse polyphenols have been evidenced diminish the ability of STAT to signal transcription of inflammatory genes, the most studied are quercetin, curcumin, resveratrol and luteolin (figure 3). Quercetin directly inhibits the phosphorylation of STAT and has been found effective in preventing apoptosis in dopaminergic cells through down regulation of pro-apoptotic genes like Bax and up-regulation of anti-apoptotic gene Bcl-2 [149]. Kumar and colleagues showed that quercetin possesses neuroprotective effects in diabetes induced neuro-degeneration and inflammation, partly due to its STAT inhibition [150]. Coupling quercetin with nanoparticles like β -cyclodextrin-dodecylcarbonate increases its permeability through blood-brain barrier making it a suitable drug for STAT intervention and prevention of neuroinflammation [151].

Curcumin also presents its anti-inflammatory properties partly by suppressing JAK/STAT pathway [152]. It has been shown to decrease activation of microglial cells by preventing STAT 1 and 3 phosphorylation indirectly through activation of SHP-2, a tyrosine phosphatase which negatively regulates JAK, thereby weakening the inflammatory action of microglia. [153]. Curcumin also suppresses Oncostatin M (a cytokine from interleukin 6 family) induced p-STAT1 [154]. Curcumin and 1,25 dihydroxyvitamin-D3 at dosages of 20 μ g/ml and 250 nM respectively, independently prevent the development of autoimmune encephalomyelitis, leading to multiple sclerosis (MS-EAE) by inhibiting the differentiation of neural antigen specific Th1 cells, through blockage of IL-12 mediated STAT3/4 activation [155,156].

Resveratrol is a wine-derived polyphenol that suppresses LPS (lipopolysaccharide) induced inflammation in part by inhibiting phosphorylation of STAT1/3 which mediates the release of pro-inflammatory cytokines [157]. It significantly inhibits the activation of STAT1 in a lower dose than 5-aminosalicylic acid (25µM for resveratrol vs 500 µM for 5-aminosalicylic acid) in

HT-29 cells [158] However, the co-treatment with the two inhibitors did not exert synergistic effect against the expression of pro-inflammatory mediators in cytokine-stimulated cells mediated by the JAK-STAT pathway [158]. However, resveratrol does not interfere with the MAPK mediated negative feedback mechanism of STAT showing its STAT1 specificity [158].

Luteolin supplementation leads to decreased IL-6, a microglial cell activator, indirectly suppressing STAT activation in children with autism spectrum disorders (ASD) which resulted in improvement in their communication and living skills [159], thus preventing microglial activation associated with an anti-inflammatory phenotype at dose of luteolin of 100 U/ml [160,161].

In addition to these four polyphenols, genistein, kaempferol and daidzein have been reported to inhibit the activation of STAT-1 and NF- κ B pathways [162]. Hämäläinen et al. [162], analyzed the effects of 36 naturally occurring flavonoids as anti-inflammatory agents. Within these compounds, the above-mentioned polyphenols were capable to inhibit the activation of STAT-1 in macrophages stimulated with LPS. A recent study also reported that kaempferol 7-*O*- β -D-glucoside, isolated from *Cudrania tricuspidata*, inhibits the phosphorylation of STAT-1 in LPS-stimulated macrophages [163].

Extracts from a number of herbal plants have been found to target JAK/STAT pathway, and could present as a therapeutic regime for neuroinflammation. *Agrimonia pilosa* has long been used in traditional Chinese medicine for its anti-inflammatory properties as it is rich in flavonoids like Agrimonolide, which at a dose of 80 µM has been found to inhibit LPS-induced JAK/STAT pathway directly by preventing phosphorylation of JAK1 and consequently STAT1 and 3, while indirectly by down regulating IL-6 expression [164]. Plumbagin, a compound extracted from *Plumbago zeylanica*, directly blocks the phosphorylation of STAT1/4 and STAT3

to downregulate differentiation of Th1 and Th17 cells respectively (at 2 mg/kg in experimental autoimmune encephalomyelitis-EAE), to control encephalitis [165]. Progression of disease is substantially controlled when plumbagin is administered pre-immunization in the model [165]. Similarly, tripchlorolide (T4) a constituent of *Tripterygium wilfordii Hook F*. extract reduces the disease severity of EAE in the same manner by blocking the JAK/STAT along with ERK1/2-NF-kB involved in Th1 and 17 proliferation at a mere dose of 40 µg/kg [166]. A compound extracted from *Cornus officinalis*, cornel iridoid glycoside (CIG) halts multiple sclerosis progression by suppressing the JAK/STAT1 and JAK/STAT3 and subsequently IL-6 in a dose dependent manner, also returning the microglial population to normal at 120 mg/kg in EAE animal model [167].

4.2. Indirect inhibitors of STAT

Berberis vulgaris L derived Berbamine extract indirectly stimulates STAT4 degradation through STAT-interacting LIM protein (SLIM) without affecting its phosphorylation, leading to decreased inflammatory cytokine assembly in EAE [168]. Berberine at the dose of 50 mg/kg targets JAK1/2 and STAT1/3/4 activation among many others to halt the differentiation of T helper cells Th1 and 17 in the EAE [169]. A novel caffeic acid derivative [(*E*)-2-cyano-*N*-[(*S*)-1-phenylethyl]-3-(pyridin-2-yl)acrylamide], which resembles in structure like AG490, a developed JAK inhibitor (also called Jakinibs), has been found to significantly down regulate levels of pSTAT3 and its dependent genes in glioma cells in a long term manner by acting on JAK1/2 activators, rendering low phosphorylation levels and inducing apoptosis at half the dose of AG490 (i.e., 25 μ M). AG490 at 50 μ M only slightly lowers p-JAK levels [170].

Sophora flavonone G extracted from *Sophora alopecuroides* partly exhibits its antineuroinflammatory properties by suppression of JAK1/2 and STAT1/3/5, their phosphorylation was significantly reduced in LPS induced inflamed microglial cells treated with the compound [171]. In the same manner, *Serenoa repens* is another phyto-therapeutic compound which along with inhibiting neuroinflammation by blocking the inflammation regulator (STAT3), also decreases growth of glioma cells; this effect of *Serenoa repens* is synergized by the JAK inhibitor AG490, where the co-treatment of the two compounds at 1 µl/ml and 20 nM respectively, markedly increased apoptotic processes in glioma cells [172]. Moreover, it down regulated both basal p-STAT and IL-6 induced p-STAT, facilitating the chemotherapeutic activity of docetaxel in treatment of myelomas by increasing apoptosis, however, lower cancer cell viability is also evidenced after co-treatment with docetaxel and AG490 [173].

5. DRUGS

Certain JAK/STAT inhibitory drugs have been approved by FDA for use in treatment of neuroinflammatory diseases while other potential suppressors are vigorously under investigation and could be developed as standard intervention strategy for inflammatory and autoimmune disorders of nervous system.

5.1. Direct STAT inhibiting drugs

Approved STAT inhibitors include laquinimod, glatiramer acetate, and fumarates. These halt multiple sclerosis progression by blocking STAT1/3 activation leading to decreased cytokine production and increased population of macrophages and type II dendritic cells; where they

additionally block the differentiation of Th1 and Th17 cells [174]. Laquinimod and fumarates have the potential to reverse EAE at the doses of 25 mg/kg and 70 μ M respectively [175,176].

AG490 is an established JAK inhibitor and is used for blocking JAK/STAT pathway in many inflammatory diseases including multiple sclerosis and cancers like the glioblastoma, where it down-regulates transcription of a number of inflammatory cytokines like IL-2, 6 and 12. In the LPS induced microglial apoptosis, AG490 (10 μ M) prevents cellular death and inflammation by preventing JAK2/STAT induced NO release and protects endothelial cell mediated blood brain barrier destruction [177]. It is also used as a standard when there is a need for comparing the JAK inhibitory potential of novel synthetic drugs and natural compounds under study.

Among others, Cucurbitacin-I (JSI-124) inhibits both JAK2/STAT3 directly by blocking STAT's phosphorylation via JAK and also preventing STAT dimerization [178]. Sorafenib and WP1066, both are known STAT inhibitors in CNS. Sorafenib at 10 μ M blocks the phosphorylation of STAT3 in glioma cells inducing apoptosis, the upstream JAK1 and 2 inhibition by Sorafenib is reversible by mutant STAT3, and its phosphatase activity is revoked by tyrosine phosphatase inhibitor [179]. A novel molecule structurally similar to AG490, WP1066, which at the concentration of 10 μ M significantly inhibits STAT3 and exhibits anti-cancer apoptotic properties in glioma cells and in vivo conditions, and is found to be almost 17 times more cytotoxic to U87-malignant glioma cell line than AG490 [180]. It also blocks JAK2/STAT3 to abolish growth of multiple brain tumor sphere cultures (BTSC) and their xenografts irrespective of the mutations they carry [181].

AZD1480 suppresses STAT1/4 in Th1 cells and STAT3 in Th17 cells of EAE models at 25 mg/kg to inhibit their differentiation and release of other pro-inflammatory cytokines thereby preventing neuronal inflammation in multiple sclerosis [182], it also target JAK/STAT pathway

to inhibit dopaminergic degeneration in PD cell line. Acute administration of 10 μ M AZD1480 exhibits anti-inflammatory properties by suppressing α -synuclein induced immune system activation [183]. AZD1480 supplied to 50 mg/kg induce apoptosis and reduce proliferation in human and murine glioblastoma tumor cells by suppressing basal and induced JAK1/2 and STAT3 [184].

A JAK3/STAT5 specific inhibitor ZM39923 (ZM) completely abolishes the activation of this pathway at 30 µM [185] and also inhibits the growth of brain tumor initiating cells (BTIC) at a dose of merely 1 µM by blocking JAK3/STAT3 [186]. The drug Pacritinib (also called as SB1518) is a JAK2/STAT3 inhibitor which significantly decreases cell viability and neurosphere formation in heterogeneous BITC populations at 5 µM concentration and increases the sensitivity of BITC cultures without affecting normal astrocytes in human [187]. A synthetic molecule ORL-NIH001 is a STAT3 suppresser which prevents the progression of inflammation in experimental uveitis by halting STAT mediated IL-6 expression which suppresses upstream differentiation of CD4⁺T cells into Th17 cells, also blocking Th17 proliferation, entry into neuroretina and subsequent release of inflammatory cytokines; it can also be used in the management of multiple sclerosis due to its Th17 inhibitory activity [188]. Similarly, another synthetic molecule BRD0476 has a novel mechanism of action against JAK/STAT pathway where it decrease the levels of IFN-y- induced p-STAT1 without affecting the kinase activity of JAK2 by inducing a competition between its phosphorylation and ubiquitination, where ubiquitination is performed by inducing deubiquitinase ubiquitin-specific peptidase 9X (USP9X) [189]. In methamphetamine induced neuro-inflammation models, Asiatic acid has the capability to return the methamphetamine induced abnormal phosphorylation of STAT3 by JAK2 to standard levels at 20 µM, blocking the expression and release of inflammatory IL-6 as the DNA binding activity

of STAT is also attenuated by Asiatic acid [190]. Pyridone 6 (P6) also inhibits the phosphorylation of JAK2/STAT3 and blocking the release of its mediated pro-inflammatory cytokines, specifically TNF α and the iNOS (inducible nitric oxide synthase) to attenuate the destructive function of activated microglial cells, which arise in neuroinflammation and various neuropathological conditions like Parkinson's disease and amyotrophic lateral sclerosis [191].

5.2. Indirect STAT inhibiting drugs

Tofacitinib, ruxolitinib and baricitinib are three developed JAK1/JAK2 inhibitors FDA approved for use in autoimmune and anti-inflammatory diseases like Rheumatoid arthritis, psoriasis and lupus; and could present as an effective therapeutic regime for encephalitis and other autoimmune inflammatory disorders of nervous system. Tofacitinib is a JAK1/3 inhibitor where ruxolitinib and baricitinib suppress the activity of JAK1/2 and their downstream STATs in the signaling cascade [192]. Ruxolitinib inhibits JAK/STAT to indirectly prevent oxidative stress-induced neuroinflammation which is potentiated by IL-13, and subsequent death of dopaminergic neurons in cultured cells and animal models of Parkinson's disease [193].

Ionomycin and PMA (phorbol 12-myristate 13-acetate) inhibits IL-6 induced STAT3 promptly by activating MAPK, ERK (extracellular signal regulated kinase) family of MPAK is antagonistic to STAT in certain conditions, which is reversible on the suppression of the latter by ERK specific kinase inhibitors. However, ionomycin and PMA have no effect on IFN- α stimulated STAT3 activation, showing their specificity and potential for use in IL-6 mediated inflammatory pathways like those involved in EAE, ASD, uveitis, etc. [194]. Similarly, phenylephrine at 100 µM can also reversibly inhibit STAT3 activation through MAPK dependent pathway [195].

Among other Jakinibs, tyrphostin B42 is a JAK2 specific inhibitor reducing MS severity by diminishing IL-12 induced JAK phosphorylation and thereby, reducing Th1 proliferation [196]. A new JAK3 specific inhibitor CP-690,550, has negative potency for JAK 1 and 2; and could be used therapeutically in multiple sclerosis for its ability to target IL-2 and 12 stimulated T cell populations [197]. JAK3 blockage sufficiently prevents against autoimmune and alloimmune actions [198]. Dimethoxy-quinazolines (WHI-P154 and WHI-P131), two novel quinazoline derivatives are JAK3 inhibitors, where the latter also blocks OSM induced STAT1 phosphorylation. In a familial ALS mouse model with Cu, Zn-superoxide dismutase (SOD1) mutation, which leads to ROS (reactive oxygen species) accumulation, causing STAT mediated release of pro-inflammatory cytokines, WHI-P131 treatment inhibits STAT and increases neuronal survival in transgenic mice and improves pathological condition [154,198].

Various other JAK inhibitors possessing affinity for specific JAK target molecule are being successfully tested in clinical trials (primary and advanced stages) of numerous cancers including those of nervous system where they prevent inflammation and increases apoptosis of malignant cells; these Jakinibs include but are not limited to TG101348 specific for JAK2, NS-018 (JAK2), XL019 (JAK2), CEP-701 (JAK2), CYT-387 (JAK1/2), LY2784544 (JAK2/STAT5), BMS-911543 (JAK2/STAT1), lestaurtinib (JAK2/3) [199-206]. Lestaurtinib is a blocker of STAT5 phosphorylation and its activity in CNS has led to its use in treatment of neuroblastoma [207].

6. Conclusions and future prospects

In conclusion, the role of STAT in brain inflammation is still controversial. In fact, on the one hand, many investigations showed that STAT3 activation contributes to neuronal damage, and, on the other hand, the disruption of STAT3 signaling resulted be linked to deleterious effects

[129-131]. In particular, deregulation of the JAK/STAT pathway found in neurodegenerative diseases, such as Parkinson's disease, Alzheimer's disease, and multiple sclerosis. Therefore, over the last few years, modulation of STATs caused by many natural and synthetic substances has become a hot topic in the treatment of human diseases including those linked to neuroinflammation. The mechanisms at the basis of the modulation of STATs are different and very complex being both direct and indirect interactions depending on the type of substance. In addition, growing evidence suggests the importance of JAK inhibitors possessing affinity for specific JAK target molecule. Some JAK inhibitors resulted to be effective in treating cancers inCNS where they decrease inflammation and increase apoptosis of malignant cells.

The natural substances, characterized by low adverse and secondary effects, and with the capability to exert specific effects on JAK/STAT should be evaluated and developed for the treatment neuroinflammatory disorders opening a new horizon of hope for patients. The intervention of JAK/STAT pathway by small molecules could be a feasible therapeutic choice against many neuroinflammatory diseases due to their low cytotoxicity, usually high target specificity, low IC₅₀, and sufficient potential of crossing the blood brain barrier.

Considering the encouraging data obtained so far, clinical trials are requested to show the effectiveness and use in the clinical practice of JAK/STAT pathway inhibitors for the treatment of inflammation-based neurodegenerative pathologies. In addition, considering that the mechanisms of action of these inhibitors still remain to be fully elucidated, further studies are strongly requested to understand these aspects.

Acknowledgements

The Indian authors gratefully acknowledge the Bioinformatics Infrastructure Facility provided by the Alagappa University (funded by Department of Biotechnology, Government of India; Grant No. BT/BI/25/015/2012). D. Balan sincerely acknowledges Alagappa University Research Fund (Ph.D./1424/AURF Fellowship/2017 dated 23.11.2017), Karaikudi, Tamil Nadu, India for providing Research Fellowship. S. Xu is a recipient of Career Development Award of American Heart Association (18CDA34110359). A. Sureda was supported by the Institute of Health Carlos III (Project CIBEROBN CB12/03/30038) and by Govern de les Illes Balears (AAEE001/2017).

References

B.L. Clossen, D.S. Reddy, Catamenial-like seizure exacerbation in mice with targeted ablation of extrasynaptic δgaba-a receptors in the brain, J. Neurosci. Res. 95(2017) 1906-1916.
 E. Croisier, L.B. Moran, D.T. Dexter, R.K. Pearce, M.B. Graeber, Microglial inflammation in the parkinsonian substantia nigra: Relationship to alpha-synuclein deposition, J.

Neuroinflamm. 2(2005)14.

3 R.M. Ransohoff, M.A. Brown, Innate immunity in the central nervous system, J. Clin. Invest. 122 (2012) 1164-1171.

J.N. Ihle, Stats: Signal transducers and activators of transcription, Cell 84 (1996) 331-334.
R. Sobota, M. Szwed, A. Kasza, M. Bugno, T. Kordula, Parthenolide inhibits activation of signal transducers and activators of transcription (stats) induced by cytokines of the il-6 family, Biochem. Biophysic. Res. Commun. 267 (2000)329-333.

6 C.V. Clevenger, Roles and regulation of stat family transcription factors in human breast cancer, Am. J. Pathol. 165 (2004) 1449-1460.

J.J. O'Shea, M. Gadina, R.D. Schreiber, Cytokine signaling in 2002: New surprises in the JAK/STAT pathway, Cell 109(2002)S121-S131.

8 D.S. Aaronson, C.M. Horvath, A road map for those who don't know JAK-STAT, Science 296(2002) 1653-1655.

9 J.S. Rawlings, K.M. Rosler, D.A. Harrison, The JAK/STAT signaling pathway, J. Cell Sci. 117(2004)1281-1283.

10 J.J. O'shea, R. Plenge, JAK and STAT signaling molecules in immunoregulation and immune-mediated disease, Immunity 36(2012)542-550.

11 E.N. Benveniste, Y. Liu, B.C. McFarland, H. Qin, Involvement of the janus kinase/signal transducer and activator of transcription signaling pathway in multiple sclerosis and the animal model of experimental autoimmune encephalomyelitis, J. Interferon Cytokine Res. 34(2014)577-588.

12 K. Welsch, J. Holstein, A. Laurence, K. Ghoreschi, Targeting JAK/STAT signalling in inflammatory skin diseases with small molecule inhibitors, Eur. J. Immunol. 47(2017) 1096-1107.

D.W. Dodington, H.R. Desai, M. Woo, JAK/STAT–emerging players in metabolism,Trends Endocrinol. Metab. 29(2018) 55-65

14 Z. Yan, S.A. Gibson, J.A. Buckley, H. Qin, E.N. Benveniste, Role of the jak/stat signaling pathway in regulation of innate immunity in neuroinflammatory diseases, Clin. Immunol. 189 (2018)4-13.

L. Chen, H. Deng, H. Cui, J. Fang, Z. Zuo, J. Deng, Y. Li, X. Wang, L. Zhao, Inflammatory responses and inflammation-associated diseases in organs, Oncotarget 9 (2018) 7204.

16 J.J. O'Shea, D.M. Schwartz, A.V. Villarino, M. Gadina, I.B. McInnes, A. Laurence, The jak-stat pathway: Impact on human disease and therapeutic intervention, Ann. Rev. Med. 66(2015)311-328.

17 S.G. Rane, E.P. Reddy, Janus kinases: Components of multiple signaling pathways, Oncogene 19(2000)5662.

J.G. Williams, STAT signalling in cell proliferation and in development, Curr. Opin.Genet. Dev. 10(2000)503-507.

19 J.A. Souza-Neto, S. Sim, G. Dimopoulos, An evolutionary conserved function of the jakstat pathway in anti-dengue defense, Proc. Natl. Acad. Sci. 106(2009)17841-17846.

20 M.P. Zeidler, E.A. Bach, N. Perrimon, The roles of the drosophila JAK/STAT pathway, Oncogene 19(2000)2598.

21 N.G. Copeland, D.J. Gilbert, C. Schindler, Z. Zhong, Z. Wen, J.E. Darnell, A.L.F. Mui, A. Miyajima, F.W. Quelle, J.N. Ihle, Distribution of the mammalian STAT gene family in mouse chromosomes, Genomics 29(1995)225-228.

22 C. Barillas-Mury, Y.S. Han, D. Seeley, F.C. Kafatos, Anopheles gambiae ag-stat, a new insect member of the STAT family, is activated in response to bacterial infection, EMBO J. 18(1999)959-967.

23 T. Decker, P. Kovarik, Serine phosphorylation of stats. Oncogene 19(2000) 2628.

24 J.N. Ihle, The STAT family in cytokine signaling, Curr Opin. Cell Biol. 13(2001)211-217.

25 X. Chen, U. Vinkemeier, Y. Zhao, D. Jeruzalmi, J.E. Darnell Jr, J. Kuriyan, Crystal structure of a tyrosine phosphorylated STAT-1 dimer bound to DNA, Cell 93(1998) 827-839.

26 U. Vinkemeier, I. Moarefi, J.E. Darnell, J. Kuriyan, Structure of the amino-terminal protein interaction domain of STAT-4, Science 279(1998)1048-1052.

27 S. Becker, B. Groner, C.W. Müller, Three-dimensional structure of the STAT3 β homodimer bound to DNA, Nature 394(1998)145-151.

28 S. John, U. Vinkemeier, E. Soldaini, J.E. Darnell, Jr., W.J. Leonard, The significance of tetramerization in promoter recruitment by STAT5, Mol. Cell. Biol. 19(1999)1910-1918.

29 X. Xu, Y.L. Sun, T. Hoey, Cooperative DNA binding and sequence-selective recognition conferred by the STAT amino-terminal domain, Science 273(1996) 794-797.

30 A. Begitt, T. Meyer, M. van Rossum, U. Vinkemeier, Nucleocytoplasmic translocation of STAT1 is regulated by a leucine-rich export signal in the coiled-coil domain, Proc. Natl. Acad. Sci. USA 97(2000)10418-10423.

31 T. Zhang, W.H. Kee, K.T. Seow, W. Fung, X. Cao, The coiled-coil domain of STAT3 is essential for its SH2 domain-mediated receptor binding and subsequent activation induced by epidermal growth factor and interleukin-6, Mol. Cell. Biol. 20(2000)7132-7139.

32 C.M. Horvath, STAT proteins and transcriptional responses to extracellular signals. Trends Biochem. Sci. 25(2000)496-502.

33 C.M. Horvath, Z. Wen, J.E. Darnell, Jr., A STAT protein domain that determines DNA sequence recognition suggests a novel DNA-binding domain, Genes Dev. 9(1995) 984-994.

34 S. Becker, B. Groner, C.W. Muller, Three-dimensional structure of the STAT3beta homodimer bound to DNA, Nature 394(1998)145-151.

35 K. Shuai, The STAT family of proteins in cytokine signaling, Prog. Biophys. Mol. Biol. 71(1999)405-422.

36 J.E. Darnell, Jr., STATs and gene regulation, Science 277(1997)1630-1635.

37 M. Chatterjee-Kishore, F. van den Akker, G.R. Stark, Association of STATs with relatives and friends, Trends Cell. Biol. 10 (2000) 106-111.

38 J. Bromberg, J.E. Darnell, Jr., The role of STATs in transcriptional control and their impact on cellular function, Oncogene 19(2000) 2468-2473.

39 Z. Wen, Z. Zhong, J.E. Darnell, Jr., Maximal activation of transcription by stat1 and stat3 requires both tyrosine and serine phosphorylation, Cell 82(1995)241-250.

40 X. Zhang, J. Blenis, H.C. Li, C. Schindler, S. Chen-Kiang, Requirement of serine phosphorylation for formation of STAT-promoter complexes, Science 267(1995) 1990-1994.

41 A. Breit, V. Besik, H.J. Solinski, S. Muehlich, E. Glas, S.J. Yarwood, T. Gudermann, Serine-727 phosphorylation activates hypothalamic STAT-3 independently from tyrosine-705 phosphorylation, Mol. Endocrinol. 29(2015)445-459.

42 T.J. Mitchell, S. John, Signal transducer and activator of transcription (STAT) signalling and t-cell lymphomas, Immunology 114(2005)301-312.

43 L.N. Heppler, D.A. Frank, Targeting oncogenic transcription factors: Therapeutic implications of endogenous stat inhibitors, Trends Cancer 3(2017)816-827.

44 D. Saleiro, L.C. Platanias, Intersection of mtor and stat signaling in immunity, Trends Immunol. 36(2015)21-29.

45 G. Donninelli, I. Sanseverino, C. Purificato, S. Gessani, M.C. Gauzzi, Dual requirement for stat signaling in dendritic cell immunobiology, Immunobiology 223(2018) 342-347.

46 S. Haricharan, Y. Li, STAT signaling in mammary gland differentiation, cell survival and tumorigenesis, Mol. Cell. Endocrinol. 382(2014)560-569.

47 V. Calò, M. Migliavacca, V. Bazan, M. Macaluso, M. Buscemi, N. Gebbia, A. Russo, STAT proteins: From normal control of cellular events to tumorigenesis, J. Cell. Physiol. 197(2003)157-168.

48 K. Linher-Melville, G. Singh, The complex roles of STAT3 and STAT5 in maintaining redox balance: Lessons from stat-mediated xct expression in cancer cells, Mol. Cell. Endocrinol. 451(2017)40-52.

49 X. Li, D. Wang, Q.C. Zhao, T. Shi, J. Chen, Resveratrol inhibited non–small cell lung cancer through inhibiting STAT-3 signaling. Am. J. Med. Sci 352(2016) 524-530.

50 K. Friedrich, H. Dolznig, X. Han, R. Moriggl, Steering of carcinoma progression by the yin/yang interaction of STAT1/STAT3, BioScience Trends 11(2017)1-8.

51 Y. Chen, B. Jiao, M. Yao, X. Shi, Z. Zheng, S. Li, L. Chen, ISG12A inhibits HCV replication and potentiates the anti-HCV activity of ifn- α through activation of the JAK/STAT signaling pathway independent of autophagy and apoptosis, Virus Res. 227(2017) 231-239.

52 B. Rehermann, Hepatitis c virus versus innate and adaptive immune responses: A tale of coevolution and coexistence, J. Cin. Invest. 119(2009)1745-1754.

53 K. Blaszczyk, H. Nowicka, K. Kostyrko, A. Antonczyk, J. Wesoly, H.A. Bluyssen, The unique role of STAT2 in constitutive and ifn-induced transcription and antiviral responses, Cytokine Growth Factor Rev. 29(2016)71-81.

54 B. Gao, H. Wang, F. Lafdil, D. Feng, STAT proteins - key regulators of anti-viral responses, inflammation, and tumorigenesis in the liver, J. Hepatol. 57(2012)430-441.

55 T. Marcello, A. Grakoui, G. Barba–Spaeth, E.S. Machlin, S.V. Kotenko, M.R. Macdonald, C.M. Rice, Interferons α and λ inhibit hepatitis C virus replication with distinct signal transduction and gene regulation kinetics, Gastroenterology 131(2006)1887-1898.

56 E. Serti, H. Park, M. Keane, A.C. O'keefe, E. Rivera, T.J. Liang, M. Ghany, B. Rehermann, Rapid decrease in hepatitis C viremia by direct acting antivirals improves the natural killer cell response to IFNα. Gut 66(2017)724-735.

57 T. Miyagi, T. Takehara, K. Nishio, S. Shimizu, K. Kohga, W. Li, T. Tatsumi, N. Hiramatsu, T. Kanto, N. Hayashi, Altered interferon- α -signaling in natural killer cells from patients with chronic hepatitis C virus infection, J. Hepatol. 53(2010)424-430.

58 P.A. Johnston, J.R. Grandis, STAT3 signaling: Anticancer strategies and challenges, Mol.Intervent. 11(2011)18.

59 P. Tammineni, C. Anugula, F. Mohammed, M. Anjaneyulu, A.C. Larner, N.B.V. Sepuri, The import of the transcription factor stat3 into mitochondria depends on grim-19, a component of the electron transport chain, J. Biol. Chem. 288(2013) 4723-4732.

J. Wegrzyn, R. Potla, Y.J. Chwae, N.B. Sepuri, Q. Zhang, T. Koeck, M. Derecka, K. Szczepanek, M. Szelag, A. Gornicka, Function of mitochondrial stat3 in cellular respiration, Science 323(2009)793-797.

61 Z. Wen, Z. Zhong, J.E. Darnell Jr, Maximal activation of transcription by statl and stat3 requires both tyrosine and serine phosphorylation, Cell 82(1995) 241-250.

M. Demaria, C. Giorgi, M. Lebiedzinska, G. Esposito, L. D'Angeli, A. Bartoli, D.J. Gough, J. Turkson, D.E. Levy, C.J. Watson, A STAT3-mediated metabolic switch is involved in tumour transformation and STAT3 addiction, Aging (Albany NY) 2010;2:823.

63 D.J. Gough, A. Corlett, K. Schlessinger, J. Wegrzyn, A.C. Larner, D.E. Levy, Mitochondrial STAT3 supports ras-dependent oncogenic transformation, Science 324(2009)1713-1716.

64 R. Goswami, M.H. Kaplan, Chapter four - STAT transcription factors in t cell control of health and disease. In: Galluzzi L, ed. International review of cell and molecular biology: Academic Press, 2017: pp 123-180.

31

65 S.F. Ahmad, M.A. Ansari, A. Nadeem, K.M.A. Zoheir, S.A. Bakheet, A.M.S. Alsaad, Al-O.A. Shabanah, S.M. Attia, STA-21, a STAT-3 inhibitor, attenuates the development and progression of inflammation in collagen antibody-induced arthritis, Immunobiology 222(2017)206-217.

66 B. Gao, H. Wang, F. Lafdil, D. Feng, STAT proteins–key regulators of anti-viral responses, inflammation, and tumorigenesis in the liver. J. Hepatol. 57(2012)430-441.

67 N.C. Reich, STATS get their move on, JAK-STAT 2(2013)e27080.

N.G. Jacobson, S.J. Szabo, R.M. Weber-Nordt, Z. Zhong, R.D. Schreiber, J. Darnell,
K.M. Murphy, Interleukin 12 signaling in t helper type 1 (th1) cells involves tyrosine
phosphorylation of signal transducer and activator of transcription (STAT) 3 and STAT4, J Exp.
Med. 181(1995)1755-1762.

69 A. Martinez, J. Varade, A. Marquez, M. Cenit, L. Espino, N. Perdigones, J. Santiago, M. Fernández-Arquero, H. De La Calle, R. Arroyo, Association of the STAT4 gene with increased susceptibility for some immune-mediated diseases, Arthritis Rheum.58(2008)2598-2602.

70 P.M. Grimley, F. Dong, H. Rui, STAT5a and STAT5b: Fraternal twins of signal transduction and transcriptional activation, Cytokine Growth factor Rev. 10(1999)131-157.

71 I.C. Furigo, A.M. Ramos-Lobo, R. Frazão, J. Donato Jr, Brain stat5 signaling and behavioral control, Mol. Cell. Endocrinol. 438(2016)70-76.

P. Krishnamurthy, M.H. Kaplan, STAT6 and PARP family members in the development of t cell-dependent allergic inflammation, Immune network 16(2016)201-210.

Y. Liu, H. Zhang, R. Ni, W.Q. Jia, Y.Y. Wang, Il-4r suppresses airway inflammation in bronchial asthma by inhibiting the IL-4/STAT6 pathway. Pulm. Pharmacol. Ther. 43(2017)32-38.

74 T. Miyake, T. Miyake, M. Sakaguchi, H. Nankai, T. Nakazawa, R. Morishita, Prevention of asthma exacerbation in a mouse model by simultaneous inhibition of NF-κB and STAT6 activation using a chimeric decoy strategy. Mol. Ther. - Nucleic Acids 10(2018)159-169.

75 Y. Li, B. Wu, H. Xiong, C. Zhu, L. Zhang, Polymorphisms of STAT-6, STAT-4 and IFN- γ genes and the risk of asthma in chinese population, Respirat. Med. 101(2007)1977-1981.

U. Gupta, S.S. Mir, A. Srivastava, N. Garg, S.K. Agarwal, S. Pande, B. Mittal, Signal transducers and activators of transcription (STATs) gene polymorphisms related with susceptibility to rheumatic heart disease in north indian population, Immunol. Lett. 161(2014)100-105.

A. Brieger, L. Rink, H. Haase, Differential regulation of TLR-dependent MyD88 and TRIF signaling pathways by free zinc ions, J. Immunol. 191(2013)1808-1817.

78 C. Reiber, A. Brieger, G. Engelhardt, S. Hebel, L. Rink, H. Haase, Zinc chelation decreases IFN-β-induced STAT1 upregulation and inos expression in raw 264.7 macrophages, J. Trace Elem. Med. Biol. 2017;44:76-82.

79 K. Yuasa, T. Hijikata, Distal regulatory element of the stat1 gene potentially mediates positive feedback control of stat1 expression, Genes to Cells 21(2016)25-40.

80 C. Le Saout, M.A. Luckey, A.V. Villarino, M. Smith, R.B. Hasley, T.G. Myers, H. Imamichi, J.H. Park, J.J. O'Shea, H.C. Lane, IL-7–dependent STAT1 activation limits homeostatic CD4+ T cell expansion, JCI insight 2(2017)

Y.S. Choi, D. Eto, J.A. Yang, C. Lao, S. Crotty, Cutting edge: STAT1 is required for IL6-mediated BCl6 induction for early follicular helper cell differentiation, J. Immunol.
190(2013)3049-3053.

33

X. Ma, W. Yan, H. Zheng, Q. Du, L. Zhang, Y. Ban, N. Li, F. Wei, Regulation of IL-10 and IL-12 production and function in macrophages and dendritic cells, F1000Research 4(2015)

N.A. Young, G.R. Valiente, J.M. Hampton, L.C. Wu, C.J. Burd, W.L. Willis, M. Bruss,
H. Steigelman, M. Gotsatsenko, S.A. Amici, Estrogen-regulated STAT1 activation promotes
TLR8 expression to facilitate signaling via microRNA-21 in systemic lupus erythematosus, Clin.
Immunol. 176(2017)12-22.

R. Khorooshi, A.A. Babcock, T. Owens, NF-κB-driven STAT2 and CCL2 expression in astrocytes in response to brain injury, J. Immunol.181(2008)7284-7291.

A. Laguna-Fernández, S. Novella, C. Bueno-Betí, J. Marrugat, C. Hermenegildo, Endothelial transcriptomic changes induced by oxidized low density lipoprotein disclose an upregulation of JAK–STAT pathway, Vasc. Pharmacol. 73(2015)104-114.

L.C. Vona-Davis, K.A. Frankenberry, U. Waheed, E. Peterson, D.W. McFadden, Expression of STAT3 and SOCS3 in pancreatic acinar cells1, 2, J. Surg. Res. 127(2005)14-20.

M. Severgnini, S. Takahashi, L.M. Rozo, R.J. Homer, C. Kuhn, J.W. Jhung, G. Perides,
M. Steer, P.M. Hassoun, B.L. Fanburg, Activation of the STAT pathway in acute lung injury,
Am. J. Physiol. Lung Cell. Mol. Physiol. 286(2004)L1282-L1292.

X. Zhang, P. Yin, D. Di, G. Luo, L. Zheng, J. Wei, J. Zhang, Y. Shi, J. Zhang, N. Xu, IL-6 regulates MMP-10 expression via JAK2/STAT3 signaling pathway in a human lung adenocarcinoma cell line, Anticancer Res. 29(2009)4497-4501.

89 S. Yan, C. Zhou, W. Zhang, G. Zhang, X. Zhao, S. Yang, Y. Wang, N. Lu, H. Zhu, N. Xu, B-catenin/TCF pathway upregulates STAT3 expression in human esophageal squamous cell carcinoma, Cancer Lett. 271(2008)85-97.

34

90 H. Gao, G. Bryzgalova, E. Hedman, A. Khan, S. Efendic, J.A.K. Gustafsson, K. Dahlman-Wright, Long-term administration of estradiol decreases expression of hepatic lipogenic genes and improves insulin sensitivity in ob/ob mice: A possible mechanism is through direct regulation of signal transducer and activator of transcription 3. Mol. Endocrinol. 20(2006)1287-1299.

91 S.R. Gonzales-van Horn, L.D. Estrada, N.S. Oers, J.D. Farrar, STAT4-mediated transcriptional repression of the il5 gene in human memory th2 cells, Eur. J. Immunol. 46(2016)1504-1510.

92 S.J. Szabo, B.M. Sullivan, S.L. Peng, L.H. Glimcher, Molecular mechanisms regulating Th1 immune responses, Ann. Rev. Immunol. 21(2003)713-758.

93 Y. Wang, A. Qu, H. Wang, Signal transducer and activator of transcription 4 in liver diseases, Int. J. Biol. Sci.11(2015)448.

H.J. Shin, H.Y. Park, S.J. Jeong, H.W. Park, Y.K. Kim, S.H. Cho, Y.Y. Kim, M.L. Cho,
H.Y. Kim, K.U. Min, STAT4 expression in human t cells is regulated by DNA methylation but not by promoter polymorphism, J. Immunol. 175(2005)7143-7150.

F. Gouilleux, C. Pallard, I. Dusanter-Fourt, H. Wakao, L. Haldosen, G. Norstedt, D. Levy,
B. Groner, Prolactin, growth hormone, erythropoietin and granulocyte-macrophage colony stimulating factor induce MGF-STAT5 DNA binding activity, EMBO J. 14(1995)2005-2013.

96 W. Liao, D.E. Schones, J. Oh, Y. Cui, K. Cui, T.Y. Roh, K. Zhao, W.J. Leonard, Priming for T helper type 2 differentiation by interleukin 2–mediated induction of interleukin 4 receptor α-chain expression, Nat. Immunol. 9(2008)1288.

97 Z.S. Nagy, J.A. Ross, G. Rodriguez, B.L. Balint, L. Szeles, L. Nagy, R.A. Kirken, Genome wide mapping reveals PDE4B as an IL-2 induced STAT5 target gene in activated human pbmcs and lymphoid cancer cells, PloS one 8(2013)e57326.

J. Lee, S. Seong, J.H. Kim, K. Kim, I. Kim, B.C. Jeong, K.I. Nam, K.K. Kim, L. Hennighausen, N. Kim, STAT5 is a key transcription factor for IL-3-mediated inhibition of rankl-induced osteoclastogenesis, Sci. Rep. 6(2016) 30977.

99 B.A. Stout, M.E. Bates, L.Y. Liu, N.N. Farrington, P.J. Bertics, IL-5 and granulocytemacrophage colony-stimulating factor activate STAT3 and STAT5 and promote PIM-1 and cyclin d3 protein expression in human eosinophils, J. Immunol. 173(2004)6409-6417.

100 A.J. Tormo, M.C. Letellier, M. Sharma, G. Elson, S. Crabé, J.F. Gauchat, IL-6 activates STAT5 in T cells, Cytokine 60(2012)575-582.

101 M. Mandal, S.E. Powers, M. Maienschein-Cline, E.T. Bartom, K.M. Hamel, B.L. Kee, A.R. Dinner, M.R. Clark, Epigenetic repression of the igk locus by STAT5-mediated recruitment of the histone methyltransferase EZH2, Nat. Immunol. 12(2011)1212.

D. Hebenstreit, G. Wirnsberger, J. Horejs-Hoeck, A. Duschl, Signaling mechanisms, interaction partners, and target genes of STAT6, Cytokine Growth Factor Rev. 17(2006)173-188.
R. Vento-Tormo, J. Rodríguez-Ubreva, L. de la Rica, J.M. Urquiza, B.M. Javierre, R. Sabarinathan, A. Luque, M. Esteller, J.M. Aran, D. Álvarez-Errico, IL-4 orchestrates STAT6-mediated DNA demethylation leading to dendritic cell differentiation, Genome Biol. 2016;17:4.

P. Dames, T. Bergann, A. Fromm, R. Bücker, C. Barmeyer, S.M. Krug, M. Fromm, J.D. Schulzke, Interleukin-13 affects the epithelial sodium channel in the intestine by coordinated modulation of STAT6 and p38 mapk activity, J. Physiol. 593(2015)5269-5282.

105 L.N. Heppler, D.A. Frank, Targeting oncogenic transcription factors: Therapeutic implications of endogenous stat inhibitors, Trends Cancer 3(2017)816-827.

106 J. Morales, Y. Falanga, A. Depcrynski, J. Fernando, J. Ryan, Mast cell homeostasis and the JAK–STAT pathway, Genes Immun.11(2010)599.

107 B.T. Kile, B.A. Schulman, W.S. Alexander, N.A. Nicola, H.M. Martin, D.J. Hilton, The socs box: A tale of destruction and degradation, Trends Biochem. Sci. 27(2002)235-241.

108 T. Kisseleva, S. Bhattacharya, J. Braunstein, C. Schindler, Signaling through the JAK/STAT pathway, recent advances and future challenges, Gene 285(2002)1-24.

109 G. Kohanbash, H. Okada, Micrornas and stat interplay, Semin. Cancer Biol. 22(2012)70-75.

110 D.P. Beiting, S. Hidano, J.E. Baggs, J.M. Geskes, Q.Fang, E.J. Wherry, C.A. Hunter, D.S. Roos, S. Cherry, The orphan nuclear receptor TLX is an enhancer of STAT1-mediated transcription and immunity to toxoplasma gondii, PLoS Biol. 13(2015)e1002200.

111 G. Wang, Y. Wang, M. Teng, D. Zhang, L. Li, Y. Liu, Signal transducers and activators of transcription-1 (STAT1) regulates microrna transcription in interferon γ -stimulated hela cells, PLoS One 5(2010)e11794.

112 M.C. Kwon, B.K. Koo, J.S. Moon, Y.Y. Kim, K.C. Park, N.S. Kim, M.Y. Kwon, M.P. Kong, K.J. Yoon, S.K. Im, CRIF1 is a novel transcriptional coactivator of STAT3, EMBO J. 27(2008)642-653.

113 G. Waris, A. Siddiqui, Interaction between STAT-3 and HNF-3 leads to the activation of liver-specific hepatitis b virus enhancer 1 function, J. Virol. 76(2002)2721-2729.

114 A.A. Fiorillo, T.R. Medler, Y.B. Feeney, Y. Liu, K.L. Tommerdahl, C.V. Clevenger, HMGN2 inducibly binds a novel transactivation domain in nuclear prlr to coordinate stat5amediated transcription, Mol. Endocrinol. 25(2011)1550-1564.

115 K. Paukku, O. Silvennoinen, STATS as critical mediators of signal transduction and transcription: Lessons learned from STAT5, Cytokine Growth Factor Rev. 15(2004)435-455.

116 E. Pfitzner, R. Jahne, M. Wissler, E. Stoecklin, B. Groner, P300/CREB-binding protein enhances the prolactin-mediated transcriptional induction through direct interaction with the transactivation domain of STAT5, but does not participate in the STAT5-mediated suppression of the glucocorticoid response, Mol. Edocrinol. 12(1998)1582-1593.

117 L. Bao, J.B. Alexander, H. Zhang, K. Shen, L.S. Chan, Interleukin-4 downregulation of involucrin expression in human epidermal keratinocytes involves STAT6 sequestration of the coactivator CREB-binding protein, J. Interferon Cytokine Res. 36(2016)374-381.

118 T. Münz, C.M. Litterst, E. Pfitzner, Interaction of STAT6 with its co-activator SRC-1/NCOA-1 is regulated by dephosphorylation of the latter via PP2A, Nucleic Acids Res. 39(2010)3255-3266.

119 X. Wang, X. Liu, J. Fang, Y. Lu, J. He, X. Yao, Z. Yao, J. Yang, Coactivator p100 protein enhances STAT6-dependent transcriptional activation but has no effect on STAT1-mediated gene transcription, Anatom. Rec. 293(2010)1010-1016.

120 O.S. Kim, E.J. Park, E.H. Joe, I. Jou, JAK-STAT signaling mediates gangliosidesinduced inflammatory responses in brain microglial cells, J. Biol. Chem. 277(2002) 40594-40601.

121 C.S. Nicolas, M. Amici, Z.A. Bortolotto, A. Doherty, Z. Csaba, A. Fafouri, P. Dournaud,P. Gressens, G.L. Collingridge, S. Peineau, The role of JAK-STAT signaling within the CNS.JAKSTAT 2(2013)e22925.

122 G. Brown, Mechanisms of inflammatory neurodegeneration: Inos and nadph oxidase. Portland Press Limited, 2007.

123 M.D. Burton, N.L. Sparkman, R.W. Johnson, Inhibition of interleukin-6 trans-signaling in the brain facilitates recovery from lipopolysaccharide-induced sickness behavior, J. neuroinflammation 8(2011)54.

124 F.W. Schwaiger, A. Horvat, G. Hager, R. Streif, C. Spitzer, S. Gamal, S. Breuer, G.A. Brook, W. Nacimiento, G.W. Kreutzberg, Peripheral but not central axotomy induces changes in janus kinases (JAK) and signal transducers and activators of transcription (STAT), Eur. J. Neurosci. 12(2000)1165-1176.

P.D. Smith, F. Sun, K.K. Park, B. Cai, C. Wang, K. Kuwako, I. Martinez-Carrasco, L.
Connolly, Z. He, SOCS3 deletion promotes optic nerve regeneration *in vivo*, Neuron 64(2009)
617-623.

126 G. Regis, S. Pensa, D. Boselli, F. Novelli, V. Poli, Ups and downs: The stat1: STAT3 seesaw of interferon and gp130 receptor signalling, Semin. Cell. Dev. Biol. 19(2008) 351-359.

127 A. Planas, R. Gorina, A. Chamorro, Signalling pathways mediating inflammatory responses in brain ischaemia. Portland Press Limited, 2006.

128 P.J. Murray, Understanding and exploiting the endogenous interleukin-10/stat3-mediated anti-inflammatory response, Curr. Opin. Pharmacol. 6(2006)379-386.

129 I. Satriotomo, K.K. Bowen, R. Vemuganti, JAK2 and STAT3 activation contributes to neuronal damage following transient focal cerebral ischemia, J. Neurochem. 98(2006)1353-1368.

130 J.E. Jung, G.S. Kim, P. Narasimhan, Y.S. Song, P.H. Chan, Regulation of mn-superoxide dismutase activity and neuroprotection by STAT3 in mice after cerebral ischemia, J. Neurosci. 29(2009)7003-7014.

131 T.A. Sarafian, C. Montes, T. Imura, J. Qi, G. Coppola, D.H. Geschwind, M.V. Sofroniew, Disruption of astrocyte STAT3 signaling decreases mitochondrial function and increases oxidative stress in vitro, PloS one 5(2010)e9532.

132 J.H. Woo, J.H. Lee, H. Kim, S.J. Park, E.H. Joe, I. Jou, Control of inflammatory responses: A new paradigm for the treatment of chronic neuronal diseases, Exp. Neurobiol. 24(2015) 95-102.

133 A.P. Hutchins, S. Poulain, D. Miranda-Saavedra, Genome-wide analysis of stat3 binding in vivo predicts effectors of the anti-inflammatory response in macrophages, Blood 119(2012)e110-e119.

134 T. Escobar, C.R. Yu, S.A. Muljo, C.E. Egwuagu, STAT3 activates miR-155 in Th17 cells and acts in concert to promote experimental autoimmune uveitis, Invest. Ophthalmol. Vis. Sci. 54(2013)4017-4025.

135 B.A. Croker, H. Kiu, S.E. Nicholson, SOCS regulation of the JAK/STAT signalling pathway, Semin. Cell. Dev. Biol. 19(2008) 414-422.

F.S. Machado, J.E. Johndrow, L. Esper, A. Dias, A. Bafica, C.N. Serhan, J. Aliberti,
Anti-inflammatory actions of lipoxin a 4 and aspirin-triggered lipoxin are socs-2 dependent, Nat.
Med. 12(2006)330-334.

137 H. Qin, S.A. Niyongere, S.J. Lee, B.J. Baker, E.N. Benveniste, Expression and functional significance of SOCS-1 and SOCS-3 in astrocytes, J. Immunol. 181(2008)3167-3176.

138 G.M. O'Keefe, V.T. Nguyen, L.P. Tang, E.N. Benveniste, IFN-gamma regulation of class II transactivator promoter IV in macrophages and microglia: involvement of the suppressors of cytokine signaling-1 protein, J. Immunol. 166(2001) 2260-2269.

D.R. Wesemann, Y. Dong, G.M. O'Keefe, V.T. Nguyen, E.N. Benveniste, Suppressor of cytokine signaling 1 inhibits cytokine induction of CD40 expression in macrophages, J. Immunol. 169(2002)2354-2360.

140 H. Qin, C.A. Wilson, S.J. Lee, E.N. Benveniste, H. Qin, C.A. Wilson, S.J. Lee, E.N. Benveniste, Ifn- β -induced socs-1 negatively regulates cd40 gene expression in macrophages and microglia, FASEB J 20(2006)985-987.

141 J. Elliott, J.A. Johnston, SOCS: Role in inflammation, allergy and homeostasis, Trends Immunol. 25(2004)434-440.

H. Qin, J.A. Buckley, X. Li, Y. Liu, T.H. Fox, G.P. Meares, H. Yu, Z. Yan, A.S. Harms,
Y. Li, Inhibition of the JAK/STAT pathway protects against α-synuclein-induced neuroinflammation and dopaminergic neurodegeneration, J. Neurosci. 36(2016;36:)5144-5159.

E. Jakkula, V. Leppä, A.M. Sulonen, T. Varilo, S. Kallio, A. Kemppinen, S. Purcell, K.
Koivisto, P. Tienari, M.L. Sumelahti, Genome-wide association study in a high-risk isolate for multiple sclerosis reveals associated variants in STAT3 gene, Am. J. Hum. Genet. 86(2010)285-291.

144 H. Qin, W.I. Yeh, P. De Sarno, A.T. Holdbrooks, Y. Liu, M.T. Muldowney, S.L. Reynolds, L.L. Yanagisawa, T.H. Fox, K. Park, Signal transducer and activator of transcription-3/suppressor of cytokine signaling-3 (STAT3/SOCS3) axis in myeloid cells regulates neuroinflammation, Proc. Natl. Acad. Sci. 109(2012)5004-5009.

41

O. Norkina, A. Dolganiuc, D. Catalano, K. Kodys, P. Mandrekar, A. Syed, M. Efros, G. Szabo, Acute alcohol intake induces socs1 and socs3 and inhibits cytokine-induced stat1 and stat3 signaling in human monocytes, Alcohol. Clin. Exp. Res. 32(2008) 1565-1573.

146 T. Chiba, M. Yamada, S. Aiso, Targeting the JAK2/STAT3 axis in alzheimer's disease, Exp. Opin. Ther. Targets 13(2009)1155-1167.

147 S.D. Buckingham, A.K. Jones, L.A. Brown, D.B. Sattelle, Nicotinic acetylcholine receptor signalling: Roles in alzheimer's disease and amyloid neuroprotection, Pharmacol. Rev. 61(2009) 39-61.

148 C.K. Glass, K. Saijo, B. Winner, M.C. Marchetto, F.H. Gage, Mechanisms underlying inflammation in neurodegeneration, Cell 140(2010)918-934.

149 J. Bournival, P. Quessy, M.G. Martinoli, Protective effects of resveratrol and quercetin against MPP+ -induced oxidative stress act by modulating markers of apoptotic death in dopaminergic neurons, Cell. Mol. Neurobiol. 29(2009)1169-1180.

B. Kumar, S.K. Gupta, T.C. Nag, S. Srivastava, R. Saxena, K.A. Jha, B.P. Srinivasan, Retinal neuroprotective effects of quercetin in streptozotocin-induced diabetic rats, Exp. Eye Res. 125(2014)193-202.

G. Testa, P. Gamba, U. Badilli, S. Gargiulo, M. Maina, T. Guina, S. Calfapietra, F. Biasi ,R. Cavalli, G. Poli, G. Leonarduzzi, Loading into nanoparticles improves quercetin's efficacy in preventing neuroinflammation induced by oxysterols, PloS One 9(2014)e96795.

152 P. Rangarajan, A. Karthikeyan, S.T. Dheen, Role of dietary phenols in mitigating microglia-mediated neuroinflammation, Neuromol. Med. 18(2016)453-464.

153 P.E. Kim HY, E.H. Joe, I. Jou, Curcumin suppresses janus kinase-stat inflammatory signaling through activation of src homology 2 domain-containing tyrosine phosphatase 2 in brain microglia, J. Immunol. 171(2003)6072-6079.

154 W.Q. Li, F. Dehnade, M. Zafarullah, Oncostatin m-induced matrix metalloproteinase and tissue inhibitor of metalloproteinase-3 genes expression in chondrocytes requires janus kinase/stat signaling pathway, J. Immunol. 166(2001)3491-3498.

155 G. Muthian, H.P. Raikwar, J. Rajasingh, J.J. Bright, 1,25 dihydroxyvitamin-D3 modulates JAK-STAT pathway in IL-12/ifngamma axis leading to th1 response in experimental allergic encephalomyelitis, J. Neurosci. Res. 83(2006)1299-1309.

B.J. Natarajan C, Curcumin inhibits experimental allergic encephalomyelitis by blocking
IL-12 signaling through janus kinase-stat pathway in T lymphocytes, J. Immunol.
168(2002)6506-6513.

157 C. Ma, Y. Wang, L. Dong, M. Li, W. Cai, Anti-inflammatory effect of resveratrol through the suppression of NF-kappaB and JAK/STAT signaling pathways, Acta Biochim. Biophys. Sin. (Shanghai) 47(2015)207-213.

D. Serra, A.T. Rufino, A.F. Mendes, L.M. Almeida, T.C. Dinis, Resveratrol modulates cytokine-induced JAK/STAT activation more efficiently than 5-aminosalicylic acid: An in vitro approach, PLoS One 9(2014)e109048.

159 I. Tsilioni, A. Taliou, K. Francis, T.C. Theoharides, Children with autism spectrum disorders, who improved with a luteolin-containing dietary formulation, show reduced serum levels of tnf and IL-6, Transl. Psychiatr. 5(2015)e647.

160 K. Rezai-Zadeh, J. Ehrhart, Y. Bai, P.R. Sanberg, P. Bickford, J. Tan, R.D. Shytle, Apigenin and luteolin modulate microglial activation via inhibition of stat1-induced cd40 expression, J. Neuroinflamm. 5(2008)41.

161 E. Parker-Athill, D. Luo, A. Bailey, B. Giunta, J. Tian, R.D. Shytle, T. Murphy, G. Legradi, J. Tan, Flavonoids, a prenatal prophylaxis via targeting JAK2/STAT3 signaling to oppose II-6/MIA associated autism, J. Neuroimmunol. 217(2009)20-27.

M. Hämäläinen, R. Nieminen, P. Vuorela, M. Heinonen, E. Moilanen, Anti-inflammatory effects of flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF-kappaB activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF-kappaB activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages, Mediators Inflamm. 2007(2007)45673.

163 S.B. Lee, J.S. Shin, H.S. Han, H.H. Lee, J.C. Park, K.T. Lee, Kaempferol 7-O- β -D-glucoside isolated from the leaves of Cudrania tricuspidata inhibits LPS-induced expression of pro-inflammatory mediators through inactivation of NF- κ B, AP-1, and JAK-STAT in RAW 264.7 macrophages, Chem. Biol. Interact. 284 (2018)101-111.

164 T.H. Chen L, T. Fang, J. Xiao, Agrimonolide from *Agrimonia pilosa* suppresses inflammatory responses through down-regulation of COX-2/iNOS and inactivation of NF-κB in lipopolysaccharide-stimulated macrophages, Phytomedicine 23(2016) 846-855.

165 J.J. Jia Y, Y. Bai, Z. Li, L. Liu, J. Luo, M. Liu, H. Chen, Amelioration of experimental autoimmune encephalomyelitis by plumbagin through down-regulation of JAK-STAT and NFkB signaling pathways, PLoS One 6(2011)e27006.

166. J. Zhang, Y.Q. Zeng, J. Zhang, X.D. Pan, D.Y. Kang, T.W. Huang, X.C. Chen, Tripchlorolide ameliorates experimental autoimmune encephalomyelitis by down-regulating ERK1/2-NF-kappaB and JAK/STAT signaling pathways, J. Neurochem. 133(2015)104-112.

167 C.Y. Yin L, Z. Qu, L. Zhang, Q. Wang, Q. Zhang, L. Li, Nvolvement of JAK/STAT signaling in the effect of cornel iridoid glycoside on experimental autoimmune encephalomyelitis amelioration in rats, J. Neuroimmunol. 274(2014) 28-37.

168 J.J. Jia Y, Y. Bai, Z. Li, L. Liu, J. Luo, M. Liu, H. Chen, Amelioration of experimental autoimmune encephalomyelitis by plumbagin through down-regulation of JAK-STAT and NF-kappaB signaling pathways, PLoS One 6(2011)e27006.

169 L.L. Ren Y, T.B. Guo, J. Qiu, Y. Yang, A. Liu, J.Z. Zhang, Novel immunomodulatory properties of berbamine through selective down-regulation of STAT4 and action of IFN- γ in experimental autoimmune encephalomyelitis, J. Immunol. 181(2008)1491-1498.

170 K. Swiatek-Machado, J. Mieczkowski, A. Ellert-Miklaszewska, P. Swierk, I. Fokt, S. Szymanski, S. Skora, W. Szeja, G. Grynkiewicz, B. Lesyng, W. Priebe, B. Kaminska, Novel small molecular inhibitors disrupt the JAK/STAT3 and FAK signaling pathways and exhibit a potent antitumor activity in glioma cells, Cancer Biol. Ther. 13(2012)657-670.

171 C. Guo, L. Yang, C.X. Wan, Y.Z. Xia, C. Zhang, M.H. Chen, Z.D. Wang, Z.R. Li, X.M. Li, Y.D. Geng, L.Y. Kong, Anti-neuroinflammatory effect of sophoraflavanone g from sophora alopecuroides in LPS-activated BV2 microglia by MAPK, JAK/STAT and Nrf2/HO-1 signaling pathways, Phytomedicine 23(2016)1629-1637.

172 T. Zhou, Y. Yang, H. Zhang, Y. Che, W. Wang, H. Lv, J. Li, Y. Wang, S. Hou, Serenoa repens induces growth arrest, apoptosis and inactivation of STAT3 signaling in human glioma cells, Technol Cancer Res. Treat. 14(2015)729-736.

173 Y. Che, S. Hou, Z. Kang, Q. Lin, Serenoa repens induces growth arrest and apoptosis of human multiple myeloma cells via inactivation of STAT 3 signaling, Oncol. Rep. 22(2009)377-383.

174 Y. Liu, S.A. Gibson, E.N. Benveniste, H. Qin, Opportunities for translation from the bench: Therapeutic intervention of the JAK/STAT pathway in neuroinflammatory diseases, Crit. Rev. Immunol. 35(2015)505-527.

175 S.A. Schulze-Topphoff U, M. Varrin-Doyer, N. Molnarfi, S.A. Sagan, R.A. Sobel, P.A. Nelson, S.S. Zamvil, Laquinimod, a quinoline-3-carboxamide, induces type ii myeloid cells that modulate central nervous system autoimmunity, PLoS One 7(2012)e33797.

K. Ghoreschi, J. Bruck, C. Kellerer, C. Deng, H. Peng, O. Rothfuss, R.Z. Hussain, A.R.
Gocke, A. Respa, I. Glocova, N. Valtcheva, E. Alexander, S. Feil, R. Feil, K. Schulze-Osthoff,
R.A. Rupec, A.E. Lovett-Racke, R. Dringen, M.K. Racke, M. Rocken, Fumarates improve
psoriasis and multiple sclerosis by inducing type ii dendritic cells, J. Exp. Med. 208(2011)22912303.

177 R. Kacimi, R.G. Giffard, M.A. Yenari, Endotoxin-activated microglia injure brain derived endothelial cells via NF-kappaB, JAK-STAT and JNK stress kinase pathways, J. Inflamm. (Lond) 8(2011)7.

C.S. Nicolas, M. Amici, Z.A. Bortolotto, A. Doherty, Z. Csaba, A. Fafouri, P. Dournaud,P. Gressens, G.L. Collingridge, S. Peineau, The role of JAK-STAT signaling within the CNS.JAKSTAT 2(2013)e22925.

F. Yang, C. Brown, R. Buettner, M. Hedvat, R. Starr, A. Scuto, A. Schroeder, M. Jensen,R. Jove, Sorafenib induces growth arrest and apoptosis of human glioblastoma cells via dephosphorylation of STAT3, Mol. Cancer Ther. 9(2010)953-962.

A. Iwamaru, S. Szymanski, E. Iwado, H. Aoki, T. Yokoyama, I. Fokt, K. Hess, C. Conrad,
T. Madden, R. Sawaya, S. Kondo, W. Priebe, Y. Kondo, A novel inhibitor of the STAT3 pathway induces apoptosis in malignant glioma cells both in vitro and in vivo, Oncogene 26(2007)2435-2444.

181 O.D. Stechishin, H.A. Luchman, Y. Ruan, M.D. Blough, S.A. Nguyen, J.J. Kelly, J.G. Cairncross, S. Weiss, On-target JAK2/STAT3 inhibition slows disease progression in orthotopic xenografts of human glioblastoma brain tumor stem cells, Neuro Oncol. 15(2013)198-207.

182 Y. Liu, A.T. Holdbrooks, P. De Sarno, A.L. Rowse, L.L. Yanagisawa, B.C. McFarland, L.E. Harrington, C. Raman, S. Sabbaj, E.N. Benveniste, H. Qin, Therapeutic efficacy of suppressing the JAK/STAT pathway in multiple models of experimental autoimmune encephalomyelitis, J. Immunol. 192(2014)59-72.

183 H. Qin, J.A. Buckley, X. Li, Y. Liu, T.H. Fox, 3rd, G.P. Meares, H. Yu, Z. Yan, A.S. Harms, Y. Li, D.G. Standaert, E.N. Benveniste, Inhibition of the JAK/STAT pathway protects against alpha-synuclein-induced neuroinflammation and dopaminergic neurodegeneration, J. Neurosci. 36(2016)5144-5159.

B.C. McFarland, J.Y. Ma, C.P. Langford, G.Y. Gillespie, H. Yu, Y. Zheng, S.E. Nozell,D. Huszar, E.N. Benveniste, Therapeutic potential of AZD1480 for the treatment of human glioblastoma, Mol. Cancer Ther. 20(2011)2384-2393.

J.D. Goldstein, A. Burlion, B. Zaragoza, K. Sendeyo, J.K. Polansky, J. Huehn, E. Piaggio,
B.L. Salomon, G. Marodon, Inhibition of the jak/stat signaling pathway in regulatory T cells
reveals a very dynamic regulation of FOXP3 expression, PLoS One 11(2016)e0153682.

186 S.A. Nguyen, Novel drug discovery for glioblastoma multiforme using brain tumour initiating cells. Department of Neuroscience: University of Calgary, 2013.

187 K.V. Jensen, O. Cseh, A. Aman, S. Weiss, H.A. Luchman, The JAK2/STAT3 inhibitor pacritinib effectively inhibits patient-derived gbm brain tumor initiating cells in vitro and when used in combination with temozolomide increases survival in an orthotopic xenograft model, PLoS One 12(2017)e0189670.

188 C.R. Yu, Y.S. Lee, R.M. Mahdi, N. Surendran, C.E. Egwuagu, Therapeutic targeting of STAT3 (signal transducers and activators of transcription 3) pathway inhibits experimental autoimmune uveitis, PLoS One 7(2012)e29742.

189 V.A. Chou DHC, A. Choudhary, S.S. Scully, M. Schenone, A. Tang, R. Gomez, S.M. Burn, M. Lundh, T. Vital, E. Comer, P.W. Faloon, V. Dančík, C. Ciarlo, J. Paulk, M. Dai, C. Reddy, H. Sun, M. Young, N. Donato, J. Jaffe, P.A. Clemons, M. Palmer, S.A. Carr, S.L. Schreiber, B.K. Wagner, Kinase-independent small-molecule inhibition of JAK-STAT signaling, J. Am. Chem. Soc. 137(2015)7929-7934.

190 S.Y. Park JH, J.H. Jang, C.H. Jeong, S. Lee, B. Park, Asiatic acid attenuates methamphetamine induced neuroinflammation and neurotoxicity through blocking of nfkb/stat3/erk and mitochondria-mediated apoptosis pathway, J. Neuroinflamm. 14(2017) 240.

191 X. Yang, G. He, Y. Hao, C. Chen, M. Li, Y. Wang, G. Zhang, Z. Yu, The role of the JAK2-STAT3 pathway in pro-inflammatory responses of EMF-stimulated N9 microglial cells, J. Neuroinflamm. 7(2010)54.

192 S. Banerjee, A. Biehl, M. Gadina, S. Hasni, D.M. Schwartz, JAK-STAT signaling as a target for inflammatory and autoimmune diseases: Current and future prospects, Drugs 77(2017)521-546.

193 P. Maher, B. Conti, Deciphering the pathways that protect from IL-13-mediated potentiation of oxidative stress-induced dopaminergic nerve cell death, Cytokine 103(2018)114-120.

T.K. Sengupta, E.S. Talbot, P.A. Scherle, L.B. Ivashkiv, Rapid inhibition of interleukin-6
 signaling and STAT3 activation mediated by mitogen-activated protein kinases, Proc. Natl. Acad.
 Sci. USA 95(1998)11107-11112.

195 G.B. Nguyen VA, Cross-talk between alpha(1b)-adrenergic receptor (alpha(1b)ar) and interleukin-6 (IL-6) signaling pathways: Activation of alpha(1b)ar inhibits IL-6-activated STAT3 in hepatic cells by a p42/44 mitogen-activated protein kinase-dependent mechanism, J. Biol. Chem. 274(1999)35492-35498.

196 E.N. Benveniste, Y. Liu, B.C. McFarland, H. Qin, Involvement of the janus kinase/signal transducer and activator of transcription signaling pathway in multiple sclerosis and the animal model of experimental autoimmune encephalomyelitis, J. Interferon Cytokine Res. 34(2014)577-588.

197 J.J. O'Shea, M. Pesu, D.C. Borie, P.S. Changelian, A new modality for immunosuppression: Targeting the JAK/STAT pathway, Nat. Rev. Drug Discov. 3(2004)555-564.

198 V.N. Trieu, R. Liu, X.P. Liu, F.M. Uckun, A specific inhibitor of janus kinase-3 increases survival in a transgenic mouse model of amyotrophic lateral sclerosis, Biochem. Biophys. Res. Commun. 267(2000)22-25.

199 H.C. Lee, K.L. Tan, P.S. Cheah, K.H. Ling, Potential role of JAK-STAT signaling pathway in the neurogenic-to-gliogenic shift in down syndrome brain, Neural. Plast 2016(2016)7434191.

200 A. Pardanani, J.R. Gotlib, C. Jamieson, J.E. Cortes, M. Talpaz, R.M. Stone, M.H. Silverman, D.G. Gilliland, J. Shorr, A. Tefferi, Safety and efficacy of TG101348, a selective JAK2 inhibitor, in myelofibrosis, J. Clin. Oncol. 29(2011)789-796.

201 Y. Nakaya, K. Shide, T. Niwa, J. Homan, S. Sugahara, T. Horio, K. Kuramoto, T. Kotera, H. Shibayama, K. Hori, H. Naito, K. Shimoda, Efficacy of NS-018, a potent and selective JAK2/SRC inhibitor, in primary cells and mouse models of myeloproliferative neoplasms, Blood Cancer J. 1(2011)e29.

202 N.P. Shah PO, L. Sokol, S. Verstovsek, R. Hoffman, A.F. List, J. Cortes, H.M. Kantarjian, D.G. Gilliland, D.O. Clary, L.A. Bui, M. Wadleigh. A phase i study of XL019, a selective JAK2 inhibitor, in patients with primary myelofibrosis, post-polycythemia vera, or post-essential thrombocythemia myelofibrosis, Blood Cancer J. 112(2008)98.

203 F.P. Santos, H.M. Kantarjian, N. Jain, T. Manshouri, D.A. Thomas, G. Garcia-Manero, D. Kennedy, Z. Estrov, J. Cortes, S. Verstovsek, Phase 2 study of CEP-701, an orally available JAK2 inhibitor, in patients with primary or post-polycythemia vera/essential thrombocythemia myelofibrosis, Blood 115(2010)1131-1136.

A. Pardanani JG, V. Gupta, A.W. Roberts, M. Wadleigh, S. Sirhan, M.R. Litzow, W.J. Hogan, K. Begna, G. Smith, L.M. Bavisotto, M. Kowalski, A. Tefferi. An expanded multicenter phase I/II study of CYT387, a JAK- 1/2 inhibitor for the treatment of myelofibrosis, Blood 118(2011)3849.

205 S. Verstovsek RAM, S.K. Rhoades, J.L. Giles, C. Pitou, E. Jones, R.A. Walgren, J.T. Prchal, Phase I study of the JAK2 V617F inhibitor, LY2784544, in patients with myelofibrosis (MF), polycythemia vera (PV), and essential thrombocythemia (ET). Blood 118(2011)2814.

A.V. Purandare, T.M. McDevitt, H. Wan, D. You, B. Penhallow, X. Han, R. Vuppugalla,
Y. Zhang, S.U. Ruepp, G.L. Trainor, L. Lombardo, D. Pedicord, M.M. Gottardis, P. Ross-Macdonald, H. de Silva, J. Hosbach, S.L. Emanuel, Y. Blat, E. Fitzpatrick, T.L. Taylor, K.W.
McIntyre, E. Michaud, C. Mulligan, F.Y. Lee, A. Woolfson, T.L. Lasho, A. Pardanani, A.
Tefferi, M.V. Lorenzi, Characterization of BMS-911543, a functionally selective small-molecule
inhibitor of JAK2, Leukemia 26(2012)280-288.

207 V.J. Minturn JE, G.A. Yanik, J.R. Park, S.G. Groshen, E.T. Hellriegel, D. Bensen-Kennedy, K.K. Matthay, Phase I trial of lestaurtinib for children with refractory neuroblastoma: A new approach to neuroblastoma therapy consortium study, J. Clin. Oncol. 28(2010)9532-9532

Figures and figure legends

Figure 1. Schematic representation of JAK/STAT signaling pathway JAK, Janus kinase; STAT, signal transducer and activator of transcription; P, phosphorylated molecule.



Figure 2. Commonly conserved structure domains of the STATs family protein: N-terminal domain (ND); coiled-coil domain (CCD); DNA-binding domain (DBD); linker domain (LD); Src homology 2 domain (SH2); Tyrosine residue (Tyr) crucial for phosphorilation (P); transcriptional activation domain (TAD).



Figure 3. Chemical structure of curcumin (a), quercetin (b), plumbagin (c), luteolin (d) and resveratrol (e)

53

