Evaluation of a standard protocol for drying off and drying off therapy in dairy 1 cows based on the comparison of two different commercial antimicrobials 2 Turini L¹, Fratini F^{1,2}, Conte G³, Turchi B¹, Cerri D^{1,2}, Bertelloni F¹, Bonelli F¹ 3 ¹Dipartimento di Scienze Veterinarie, University of Pisa, Italy. ²Interdepartmental 4 Research Center "Nutraceuticals and Food for Health", Via del Borghetto 80, University 5 of Pisa (Italy). ³Dipartimento di Scienze Agrarie, Alimentari e Agro-ambientali, 6 7 University of Pisa, Italy. 8 Running title: 9 10 Drying off therapy in a dairy farm Turini et al. 11 12 13 14 15 Corresponding author: 16 Dr. Luca Turini 17 Dipartimento di Scienze Veterinarie, 18 Viale delle Piagge 2, 56122, Pisa, Italy 19 Phone number: +390502210115, Fax: +390502210654 20 Email: <u>luca.turini@phd.unipi.it</u> 21

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ORCID: 0000-0002-4164-8263

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This study aims to evaluate two commercial antibiotics for dry-off. Ninety-five Friesian cows and 380 quarters were included. Cows were classified as Control and Subclinical Mastitis based on somatic cell count. The California Mastitis Test and the teat end score have been performed. Quarters were randomly treated with Cloxacillin and Ampicillin (TA) and Cephalexin (TB). The effect of the therapy (TA vs TB) was estimated by X2 analysis based on the Wald test. The preservative and therapeutic action of TA vs TB were evaluated by the Kruskall-Wallis test. TB showed a statistically significant therapeutic effect in the control group, that might be related to the pharmacological activity of the two antibiotics. Also, the subclinical mastitis group most commonly presented more quarters affected compared to the control group, leading to a worse improvement despite proper therapy. In conclusion, an abruptly dried off, the California mastitis test, teat end score and somatic cell count evaluation, as long as microbial herd data might represent key concepts for an efficient drying off standard protocol in a dairy farm. In line with the herd bacterial population, both TA and TB might be employed for drying off therapy.

Key words: Veterinary, dairy cow, mastitis, dry period, drying-off therapy.

Introduction

- The dry period is defined as the nonlactating period prior to parturition in dairy cows.
- 44 Usually, this period begins at the end of the lactation curve, whom shape, and length is
- 45 strongly influenced by several environmental and genetic factors (Macciotta et al.
- 46 2011). The length of the dry period is about 60 days in Europe and United States of

America (Capuco et al. 1997; Annen et al. 2004). The dry period has a critical role for 47 udder health, due to an increased risk of intra-mammary infections (IMI) during this 48 time (Bradley and Green 2000; Whist and Østeras 2007). Despite a good dry period 49 50 management, some animals appear more prone to new IMI than others and may show 51 clinical mastitis in the next lactation (Hogan and Smith 2003). The most frequently isolated microorganisms at dry-off are *Streptococcus* spp., coagulase-negative 52 53 staphylococci (CNS), Staphylococcus aureus and Corynebacterium spp. (Pantoja et al. 54 2009). Dry cow therapy (DCT) is an intra-mammary treatment with an antibiotic, 55 administered at the beginning of the dry period. Since many years, the treatment of IMI at drying off has been a basis for mastitis control and management (Bradley and Green 56 2000). Dry cow therapy eliminates existing IMI and preventing new IMIs (Bradley and 57 58 Green 2000; Dingwell et al. 2004; Whist and Østeras 2007). The elimination of 59 infection and the prevention of new IMI in the dry period is easier than during lactation. The drug is not milked out, so the antibiotic can remain longer in the udder. Moreover, 60 the absence of regular milking reduces the exposition to pathogens by teat penetration 61 62 (Berry et al. 2004; Kashif et al. 2016). 63 The most common drugs used as dry-off therapy are intra-mammary tubes containing antibiotics such as penicillin, cloxacillin, cephalosporin and spiramycin (Kashif et al. 64 65 2016). Despite the large amount of literature about dry-off therapy strategies, usually 66 the decision concerning which antibiotic therapy would be better in a specific herd is 67 still made by drug popularity and farmer's preference and not based on scientific 68 evidences.

The aims of the present study were to evaluate the effects of two different commercial antibiotics at dry-off in the same herd on the mammary microbiological populations, in order to set a standard protocol for the dry off therapy in line with isolated bacteria.

Materials and methods

73 Animals

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The present study was carried out in an intensive dairy farm located in the North of 74 75 Italy. A total of 108 Italian Holstein Friesian cows in the same management conditions 76 were considered for the trial. An owner's written consent for participating on the study 77 have been recorded. All the experimental procedure were in compliance with the 2010/63/UE directive about the protection of animals in the scientific experiments. 78 79 Cows were between 210 and 220 days of gestation and were considered healthy based 80 on the complete physical examination. Animals included in the trial were not subjected to antibiotic and/or nonsteroidal anti-inflammatory drugs (NSAIDs) treatments for 30 81 days period before the admission time and did not showed udder or milk abnormalities 82 at dry-off. All the included animals were observed from drying-off until 15 days of 83 84 lactation. The drying off standard protocol proposed in this study has been carried out as 85 follow. All eligible cows were dried off abruptly. During the last milking session, the teat end score (TESa) was recorded for each quarter as reported in literature (Hamann 86 87 and Mein 1996; Neijenhuis et al. 2000), along with the udder evaluation. The California 88 Mastitis Test (CMTa) was used as the screening test for clinical or subclinical mastitis 89 detection (Barnum and Newbould 1961). A sample has been considered positive for 90 CMT ≥ 1 (Blowey and Edmondson 2010). Before the antibiotic administration, a sterile 91 milk sample for bacteriologic analysis was collected from positive CMT quarter in a 15 mL tubes (Falcon®, BD, Italy) and was stored at 4° C $\pm 2^{\circ}$ C until the time of analysis. 92

93 All the samples were processed within 3 hours. After these evaluations, the animals were treated with Cloxacillin and Ampicillin (Cloxalene Max, Fatro, Italy), or 94 Benzathine Cephalexin (Rilexine, 500 HL, Virbac, Italy) as drying-off therapy, despite 95 96 the CMT results. Dry-off cows were then transferred to a separate box, located in a 97 different barn. The dry-cow's box was composed by a pen for the daily exercise, in between the feeding area, where all animals were fed at the same time. Moreover, in this 98 99 area there was a resting zone composed by an appropriate number of cubicles bedded 100 with daily-changed straw. At dry-off, cows were fed a low energy density diet, 101 administered as total mixed ration (TMR) two times per day (Table 1). At the inclusion time, animals were divided in two groups (Treatment A and Treatment B) based on the 102 103 drugs supply at the farms. Treatment A (TA) quarters received 600 mg of Cloxacillin 104 and 300 mg of Ampicillin (Cloxalene Max, Fatro, Italy), while treatment B (TB) 105 quarters received 375 mg of Benzathine Cephalexin (Rilexine, 500 HL, Virbac, Italy). The whole 4 quarters of a single cow were treated with the same antibiotic. For a better 106 classification of the cows, the average somatic cells count (SCCa) of the 4 quarters has 107 108 been evaluated at the inclusion time by the MilkoScan (FOSS, Italy). Thus, animals 109 were retrospectively classified as "C cows" when the SCC was < 150'000 cells/mL for primiparous and < 250'000 cells/mL for multiparous cows and as "SCM cows" when 110 111 the SCC was > 150'000 cells/mL for primiparous and > 250'000 cells/mL for 112 multiparous (de Haas et al. 2008; Windig et al. 2010). The "C cows" has been 113 considered as control animals. A second evaluation took place 15 days after parturition 114 for each quarter included. The same parameters collected at the inclusion time were 115 considered: 1) teat end score for each quarter (TESb); 2) CMT performed for each

116 quarter 15 days after parturition (CMTb); 3) the average SCC (SCCb) of each single 117 quarter. Bacteriology analysis 118 119 To isolate the mastitis agents and to screening the microbials population of the studied herd, 0.01 mL of each CMTa positive milk sample was streaked on blood agar plates 120 containing 10% sheep blood. Each plate was incubated aerobically at 37 °C for 24-48 h. 121 After observation of colony morphology and hemolytic patterns on blood agar, isolates 122 123 were submitted to Gram staining, catalase and oxidase testing and additional 124 biochemical and metabolic evaluations as needed. Gram-negative organisms were successively identified by sowing on appropriate 125 126 selective and differential media; furthermore, enzyme activities, acid production from 127 different carbohydrates, assimilation of various substrates were determined using 128 commercial systems- API ZYM, API 20E and API 20NE (BioMerieux ®) according to the manufacturer's instructions. 129 Staphylococcus spp., grown on Baird Parker medium with the typical halo associated 130 with lecithinase positivity and characterized by typical zones of complete and 131 132 incomplete hemolysis and nonhemolytic Staphylococcus spp. that had a positive tube test for free coagulase were classified as Staphylococcus aureus; all other staphylococci 133 134 were classified as CNS. Moreover, all isolates belonging to Staphylococcus genus were 135 also identified using API Staph (BioMerieux ®). The other Gram-positive cocci, grown 136 on blood agar plates and negative for catalase test, were phenotypically identified by 137 means of API 20 Strep (BioMerieux ®). 138 Statistical analysis

Data concerning SCCa and SCCb, CMTa and CMTb, type of treatment (TA vs TB) and 139 TESa and TESb of C and SCM groups were expressed as prevalence. 140 Data obtained in this work were analyzed by R software (R Development Core Team). 141 142 Firstly, the effect of the therapy (use of the antibiotics TA or TB) on the number of positive or negative teats to CMTa were estimated by $\chi 2$ analysis based on the Wald 143 test. The analysis was repeated both for cows classified as "C" or "SCM". 144 145 Subsequently, we estimated the different effect of the two antibiotics by the non-146 parametric Kruskall-Wallis test. This analysis was repeated in two different situations: 147 firstly, we evaluated the protective action of the two antibiotics considering the percentage of negative CMT teats that remain unchanged after the treatment; while the 148 149 therapeutic effect was estimated considering the percentage of positive CMT teats that 150 became negative after treatment. 151 **Results** A total of 380 quarter in 95 cows were included, from 46 primiparous cows (48%) and 152 153 49 multiparous cows (52%). Thirteen animals from the 108 enrolled were excluded 154 from the study for the presence of only three functional udder quarters or because of 155 treatments during the 30 days before the inclusion time. Cows were in first to fifth 156 lactation, with an average of 4.5 (3-7) years old, average body weight of 660 kg (520-157 710) and average body condition score (BCS) of 3.25 (2.5-3.75). One hundred and thirty-six/380 quarters (referred to 34 cows) received TA while 158 159 244/380 quarters (referred to 61 cows) were treated with TB. Based on SCCa evaluation, a total of 29 cows (18 primiparous and 11 multiparous) were included in the 160 C group, while 66 cows (28 primiparous and 38 multiparous) were included in the SCM 161 162 group. A total of 32 quarters belong to C group were treated with TA, while 84 quarters

were treated with TB. A total of 104 quarters belong to SCM group were treated with 163 164 TA, while 160 quarters were treated with TB. Data concerning SCCa, CMTa and TESa of C and SCM groups were reported in Table 2. 165 166 A total of 68 out of 380 milk samples were CMTa positive, thus they were sampled for 167 bacteriologic analysis. Fiftythree out of 68 milk samples (78%) were culture negative, 9 (13%) were positive for environmental pathogens (CNS and Serratia), while the 168 remaining 6 samples (9%) were positive for contagious pathogens. The pH was out of 169 the normal range (6.5-6.7) in 41 out of 68 (60%) milk samples (Ruegg e Erskine, 2015). 170 171 The average pH value was 6.8±0.2. Results concerning CMTb and TESb of C and SCM cows, grouped based on SCCa are 172 reported in Table 3, while results concerning CMTb and TESb of C and SCM cows, 173 174 grouped based on SCCb are reported in Table 4. 175 Results concerning changing between CMTa vs CMTb in TA vs TB cows, classified based on SCCa are reported in Table 5. 176 Table 6 showed the result of contingency analysis related to the number of quarters 177 treated with TA and TB. Statistical differences between expected and observed number 178 179 of quarters were revealed for SCM cows (p < 0.001) with higher number of positive quarters observed for TA. 180 181 Results concerning the effect of the different antibiotic on teat in a preventive 182 (expressed as % of teats that continued to show CMT negative after treatment) and 183 therapy (expressed as % of teats that showed CMT negative after treatment) are reported in Table 7. Different effect between the two treatments was only observed only 184 185 for the C cows (p = 0.022) during the rapeutic action, while no difference was revealed

for SCM cows (p = 0.990). On the contrary, the preventive effect was similar between TA and TB.

Discussion

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Dry period is crucial for two different reasons: 1) high rates of IMIs healing during this period and 2) the develop rates of new IMIs during this period are higher than during the lactating period (Dingwell et al. 2003; Halasa et al. 2009). For these reasons, intramammary antibiotics are used to treat any existing IMI at dry-off and to prevent new IMI during the dry period. However, the choice of the best antibiotics therapy would be set according to the kind of mammary pathogens. Prudent use of antibiotics is recommended to prevent the development of antimicrobial resistance. The isolation of different pathogens and their sensitivity or resistance to some antibiotics is the key to choose the most suitable antibiotics to avoid the antimicrobial resistance (Scherpenzeel et al. 2014). The whole population of this study presented no udder or milk abnormalities at the dry-off but only CMT alterations, with a prevalence for subclinical mastitis of 18%, which is slightly lower value compared with data reported in literature (Busato et al. 2000; Gianneechini et al. 2002). However, our study only evaluated cows at drying-off, while data presented in literature usually came from the screening of the whole herd. This might explain the differences in results. Reported prevalence of infection at dry-off, due to any pathogen, ranges from 28 % to 50 % at cow level (Rindsig et al. 1978; Browning et al. 1994). Our results are slightly lower compared with other authors. However, the incidence of infections at dry-off in a herd could be influenced by many factors and this might explain the differences found in prevalence (Torres et al. 2008).

Otherwise, the high prevalence of subclinical mastitis obtained in this study, confirmed that subclinical mastitis still represents an important problem in the dairy cow industry (Pisoni 2007). Thus, screening for subclinical mastitis at the time of dry-off is mandatory for keeping a high standard of udder and milk health and hygiene. The screening method choose in the present study was the CMT because it still represents the commonest used one in the field (Ruegg e Erskine 2015). Also, literature recommended the use of CMT for identification of IMI when herd prevalence of IMI is lower than 15% (Torres et al. 2008). Authors knew the average prevalence of IMI from the history of the herd. However, samples CMT positive were tested for BE and the prevalence of positive BE was 22%. Despite CMT is largely used in dairy practice, our results confirmed the too high sensibility of the test (Bradley et al. 2012; Zecconi and Zanirato 2013; Sgorbini et al. 2014). The CMT may be influenced by several factors, i.e. the time of sampling (morning vs evening) or the season (summer vs fall) (Bradley et al. 2012; Zecconi and Zanirato 2013), the storage and processing of the sample (Viguier et al. 2009; Bradley et al. 2012), udder inflammation other than infectious problems (i.e. trauma, alimentary management, milking procedures) (Zecconi and Zanirato 2013). Thus, for a more proper use of antibiotic therapy at dry-off, the BE might be considered the gold standard test for drive the decision (Ruegg and Erskine 2015). Staphylococcus spp. was the most common bacterium isolated in our population, as confirmed by literature (Torres et al. 2008; Zecconi and Zanirato 2013; Ruegg and Erskine 2015). Compared with TA, TB showed a statistically significant therapeutic effect in "C cows" while no difference where observed for "SCM cows". Cloxacillin (TA) represents one of the most widely used antibiotics for the drying off in dairy cows (Halasa et al. 2009; Bhutto et al. 2011). However, Cephalexin (TB) has been

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successfully used in the treatment of subclinical mastitis of lactating cows (Tiwari et al. 2000). The difference in therapeutic effect found in the present study might be related to the pharmacological activity of the two antibiotics. Also, "SMC cows" most commonly had affected more quarters than "C cows", leading to a worse improvement despite a proper therapy. Further studies with an increasing number of animals included might complete the present findings.

Conclusions

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In conclusion, an abruptly dried off, the CMT, TEC and SCC evaluation, as well as microbiological examination might represent key concepts for an efficient drying off standard protocol in a dairy farm. The SCC evaluation is essential to find subclinical mastitis and to decide to treat the single quarter of the udder. The pathogens isolations before the drying off is the key to choose the ideal antimicrobial to treat it. In line with the herd bacterial population, both TA and TB might be employed for drying off therapy.

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References

- 1. Annen, E. L., R. J. Collier, M. A. McGuire, J. L. Vicini, J. M. Ballam, M. J.
- Lormore (2004): Effect of modified dry period lengths and bovine somatotropin
- on yield and composition of milk from dairy cows. J. Dairy Sci. 87, 3746–3761.
- 253 https://doi.org/10.3168/jds.S0022-0302(04)73513-4
- 254 2. Barnum, D. A., F. H. S. Newbould (1961): The use of the California Mastitis Test
- for the detection of bovine mastitis. Can. Vet. J. 2, 83–90.

- 3. Berry, E. A., H. Hogeveen, J. E. Hillerton (2004): Decision tree analysis to
- evaluate dry cow strategies under UK conditions. J. Dairy Res. 71, 409–418.
- 258 https://doi.org/10.1017/S0022029904000433
- 4. Bhutto, A. L., R. D. Murray, Z. Woldehiwet (2011): The effect of dry cow therapy
- and internal teat-sealant on intra-mammary infections during subsequent
- lactation. Res. Vet. Sci. 90, 316–320.
- 262 https://doi.org/10.1016/j.rvsc.2010.06.006
- 5. Blowey, R., P. Edmondson (2010): Somatic cell count. In: Mastitis control in
- Dairy Herds (Blowey, R., P. Edmondson P, Eds), 2nd Edition, CABI, UK, pp. 153-
- 265 170.
- 6. Bradley, A. J., M. J. Green (2000): A study of the incidence and significance of
- intramammary enterobacterial infections acquired during the dry period. J. Dairy
- 268 Sci. 83, 1957-1965.
- 269 https://doi.org/10.3168/jds.S0022-0302(00)75072-7
- 7. Bradley, A., H. Barkema, A. Biggs, M. Green, T. Lam (2012): Control of mastitis
- and enhancement of milk quality. In: Dairy Herd Health (Green, D., Eds), CABI,
- 272 UK, pp. 117-168.
- 8. Browning, J. W., G. A. Mein, P. Brightling, T. J. Nicholls, M. Barton (1994):
- Strategies for mastitis control: dry cow therapy and culling. Aust. Vet. J. 71, 179–
- 275 18.
- 276 https://doi.org/10.1111/j.1751-0813.1994.tb03383.x
- 9. Busato, A., P. Trachsel, M. Schallibaum, J. M. Blum (2000): Udder health and
- 278 risk factors for subclinical mastitis in organic dairy farms in Switzerland. Prev.
- 279 Vet. Med. 44, 205–220.

- 280 https://doi.org/10.1016/S0167-5877(00)00104-5
- 10. Capuco, A. V., R. M. Akers, J. J. Smith (1997): Mammary growth in Holstein
- cows during the dry period: quantification of nucleic acids and histology. J. Dairy
- 283 Sci. 80, 477–487.
- 284 https://doi.org/10.3168/jds.S0022-0302(97)75960-5
- 11. de Haas, Y., W. J. Ouweltjes ten Napel, J. J. Windig, G. de Jong (2008):
- Alternative somatic cell count traits as mastitis indicators for genetic selection. J.
- 287 Dairy Sci. 91, 2501–2511.
- 288 https://doi.org/10.3168/jds.2007-0459
- 12. Dingwell, R.T., D. F. Kelton, K. E. Leslie (2003): Management of the dry cow in
- control of peripartum disease and mastitis. Vet. Clin. North Am. Food. Anim.
- 291 Pract. 19, 235–265.
- 292 10.1016/S0749-0720(02)00072-5
- 13. Dingwell, R.T., K. E. Leslie, Y. H. Schukken, J. M. Sargeant, L. L. Timms, T. F.
- Duffield, G. P. Keefe, D. F. Kelton, K. D. Lissemore, J. Conklin (2004):
- Association of cow and quarter-level factors at drying-off with new
- intramammary infections during the dry period. Prev. Vet. Med. 63, 75-89.
- 297 https://doi.org/10.1016/j.prevetmed.2004.01.012
- 14. Gianneechini, R., C. Concha, R. River, I. Delucci, J. Moreno Lopez (2002):
- Occurrence of clinical and subclinical mastitis in dairy herds in the west littoral
- region in Uruguay. Acta Vet. Scand. 43, 221–230.
- 301 https://doi.org/10.1186/1751-0147-43-221

- 302 15. Halasa, T., O. Osteras, H. Hogeveen, T. Van Werven, T. Nielen (2009) Meta-
- analysis of dry cow management for dairy cattle. Part1. Protection against new
- intramammary infections. J. Dairy Sci. 92, 3134-3149.
- 305 https://doi.org/10.3168/jds.2008-1740
- 16. Hamann, J., G. A. Mein (1996): Teat thickness changes may provide biological
- test for effective pulsation. J. Dairy Sci. 63, 179–189.
- 308 https://doi.org/10.1017/S002202990003168X
- 17. Hogan, J. S., K. L. Smith (2003): Coliform mastitis. Vet. Res. 34:507–519.
- 310 https://doi.org/10.1051/vetres:2003022
- 18. Macciotta, N. P. P., C. Dimauro, S. P. G. Rassu, G. Steri, G. Pulina (2011): The
- mathematical description of lactation curves in dairy cattle. Ital. J. Anim. Sci.
- 313 10:51.
- 314 https://doi.org/10.4081/ijas.2011.e51
- 19. Neijenhuis, F., H. W. Barkema, H. Hogeveen, J. P. Noordhuizen (2000):
- Classification and longitudinal examination of callused teat ends in dairy cows. J.
- 317 Dairy Sci. 83, 2795–2804.
- 318 https://doi.org/10.3168/jds.S0022-0302(00)75177-0
- 20. Pantoja, J. C. F., C. Hulland, P. L. Ruegg (2009): Dynamics of somatic cell counts
- and intramammary infections across the dry period. Prev. Vet. Med. 90, 43-54.
- 321 https://doi.org/10.1016/j.prevetmed.2009.03.012
- 322 21. Pisoni G (2007) Mastitis in a dairy herd (in Italian). (Point Veterinaire Italie, La
- fenice grafica, Eds), Borghetto Lodigiano (Lo), Italy.
- 22. Rindsig, R. B., R. G. Rodewald, A. R. Smith, S. L. Spahr (1978): Complete versus
- selective dry cow therapy for mastitis control. J. Dairy Sci. 61, 483–1497.

- 326 https://doi.org/10.3168/jds.S0022-0302(78)83753-9
- 23. Ruegg, P. L., R. J. Erskine (2015): Mammary Gland Health. In: Large Animal
- Internal Medicine (Smith, B. P., Eds),5th ed., Elsevier. St Louis, pp. 1015-1043.
- 329 24. Scherpenzeel, C. G. M., I. E. M. den Uijl, G. van Schaik, R. G. M. Olde Riekerink,
- J. M. Keurentjes, T. J. G. M. Lam (2014) Evaluation of the use of dry cow
- antibiotics in low somatic cell count cows. J. Dairy Sci. 97:3606-3614.
- https://doi.org/10.3168/jds.2013-7655
- 25. Sgorbini, M., F. Bonelli, F. Fratini, A. Sbrana, M. Brombin, V. Meucci, M.
- Corazza, V. Ebani, F. Bertelloni, B. Turchi, D. Gatta, D. Cerri (2014): Mastitis in
- dairy cattle: a comparison of some screening tests and bacteriology. Large Anim.
- 336 Rev. 20(1), 9-16.
- 26. Torres, A. H., P. J. Rajala-Schultz, F. J. DeGraves, K. H. Hoblet (2008): Using
- dairy herd improvement records and clinical mastitis history to identify
- subclinical mastitis infections at dry-off. J. Dairy Res. 75, 240–247.
- 340 https://doi.org/10.1017/S0022029908003257
- 27. Tiwari, A., R. S. Sisodia, K. S. Misraulia (2000): Treatment of subclinical mastitis
- in cows with Cephalexin. Indian J. Vet. Med. 20(2), 97-98.
- 28. Kashif, M., M. Rizwan, M. Ali, T. Ahmad, A. Z. Durrani (2016): Control of
- Mastitis through dry cow therapy: A review. Veterinaria 2(2), 13-16.
- 29. Whist, A. C., O. Østeras (2007): Associations between somatic cell counts at
- calving or prior to dry-off and clinical mastitis in the remaining or subsequent
- 347 lactation. J. Dairy Res. 74, 66-73.
- 348 https://doi.org/10.1017/S0022029906002172

349	30. Windig, J. J., W. Ouweltjes, J. Ten Napel, G. de Jong, R. F. Veerkamp, Y. de Haas
350	(2010): Combining somatic cell count traits for optimal selection against mastitis.
351	J. Dairy Sci. 93, 1690–1701.
352	https://doi.org/10.3168/jds.2009-2052
353	31. Viguier, C., S. Arora, N. Gilmartin, K. Welbeck, R. O'Kennedy (2009): Mastitis
354	detection: current trends and future perspectives. Trends Biotechnol. 27, 486-
355	493.
356	https://doi.org/10.1016/j.tibtech.2009.05.004
357	32. Zecconi, A., G. Zanirato (2013): Mastitis control for a sustainable dairy farm (in
358	Italian) (diVet e GeSan, Eds), pp. 69.
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Table 1. Dry matter concentrations and chemical composition of the close-up dry (CUD)

diets fed to Holstein Friesian cows.

CUD Diet								
Ingredients	Percentage							
Corn silage	45.46							
Alfalfa hay	5.00							
Corn grains	18.28							
Cottonseed hulls	11.17							
Soybean meal	8.38							
Cottonseed with lint	5.2							
Prolak	2.55							
Springer minerals	2.92							
Salt	0.5							
Sodium bicarbonate	0.54							
Chemical composition	Percentage (DM basis)							
DM (%)	62.07							
CP	14.88							
Sol CP ¹	29.25							
ADF	26.17							
NDF	40.53							
EE^2	3.81							
TDN	67.22							
NE_L (Mcal/Kg)	1.52							
\mathbf{K}^{+}	1.14							
Na^+	0.40							
Cl ⁻	0.82							
S ⁻²	0.33							
Ca^{+2}	1.07							
\mathbf{P}^{+}	0.38							
Calculated DCAD ³	+19.4							

¹Percentage of the CP

380 ²Ether extract

381 ³Meq/100g DM

Table 2. Data concerning the evaluation of SCCa, CMTa and TESa scores performed in C and SCM groups. Legend: SCCa – somatic cells count at inclusion time (dry-off); CMTa – california mastitis test at inclusion time (dry-off); TESa - teat score for each quarter; TA – treatment A (600 mg of Cloxacillin and 300 mg of Ampicillin); TB – treatment B (375mg of Benzathine Cephalexin); C group – control group; SCM group – subclinical mastitis group.

SCCa (cell/ml)	CMTa	TESa sco	ore 1	TESa so	core 2	TESa so	core 3	TESa so	core 4
		TA	TB	TA	TB	TA	TB	TA	TB
C group	Positive	1/32	0/84	2/32	4/84	0/32	2/84	0/32	0/84
	Negative	26/32	68/84	3/32	10/84	0/32	0/84	0/32	0/84
SCM group	Positive	11/104	4/160	17/104	14/160	2/104	8/160	0/104	3/160
	Negative	54/104	94/160	19/104	32/160	1/104	3/160	0/104	2/160

Table 3. Data concerning the evaluation of CMTb and TESb scores performed in C and SCM grouped based on SCCa. Legend: SCCa – somatic cells count at inclusion time (dry-off); CMTb – california mastitis test 15 days after parturition; TA – treatment A (600 mg of Cloxacillin and 300 mg of Ampicillin); TB – treatment B (375mg of Benzathine Cephalexin); C group – control group; SCM group – subclinical mastitis group.

SCCa (cell/ml)	CMTb	TESb sco	ore 1	TESb sc	core 2	TESb s	core 3	TESb s	core 4
		TA	TB	TA	TB	TA	TB	TA	TB
C group	Positive	3/32	1/84	1/32	3/84	0/32	0/84	0/32	0/84
	Negative	24/32	67/84	4/32	11/84	0/32	2/84	0/32	0/84
SCM group	Positive	6/104	3/160	5/104	8/160	1/104	3/160	0/104	1/160
	Negative	59/104	95/160	31/104	38/160	2/104	8/160	0/104	4/160

Table 4. Data concerning the evaluation of CMTb and TESb scores performed in C and SCM groupep based on SCCb. Legend: SCCb – somatic cells count 15 days after parturition; CMTb – california mastitis test 15 days after parturition; TA – treatment A (600 mg of Cloxacillin and 300 mg of Ampicillin); TB – treatment B (375mg of Benzathine Cephalexin); C group – control group; SCM group – subclinical mastitis group.

SCCb (cell/ml)	CMTb	TESb scor	re 1	TESb sc	ore 2	TESb sc	core 3	TESb sc	ore 4
		TA	TB	TA	TB	TA	TB	TA	TB
C group	Positive	1/136	1/244	4/136	0/244	0/136	0/244	0/136	0/244
_	Negative	72/136	120/244	23/136	38/244	2/136	7/244	0/136	2/244
SCM group	Positive	6/136	9/244	5/136	5/244	1/136	3/244	0/136	2/244
	Negative	13/136	36/244	9/136	17/244	0/136	3/244	0/136	1/244

Table 5. Results concerning changing in CMTa vs CMTb for C and SCM group and the information about treatments received (TA vs TB). Legend: SCCa – somatic cells count at inclusion time (dry-off); CMTa – california mastitis test at inclusion time (dry-off); CMTb – california mastitis test 15 days after parturition; TA – treatment A (600 mg of Cloxacillin and 300 mg of Ampicillin); TB – treatment B (375mg of Benzathine Cephalexin); C group – control group; SCM group – subclinical mastitis group.

	Treatment	CMTa-neg	CMTa-neg vs	CMTa-pos	CMTa-pos
	at dry-off	VS	CMTb-pos	VS	VS
		CMTb-neg	_	CMTb-neg	CMTb-pos
C group	TA	27	2	1	2
	TB	74	4	6	0
SCM	TA	69	5	23	7
group	TB	121	10	24	5

Table 6. Table of contingency for the estimation of antibiotic treatment effect. The data refer to the number of treated teats. A) C cows; B) SCM cows.

A) P=0.297	7						
	Observe	d			Expected		
-	TA	TB	Total		TA	TB	Total
Positive	3	6	9	Positive	4	5	9
Negative	29	78	107	Negative	28	79	107
Total	32	84	116	Total	32	84	116
B) P< 0.00	1						
	Observe	d			Expected		
	TA	TB	Total		TA	TB	Total
Positive	30	29	59	Positive	23	36	59
Negative	74	131	205	Negative	81	124	205
Total	104	160	264	Total	104	160	264

Table 7. Effect of the different antibiotic on teat in a preventive (expressed as % of teats that remain negative after treatment) and therapy (expressed as % of teats that become negative after treatment).

	TA	TB	SE	P-value
Preventive effect in "C cows"	92.85 %	94.91 %	6.08	0.760
Preventive effect in "SCM cows"	92.68 %	92.37 %	4.14	0.770
Therapeutic effect in "C cows"	50.00 %	100.00 %	14.56	0.022
Therapeutic effect in "SCM cows"	76.61 %	81.60 %	15.84	0.990