

# HELMINTHOLOGIA, 57, 1: 78 - 82, 2020

# **Research Note**

# Evaluation of a marketed polyherbal dewormer against intestinal strongyles in naturally infected donkeys

# R. A. PAPINI<sup>1,\*</sup>, C. ORSETTI<sup>2</sup>, M. SGORBINI<sup>2</sup>

Ospedale Didattico Veterinario "Mario Modenato", Dipartimento di Scienze Veterinarie, Via Livornese Lato Monte, 1 – 56122 San Piero a Grado (PI), Italy. E-mail: \*roberto.amerigo.papini@unipi.it, chiara.orsetti@gmail.com, micaela.sgorbini@unipi.it

# Article infoSummaryReceived July 15, 2019<br/>Accepted September 23, 2019The study evaluated the effectiveness of a commercially available polyherbal dewormer to control<br/>intestinal strongyles in naturally infected donkeys. The animals were allotted to two groups: treated<br/>with the herbal dewormer (n=8) according to manufacturer recommendations and untreated control<br/>group (n=6). Fecal samples were taken from each animal on days 0 (day of treatment), 14, 21 (day<br/>of second additional treatment), 35, and 42. Faecal egg count reduction tests showed very negligible<br/>or no reduction in number of strongyle eggs for donkeys in the phytotherapeutic treatment group<br/>compared to those in the control group. Thus, the herbal dewormer was mostly ineffective in reduc-<br/>ing fecal egg counts in donkeys infected with intestinal strongyles. These findings can make equine<br/>practitioners aware of possible disadvantages of herbal dewormer; phytotherapy

#### Introduction

Donkeys play a key role as working animals in developing countries, and they are also used for meat and milk productions, social activities, tourism and leisure (Camillo *et al.*, 2018). Consequently, the interest in the welfare and diseases of this species is constantly increasing (Bonelli *et al.*, 2016; Sgorbini *et al.*, 2017, 2018), including parasitic diseases and strategies for their control.

The high occurrence of large and small strongyles in donkeys has been widely reported (Matthee *et al.*, 2000; Getachew *et al.*, 2010; Matthews *et al.*, 2013; Ismail *et al.*, 2016; Jajere *et al.*, 2016; Dibaba *et al.*, 2017). Intestinal strongyles may negatively affect body conditions, live weight and haematological parameters of donkeys (Matthee *et al.*, 2002; Yoseph *et al.*, 2005) and their control is typically performed by the administration of anti-helminthic drugs licensed for use in horses (Matthee *et al.*, 2002). However, the development of drug-resistant populations of small strongyles

\* - corresponding author

have been reported in donkeys (Matthee et al., 2002; Lawson et al., 2015). This factor and the growing consumer trend to ask for food products free of chemical residues, including milk from donkeys raised according to organic farming methods (Camillo et al., 2018), prompt researchers, practising veterinarians, and organic donkey farmers to search for sustainable alternatives to control gastrointestinal parasites in this host species. Among the sustainable alternatives, there are various medicinal plants that are reported to have anthelmintic properties in donkeys (Scantlebury et al., 2013). However, to the best of the authors' knowledge, the effectiveness of herbal formulations marketed for parasite control in donkeys under field conditions has not been yet investigated so far. Hence, the aim of the present study was to determine the effectiveness of a commercially available polyherbal dewormer, marketed for use in equines, to reduce the faecal output of stronayle eggs in donkeys.

# **Materials and Methods**

#### Study animals, sample collection, and laboratory procedures

The study was carried out at the facilities of the veterinary teaching hospital "Mario Modenato" (University of Pisa, Italy) between January and February 2019. The study period was chosen for practical reasons. Indeed, female donkeys selected for the study were going to start the reproductive season in the following months (late winter/early spring). So, we wanted to avoid using the animals for the study purposes during the heat period, mating, and pregnancy. Fourteen asymptomatic Amiata donkeys (Equus asinus) of both sexes (10 unpregnant females and 4 intact males) were enrolled in the study irrespective of the parasite burden. They were aged 7-13 years and none of them had received any anthelmintic treatment for at least 12 months before the beginning of the study. The donkeys were naturally infected by intestinal strongyles. Infections were confirmed prior to the beginning of the study by the Mini-FLO-TAC technique in combination with Fill-FLOTAC. A commercial sodium nitrate solution with specific gravity of 1.200 (Coprosol®, Candioli Farmaceutici S.p.A., Beinasco (TO), Italy) was used as flotation solution. The Mini-FLOTAC technique in combination with Fill-FLOTAC has a reported analytic sensitivity of five eggs per gram (EPG) of faeces and it was performed according to the detailed instructions provided at the web site of the supplier (https:// www.parassitologia.unina.it/wp-content/uploads/2015/07/001-Erbivori-fresh Layout-1-1 cut.pdf). This tool was chosen for copromicroscopic diagnosis and EPG counts because of the excellent performance previously shown in various studies to assess gastrointestinal helminth infections and anthelmintic drug efficacy in veterinary medicine (Rinaldi et al., 2014; Bellaw et al., 2018).

#### Study design

Each of the 14 donkeys was allotted to one of two groups: treated group (n=8) and untreated control group (n=6). Because there is no indication in previous studies that the response to anthelmintic treatment is different between sexes in donkeys, animals were allotted as follows. Females were randomly assigned to either treated or control group and males were assigned to control group. Thus, the treated group contained 8 females whilst the control group contained 2 females and 4 males. After allotments, the two groups were kept in separate paddocks as follows. Eight unpregnant females in the treatment group were kept all together in the same paddock. Due to management reasons, the 6 donkeys in the control group were separated by sex into five paddocks, that is 2 unpregnant females were kept together in the same paddock whilst the 4 intact males were kept separately in individual paddocks. All the donkeys allotted to each group were fed with hay from mixed-grass meadows and water ad libitum during the entire study period. On day 0, faecal samples were individually collected from the rectal ampulla of each donkey into plastic containers. Samples were appropriately labelled and brought to the

laboratory where individual EPG counts were performed by using the Mini-FLOTAC technique in combination with Fill-FLOTAC, as above reported. The same day the bodyweight of each donkey in the treated group (n=8) was estimated by a weight tape, ranging from 280 to 393 kg. Thereafter, these animals were treated by oral administration of a marketed polyherbal dewormer at a dose rate of 8.3 gr per 100 kg bodyweight. The product is commercialized in the form of paste and is licensed for use in equines in Italy. It contains extracts of Mallotus philippinensis, Carduus marianus, Urtica urens, Genziana lutea, and Eucalyptus globulus. It is considered atoxic, can be used in pregnant and young animals, and has no withdrawal time. According to the dose suggested in the drug package, a 50 gr syringe is useful for treatment of horses of weight up to 600 kg. Thus, the dose used for treatment of donkeys in the present study was extrapolated from the dose recommended for horses by the manufacturer. During the oral administration of the herbal dewormer, all donkeys in the treated group showed that they enjoyed the taste very much. The remaining 6 animals, assigned to the control group, were kept untreated. Subsequently, faecal sample collections and EPG counts were individually performed as above reported in both groups on days 14, 21, 35, and 42. Furthermore, on day 21 all the donkeys in the treated group were administered again an additional administration of a 50 gr syringe of the paste used for the initial treatment, as adjunct treatment irrespective of bodyweight. After treatment, they were observed for possible adverse reactions for about 1 hour.

#### Data analysis

The faecal egg count reduction (FECR) test was used to determine the anthelmintic efficacy of the tested polyherbal dewormer (Kaplan & Nielsen, 2010). Arithmetic means (AMs) of the EPG counts were calculated to determine the mean percentage reductions on day 14 and on day 35, according to the following formula:

FECR % = 
$$\frac{\text{Pre-treatment EPG - Post-treatment EPG}}{\text{Pre-treatment EPG}} \times 100$$

Egg shedding reduction can vary based on the drug tested and the number of horses investigated (Kaplan and Nielsen, 2010) but there are currently no available indications for phytotherapeutic products in literature. Thus results were interpreted following the classification of Kaplan and Nielsen (2010) and we arbitrarily assumed that the cut-off value for appropriate efficacy of the tested polyherbal dewormer against intestinal strongyles in donkeys was FECR at least  $\geq$ 90 %.

AMs  $\pm$  standard error of the mean (SEM) of the EPG counts were determined on each sampling day after treatment. AMs of the EPG counts were compared between treated and untreated groups by Student's t test. *P* values  $\leq 0.05$  and  $\leq 0.01$  were considered statistically significant or highly significant, respectively.

# Ethical Approval and/or Informed Consent

For this study, formal consent is not required. The research related to animals complied with all the relevant national regulations and institutional policies for the care and use of animals.

# **Results and Discussion**

On day 0, EPG counts ranged from 180 to 700 and from 100 to 770 in faeces of treated and untreated donkeys, respectively. No side effects were observed after treatment. FECR test showed that the polyherbal dewormer produced a very negligible reduction (5%) of strongyle eggs on day 0 onwards and no reduction or lower reduction (2 %) on day 21 till the end of the study, even though donkeys were treated again with a much higher dose, as adjunct treatment, on day 21. The values of AM±SEM of the EPG counts on day 0 to day 42 in treated and untreated donkeys are presented in Table 1. Unexpectedly, the AMs of the EPG counts were higher in the treated group both on day 14 and on day 21, reaching highly significant differences (P=0.0033 and P=0.0029, respectively). The values of the AMs in the treated donkeys became lower in comparison with their untreated counterpart on day 35 and on day 42 but differences did not reach statistically significant values (P=0.6530 and P=0.1796, respectively).

Results of the FECR test in the present study were compared to cut-off limits proposed by Kaplan and Nielsen (2010) as general guidelines to assess the occurrence of anthelmintic resistance after treatment with drugs currently used in horses, when a recommended range of at least 5 - 10 horses on each farm is included. Reductions >90 % for benzimidazoles and pyrantel or >95 % for ivermectin and moxidectin suggest that the strongyle population is

susceptible to the drug tested. Lack of effectiveness is suspected when reductions <80 % for benzimidazoles and pyrantel or <90 % for macrocyclic lactones are found. In this study, only a very negligible reduction (5 %) was seen on day 14. In general, the anthelmintic activity of plants has been reported to be lower than that found for synthetic anthelmintic drugs (Macedo et al., 2010). In addition, Igbal et al. (2004) reported that an increase in EPG reduction was noted with an increase in the dose of Artemisia brevifolia administered as crude powder, crude aqueous extract, and crude methanol extract against mixed infection of gastrointestinal nematodes in sheep. Similarly, Zajac and Gipson (2000) reported that a single treatment with fenbendazole was able to reduce EPG counts by only 50 % but 2 doses administered in a 12 h interval increased efficacy to 92 % in a goat herd. Thus, it is possible that the dose rate (8 gr of paste/100 kg bodyweight) recommended by the manufacturer for the polyherbal dewormer tested in our study may not be high enough to produce anthelmintic effects in donkeys. For this reason, a second treatment with a much higher dose of the product (50 gr of paste irrespective of the bodyweight) was additionally administered as adjunct treatment on day 21. Despite of this, no egg reduction could be demonstrated on day 35. Moreover, no important differences in individual FECR tests were observed among donkeys treated with the herbal dewormer. Finally, the occurrence of anthelmintic resistance can be excluded since the phytotherapeutic product tested had never been used before in the donkey population of the study. Therefore, our results show that the polyherbal dewormer administered in the present study failed to produce any suitable anthelmintic efficacy against intestinal strongyle infections in donkeys after both the first and mostly the second administration.

Our findings are in agreement with those of other authors. Lugin-

285

1115

835

1005

195

435

645 ± 159.5

215

400

585

640

285

305

 $405 \pm 70.3$ 

Groups Donkevs EPG counts Day 35 Day 42 Day 0 Day 14 Day 21 Treated 380 610 325 1000 230 I 180 120 230 315 180 Ш 240 440 570 305 495 IV 580 480 480 745 160 V 700 325 230 490 395 VI 630 600 555 865 440 VII 200 190 175 275 140 VIII 370 350 325 510 260 AM ± SEM 410 ± 71.9 389.4 ± 62.9<sup>a</sup> 361.2 ± 54.6<sup>b</sup> 563.1 ± 97.7  $287.5 \pm 48.4$ 

185

100

150

225

150

95

150.8 ± 20.3<sup>b</sup>

215

115

170

45

60

15

103.3 ± 31.7<sup>a</sup>

Table 1. Individual counts and arithmetic means ± standard error of the mean (AM±SEM) of eggs per gram (EPG) of faeces from day 0 to day 42 in donkeys treated with a commercial polyherbal dewormer on day 0 and on day 21 (I to VIII), and in donkeys of the control group kept untreated (IX to XIV).

<sup>a, b</sup>highly significant differences

IX

Х

XI

XII

XIII

XIV

AM ± SEM

770

410

280

110

100

200

 $311.6 \pm 103$ 

Control

buhl et al. (2006) reported the lack of anthelminthic effects of a commercially available herbal dewormer to reduce EPG counts in meat goats. Burke et al. (2009a) showed that another marketed herbal dewormer, involving the use of two different formulations, was ineffective to control gastrointestinal nematodes in dairy goats. The two herbal mixtures contained Artemisia absinthium. Allium sativum, Foeniculum vulgare, Juglans nigra, and Stevia rebaudiana as well as Cucurbita pepo, Hyssopus officinalis, and Thymus vulgaris. Similarly, a commercially available certified organic garlic product, fresh garlic juice, and garlic bulbs or papaya seeds failed to control gastrointestinal strongyles in goats and lambs (Burke et al., 2009b). In a field trial to assess the usefulness of a mineral lick containing herbal extracts with anti-parasitic properties for the control of gastrointestinal helminths in grazing sheep, there were no significant differences in the prevalence and intensity of helminth infections between the treated and control groups (Nosal et al., 2016). The commercially available product consisted of the extracts of A. absinthium, Artemisia cina, Tanacetum vulgare, T. vulgaris, A. sativum, Alsidium helminthochorton, Dryopteris filix-mas, Daucus carota, Chenopodium, and Punica granatum. However, contrary to the aforementioned studies, other in vivo investigations have reported the efficacy of some medicinal plants against gastrointestinal strongyles. Igbal et al. (2004), Masamha et al. (2010), Jabbar et al. (2007), and Tarig et al. (2009) reported the anthelmintic activity of A. brevifolia, A. sativum, Chenopodium album and Caesalpinia crista, or A. absintium in sheep, respectively.

To conclude, the phytotherapeutic potential and pattern of anthelmintic effectiveness of plant extracts vary widely from study to study. Consequently, though phytotherapy has become more and more popular with both donkey breeders and veterinarians, phytotherapic compounds marketed as herbal anthelmintics need evidence-based validation. For this reason, our trial tested the effectiveness of an herbal dewormer against intestinal strongyles in naturally infected donkeys. Based on the FECR test and current criteria used to evaluate the efficacy of anthelmintic drugs, the marketed polyherbal product tested in this study cannot be recommended as an effective alternative or complementary treatment for the control of intestinal strongylosis in donkeys. The findings of our study can help make practitioners aware of the possible disadvantages of natural herbal medicines for the treatment of parasitoses. In our opinion, monitoring FECR test results after treatment with herbal medicines is strongly recommended to provide the equine practitioner with the information necessary to administer extracts of anthelmintic plants consciously. Further in vivo trials focusing on suitable plants with promising potential anthelmintic properties are advisable for the development and the release of commercially available alternative and complementary anthelmintics to be used in donkeys.

# **Conflict of Interest**

Authors state no conflict of interest.

#### Acknowledgements

This study was funded by University of Pisa. We also would like to thank Ente Terre Regionali Toscane for allowing us to use of the animals for the purpose of the study.

# References

BELLAW, J.L., KREBS, K., REINEMEYER, C.R., NORRIS, J.K., SCARE, J.A., PAGANO, S., NIELSEN, M.K. (2018): Anthelmintic therapy of equine cyathostomin nematodes – larvicidal efficacy, egg reappearance period, and drug resistance. *Int. J. Parasitol.*, 48 (2): 97 – 105. DOI: 10.1016/j.ijpara.2017.08.009

BONELLI, F., BUSECHIAN, S., MEUCCI, V., CAPORRINO, G., BRIGANTI, A., RUECA, F., ZAPPULLA, F., FERINI, E., GHIANDAI, L., SGORBINI, M. (2016): pHyloGASTRO in the treatment of equine gastric ulcer lesions. *J. Equine Vet. Sci.*, 46: 69 – 72. DOI: 10.1016/j.jevs.2016.06.069 BURKE, J.M., WELLS, A., CASEY, P., KAPLAN, R.M. (2009a): Herbal dewormer fails to control gastrointestinal nematodes in goats. *Vet. Parasitol.*, 160 (1 – 2): 168 – 170. DOI: 10.1016/j.vetpar.2008.10.080

BURKE, J.M., WELLS, A., CASEY, P., MILLER, J.E. (2009b): Garlic and papaya lack control over gastrointestinal nematodes in goats and lambs. *Vet. Parasitol.*, 159 (2): 171 – 174. DOI: 10.1016/j.vet-par.2008.10.021

CAMILLO, F., ROTA, A., BIAGINI, L., TESI, M., FANELLI, D., PANZANI, D. (2018): The current situation and trend of donkey industry in Europe. *J. Equine Vet. Sci.*, 65: 44 – 49. DOI: 10.1016/j. jevs.2017.11.008

DIBABA, M.D., GETACHEW, A.M., ASSEFA, Z., FANTA, A., ETANA, M., FIREW, S., GOSHU, L., BURDEN, F. (2017): Seasonal variation of strongylosis in working donkeys of Ethiopia: a cross-sectional and longitudinal studies. *Parasitol. Res.*, 116 (7): 2009 – 2015. DOI: 10.1007/s00436-017-5485-z

Getachew, M., TRAWFORD, A., FESEHA, G., REID, S.W.J. (2010): Gastrointestinal parasites of working donkeys of Ethiopia. *Trop. Anim. Health Prod.*, 42 (1): 27 - 33.

DOI: 10.1007/s11250-009-9381-0

IQBAL, Z., LATEEF, M., ASHRAF, M., JABBAR, A. (2004): Anthelmintic activity of *Artemisia brevifolia* in sheep. *J. Ethnopharmacol.*, 93 (2 – 3): 265 – 268. DOI: 10.1016/j.jep.2004.03.046

ISMAIL, A.A., AHMED, N.K., BASHAR, A.E., SERI, H.I., TIGANI-ASIL, E., ABAKAR, A.D. (2016): A survey of seasonal gastrointestinal parasitic infections in donkeys from a semiarid sub-Saharan region, Sudan. *J. Pathog.*, 2016:4602751. DOI: 10.1155/2016/4602751

JABBAR, A., ZAMAN, M.A., IQBAL, Z., YASEEN, M., SHAMIM, A. (2007): Anthelmintic activity of *Chenopodium album* (L.) and *Caesalpinia crista* (L.) against trichostrongylid nematodes of sheep. *J. Ethnopharmacol.*, 114 (1): 86 – 91

DOI: 10.1016/j.jep.2007.07.027

JAJERE, S.M., LAWAL, J.R., BELLO, A.M., WAKIL, Y., TURAKI, U.A., WAZIRI, I. (2016): Risk factors associated with the occurrence of

gastrointestinal helminths among indigenous donkeys (*Equus asinus*) in Northeastern Nigeria. *Scientifica*, 2016:3735210. DOI: 10.1155/2016/3735210

KAPLAN, R.M., NIELSEN, M.K. (2010): An evidence-based approach to equine parasite control: it ain't the 60s anymore. *Equine Vet. Educ.*, 22 (6): 306 – 316. DOI: 10.1111/j.2042-3292.2010.00084.x LAWSON, E., BURDEN, F., ELSHEIKHA, H.M. (2015): Pyrantel resistance in two herds of donkey in the UK. *Vet. Parasitol.*, 207 (3 – 4): 346 – 349. DOI: 10.1016/j.vetpar.2014.12.026

LUGINBUHL, J.-M., PIETROSEMOLI CASTAGNI, S., HOWELL, J.M. (2006): Uso de un anthelmíntico botánico para el control de nemátodos gastrointestinales en caprinos de carne. [Use of an herbal dewormer for the control of gastric intestinal tract nematodes in meat goats]. *Arch. Latinoam. Prod. Anim.*, 14 (3): 88 – 89 (In: Spanish). MACEDO, I.T., BEVILAQUA, C.M., DE OLIVEIRA, L.M., CAMURÇA-VASCON-CELOS, A.L., VIEIRA LDA, S., OLIVEIRA, F.R., QUEIROZ-JUNIOR, E.M., TOMÉ ADA, R., NASCIMENTO, N.R. (2010): Anthelmintic effect of *Eucalyptus staigeriana* essential oil against goat gastrointestinal nematodes. *Vet. Parasitol.*, 173 (1 – 2): 93 – 98.DOI: 10.1016/j. vetpar.2010.06.004

MASAMHA, B., GADZIRAYI, C.T., MUKUTIRWA, I. (2010): Efficacy of *Allium sativum* (garlic) in controlling nematode parasites in sheep. *Inter. J. Appl. Res. Vet. Med.*, 8 (3): 161 – 169.

MATTHEE, S., KRECEK, R.C., MILNE, S.A. (2000): Prevalence and biodiversity of helminth parasites in donkeys from South Africa. *J. Parasitol.*, 86 (4): 756 – 762. DOI: 10.1645/0022-3395(2000)086[0756:PABOHP]2.0.CO;2

MATTHEE, S., KRECEK, R.C., MILNE, S.A., BOSHOFF, M., GUTHRIE, A.J. (2002): Impact of management interventions on helminth levels, and body and blood measurements in working donkeys in South Africa. *Vet. Parasitol.*, 107 (1 - 2): 103 – 113. DOI: 10.1016/S0304-4017(02)00113-9

MATTHEWS, J.B., BURDEN, F.A. (2013): Common helminth infections of donkeys and their control in temperate regions. *Equine Vet. Educ.*, 25 (9): 461 – 467. DOI: 10.1111/eve.12018

Nosal, P., Murawski, M., Bartlewski, P.M., Kowal, J., Skalska, M.,

 $Z_{I \notin BA}$ , D.A. (2016): Assessing the usefulness of mineral licks containing herbal extracts with anti-parasitic properties for the control of gastrointestinal helminths in grazing sheep – a field trial. *Helminthologia*, 53 (2): 180 – 185. DOI: 10.1515/helmin-2016-0008

RINALDI, L., LEVECKE, B., BOSCO, A., IANNIELLO, D., PEPE, P., CHARLIER, J., CRINGOLI, G., VERCRUYSSE, J. (2014): Comparison of individual and pooled faecal samples in sheep for the assessment of gastrointestinal strongyle infection intensity and anthelmintic drug efficacy using McMaster and Mini-FLOTAC. *Vet. Parasitol.*, 205: 216 – 223. DOI: 10.1016/j.vetpar.2014.06.011

SCANTLEBURY, C.E., PEACHEY, L., HODGKINSON, J., MATTHEWS, J.B., TRAWFORD, A., MULUGETA, G., TEFERA, G., PINCHBECK, G.L. (2013): Participatory study of medicinal plants used in the control of gastrointestinal parasites in donkeys in Eastern Shewa and Arsi zones of Oromia region, Ethiopia. *BMC Vet. Res.*, 9: 179. DOI: 10.1186/1746-6148-9-179

SGORBINI, M., BONELLI, F., PAPINI, R., BUSECHIAN, S., BRIGANTI, A., LAUS, F., FAILLACE, V., ZAPPULLA, F., RIZK, A., RUECA, F. (2017): Equine gastric ulcer syndrome in adult donkeys: investigation on prevalence, anatomical distribution and severity. *Equine Vet. Educ.*, 30 (4): 206 – 210. DOI: 10.1111/eve.12747

SGORBINI, M., VERONESI, F., FRATINI, M., LAUS, F. (2018): Tick-borne diseases and gastric ulcer in the donkey. *J. Equine Vet. Sci.*, 65: 62 – 65. DOI: 10.1016/j.jevs.2017.12.014

TARIQ, K.A., CHISHTI, M.Z., AHMAD, F., SHAWL, A.S. (2009): Anthelmintic activity of extracts of *Artemisia absinthium* against ovine nematodes. *Vet. Parasitol.*, 160 (1 – 2): 83 – 88. DOI: 10.1016/j. vetpar.2008.10.084

YOSEPH, S., SMITH, D.G., MENGISTU, A., TEKLU, F., FIREW, T., BETERE, Y. (2005): Seasonal variation in the parasite burden and body condition of working donkeys in East Shewa and West Shewa regions of Ethiopia. *Trop. Anim. Health Prod.*, 37 (Suppl 1): 35 – 45. DOI: 10.1007/s11250-005-9004-3

ZAJAC, A.M., GIPSON, T.A. (2000): Multiple anthelmintic resistance in a goat herd. *Vet. Parasitol.*, 87 (2 – 3): 163 – 172. DOI: 10.1016/S0304-4017(99)00174-0