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2 **Essential oils as eco-friendly biopesticides? Challenges and constraints**

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12 **Keywords:** botanical pesticides; microencapsulation; nanosynthesis; natural product research;
13 stabilization processes

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Recently, a growing number of plant essential oils (EOs) have been tested against a wide range of arthropod pests with promising results. EOs showed high effectiveness, multiple mechanisms of action, low toxicity on non-target vertebrates and possible use of by-products as reducing and stabilizing agents for the synthesis of nanopesticides. However, the number of commercial biopesticides based on EOs is still low. We analyse the main strengths and weaknesses arising from the use of EO-based biopesticides. Key challenges for future research include (i) development of efficient stabilization processes (e.g. microencapsulation), (ii) simplification of the complex and costly biopesticide authorization requirements, (iii) optimization of plant growing conditions and extraction processes leading to homogeneous chemical compositions of EOs.

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The phenomenon of essential oils

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Essential oils (EOs) are synthesized through secondary metabolic pathways of plants as communication and defence molecules. Generally, EOs play important roles in direct and indirect plant defences against herbivores and pathogens, in plant reproduction processes through attraction of pollinators and seed disseminators, as well as in plant thermotolerance [1]. The synthesis and accumulation of EOs are associated with the presence of secretory structures such as glandular trichomes (e.g. Lamiaceae), secretory cavities (e.g. Myrtaceae and Rutaceae), and resin ducts (e.g. Asteraceae and Apiaceae) [2], which can be found in various plant organs. Substances contained in EOs are classified into two chemical groups based on the metabolic pathway of their synthesis: (i) terpenoids, which are mainly represented by monoterpenes and less commonly by sesquiterpenes, and (ii)

39 phenylpropanoids with low molecular weight. The main metabolic pathways of EO synthesis
40 are shown in Box 1, Figure I [1, 3, 6].

41 Besides their communication and defence roles, EOs are responsible for the specific
42 flavour and scent of aromatic plants [3, 4]. These characteristics, together with their diverse
43 biological activities [5], have attracted high interest from industry, including food processing,
44 perfumery, and medicine [6]. Pesticides protect against many pathogens (e.g., [1, 7, 8]) and
45 arthropod pests (e.g., [9, 10]), including insects of high medical and veterinary importance
46 (see Supplementary Data Figure S1 online). Thus, many EOs are currently considered for the
47 development of plant protection products.

48 Many studies are published every year (see Supplementary Data Figure S2 online),
49 indicating great prospects of EOs as active ingredients for the production of botanical
50 pesticides [11]. Nevertheless, only very few commercial products based on EOs have been
51 marketed and the number of newly introduced products remains minimal. This paradox has
52 been discussed recently [1, 12, 13] raising the two main questions: *(i)* What are the causes for
53 this low rate of converting research results into practical application? *(ii)* Which direction
54 should the research take in order to change this unfavourable development? Here we attempt
55 to provide some answers to these questions, and at the same time, we outline key challenges
56 and potential trends for this area of fast-growing research.

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58 **Botanical pesticides based on essential oils**

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60 Pesticides have become a regular part of our lives. The yield of annual crop production
61 heavily relies on the application of pesticides [13]. However, frequent applications of some
62 synthetic pesticides has led to several problems [14]. For example, residues of many
63 pesticides can be detected in foods at concentrations above recommended limits with negative

64 effects on human health [15, 16]. Also, the effect of pesticides on non-target organisms is
65 important [17]. Lastly, the development of pathogen and insect populations resistant against
66 one or more synthetic pesticides is a major problem [18].

67 The above-mentioned issues culminated in the second half of the 20th century and
68 resulted in the current efforts to reduce the use of pesticides [19]. Eco-friendly alternatives
69 include botanical pesticides (BPs) produced from plant metabolites [1, 9, 13]. BPs, based on
70 EOs, exploit the toxicity of aromatic hydrocarbons contained in the oils. EOs, including their
71 active substances (see Supplementary Data Figure S1 online), show good biological activity
72 and provide insecticidal, nematicidal, ovicidal, fungicidal, and bactericidal effects against
73 pathogens and pests that are important factors in agricultural yield [1]. In addition, EOs are
74 able to inhibit growth, food intake, and oviposition in a number of important pests [1, 7-10,
75 20, 21]. EOs may be composed of several dozen (usually 20-60) active substances, which are
76 characterized by two or three main compounds at high concentrations (20–85%) as well as
77 other components, present at trace levels [5, 7]. Although little is known about the
78 mechanism(s) of action of individual compounds, evidence so far indicates that effects of
79 most compounds differ in the mechanism(s) of action. However, one common mode of action
80 for EOS that has been observed is based on their ability to disrupt cell wall and cytoplasmic
81 membrane of the bacteria and fungi, leading to lysis and leakage of intracellular compounds
82 or reported increased uptake of PI and leakage of K⁺ [22-26], able to cause structural
83 alterations of the outer envelope without promoting the release of cellular content. These cell
84 surface changes were sufficient to induce cell death. In addition, compounds from EOs can
85 exert their activities on insects through neurotoxic effects involving several mechanisms,
86 notably through GABA, octopamine synapses, and the inhibition of acetylcholinesterase
87 (Table 1).

88 Complex mixtures of substances contained in EOs, with different mechanisms of
89 action and often exhibiting mutual synergistic relationships [27-29], may be efficient in
90 preventing the development of resistant pathogen and pest populations. This is one of several
91 important benefits of BPs based on EOs (Box 2).

92

93 **So why is the number of commercial products so low?**

94 Although being very efficient as potential active ingredients, only few commercially
95 manufactured BPs based on EOs exist. In our opinion, this is due to four main reasons: (i)
96 Many published studies, but only few practical results; (ii) Strict legislation; (iii) Low
97 persistence of effects; (iv) Lack of quality and sufficient quantity of materials for affordable
98 prices.

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100 *Many published studies, but only few practical results*

101 As noted by Isman and Grieneisen [12], most of the published studies of biological efficacy of
102 EOs are based on screening the efficacy of one or more EOs against one or more target
103 organisms. Most studies are thus only the first step in the development of new BPs. Only very
104 few studies have dealt with the effect of EOs on non-target organisms, while such researches
105 are important for the development and authorization of BPs [30-32]. For example, EOs may
106 have a significant effect on insects even in sublethal doses [33]. However, this phenomenon
107 has been rarely studied. In addition, the number of studies focusing on the mutual
108 relationships of individual compounds contained in EOs is low [27-29], although this
109 information is crucial for the development of standardized BP formulations. We believe that
110 closer cooperation between scientific centres and potential manufactures in research is
111 missing – i.e. a type of cooperation that would lead from taking *in vitro* data on the

112 verification of biological efficacy all the way through to field experiments on pests attacking
113 plants and livestock.

114

115 *Strict legislation*

116 The BP authorization processes are complex and costly, as well as the authorization of any
117 newly synthesized compound with no history of use in the food, cosmetics or pharmaceutical
118 industry. Authorizations in EU states require safety documentation through appropriate
119 toxicological studies. However, in many cases, such studies do not exist and their preparation
120 is too expensive for local manufacturers. The manufacture of BPs, often of only local
121 importance, is usually at low scale, because production is restricted by limited availability of
122 active substances. BP manufacturers therefore try to market such products outside the scope
123 of the authorization process, for example, as fragrances or fertilizers with a secondary
124 pesticidal benefit. However, this practice is desolating for many manufacturers, because they
125 cannot openly declare the efficacy against pests on the label, which usually result in low sales.
126 BP manufacturers in the EU look with certain hope towards Regulation (EC) No. 1107/2009
127 (<http://eur-lex.europa.eu/eli/reg/2009/1107/oj>), which has introduced a new term – “basic
128 substances” (BS). This regulation [(EC) No. 1107/2009 Article 23] provides criteria for the
129 approval of BS with specific provisions to ensure that such active substances can be legally
130 used in the EU, as far as they do not have an immediate or delayed harmful effect on human
131 and animal health nor an unacceptable effect on the environment. However, this regulation
132 may not be easily applicable for the use of EOs, because the concept behind the regulation
133 was that various food additives, without any further formulation adaptations (e.g. without the
134 use of emulsifiers or additives), could be authorized as BSs. Therefore, the process for
135 approval of some active ingredients, such as EOs, could be more complicated, because for
136 their application the use of emulsifiers is necessary. Specifically, any products deviating from

137 the definition of Regulation (EC) No. 1107/2009 Article 23, including the ones containing, for
138 example, an already approved “BS” but contain a co-formulant will then be considered as
139 plant protection product and no longer qualifies as BS [34]. These strict criteria have been set
140 particularly to protect human health against risky pollutants. Among others, this was
141 motivated also by the finding of high food contamination with pesticide residues in EU states,
142 detected in analyses performed at the beginning of the 21st century (see,
143 <http://www.pesticide-residues.org/food.html>).

144 Natural substances face a number of opponents among the European Commission members
145 responsible for authorization processes, being viewed by some even as more hazardous than
146 synthetic compounds. These opponents use precisely the lack of relevant toxicological data
147 for natural products as an argument against their use. In particular, there are fears of potential
148 mutagenic or genotoxic adverse effects and negative effects on the human endocrine system.
149 However, according to available information, most of EOs and their main compounds have
150 been reported to be not mutagenic/genotoxic [5]. However, the genotoxic response may be
151 affected by the experimental model chosen and the range of concentrations assayed, and this
152 could be due in part to the induction of oxidative stress. However, some of them can be
153 metabolically activated, such as cinnamaldehyde, or they can be metabolized to a substance
154 without genotoxic activity, such as in the case of linalool [35]. Most negative effects of EOs
155 appear with high dosages, application of undiluted EO concentrates or upon long-term
156 exposure. In terms of toxicology, it should be noted that most EOs show only low acute
157 toxicity, >2 g/kg for both oral and dermal application (Table 2). Given that any residues on
158 plants are minimal (given the fumigation and degradation nature of EOs) [36], a number of
159 EOs cannot be considered as risky substances.

160

161 *Low persistence of effects*

162 Essential oils are composed of lipophilic and highly volatile secondary plant metabolites,
163 reaching a mass below a molecular weight of 300. Terpenoids tend to be both volatile and
164 thermolabile and may be easily oxidized or hydrolyzed depending on their respective
165 structure [37]. Thus, the chemical composition of essential oils is dependent on the conditions
166 during processing and storage of the plant material, upon distillation as well as in the course
167 of subsequent handling of the oil itself [36, 38]. Thus, these features may significantly reduce
168 the efficacy of Eos against pests. Although the contact effect of EOs is very good, rapid
169 fumigation into the environment and gradual biodegradation of active substances occur upon
170 application on the plants with low persistence of the effect. High attention should be given to
171 the development of suitable EO formulations as active ingredients of BPs that would show
172 higher persistence of efficacy. However, this research is only beginning, and mostly focused
173 on suitable encapsulation methods. Encapsulation is a process in which an active component
174 is entrapped or coated by a matrix wall. This matrix isolates the bioactive molecule from the
175 surrounding environment until its release in response to external conditions (pH, pressure,
176 temperature, etc.) [36]. The wall material can be selected from a wide range of natural or
177 synthetic polymers according to the desired characteristics of the final delivery system [39].
178 Although a number of EO encapsulation methods exist, developed predominantly for food
179 industry and pharmaceutical purposes [40], especially inexpensive encapsulation methods are
180 needed for the application of EOs as BP. Among the existing methods, coacervation, also
181 known as phase separation, seems the most suitable solution. In terms of the use of EOs for
182 BPs, simple coacervation is suitable, which uses one polymer, such as gelatine or ethyl
183 cellulose [41]. The use of cyclodextrins (CD) may be another suitable method. CDs are cyclic
184 glucose oligomers having six, seven or eight glucose units linked by α -1,4-glucosidic bonds,
185 called, respectively, α -, β - and γ -CD [42]. The use of CD-complexation is widespread in
186 pharmaceutical applications, foods, cosmetics, and toiletries. CDs may be considered as

187 nanoencapsulating agents and the complex formation is equivalent to molecular
188 encapsulation. The bioactive EO molecules are isolated from each other and dispersed on a
189 molecular level in an oligosaccharide matrix [40].

190 Suitable encapsulation methods, as well as some nanoparticle synthesis methods –
191 AgNP [43, 44] or understanding of the synergistic relationships [27-29], may finally result in
192 an increase of biological activity of BPs based on EOs and thus to an extension of their
193 persistence of efficacy, which is a very important part of research for manufacturers.

194

195 *Lack of quality and sufficient quantity of materials for affordable prices*

196 EOs are produced in 17,500 aromatic species of higher plants belonging mostly to a few
197 families, including the Apiaceae, Myrtaceae, Lauraceae, Lamiaceae, and Asteraceae.
198 However, only a small proportion (approximately 300 species) has found use in commercial
199 application [1]. A great part of promising EOs originates from plants whose cultivation is
200 expansive or disadvantageous due to low EO yields. Not even plants that are currently grown
201 for commercial production of EOs can be cultivated easily. One of the reasons is that the
202 physiological expression of plant secondary metabolism can differ at all developmental
203 stages. Furthermore, the proportions of monoterpenes depend on temperature and circadian
204 rhythm and vary according to the plant phenological phase [46, 47]. Finally, soil acidity and
205 climate (heat, photoperiod, and humidity) directly affect the secondary metabolism of the
206 plant [1] and EO composition. Therefore a standardized product, which is important for
207 regulatory and marketing purposes, is a timely challenge [1, 5]. To address this challenge,
208 elicitation products, genetic manipulations or new technologies of growing plants have been
209 suggested, aimed at increasing the production and standardizing qualitative and quantitative
210 parameters of EOs [45, 48, 49]. New methods for isolating EOs from plants have also been
211 investigated. At present, EOs are isolated from plants using conventional/classical methods,

212 i.e. using standard distillation of the plant material. Investing in new technologies (e.g.
213 ultrasounds, microwaves) in the last decades has led to the emergence of innovative and more
214 efficient extraction processes (i.e. reduction of extraction time and energy consumption,
215 increase of extraction yield, improvement of EOs quality) [50].

216 These new trends in the research of aromatic plants, together with the choice of
217 suitable chemotypes showing high yields or better biological efficacy [33, 48], will open new
218 prospects for the sustainable production and practical employment of EOs.

219

220 **Concluding remarks and future perspectives**

221 In our opinion, although a huge amount of studies have been published, focused on biological
222 activity of EOs on target organisms, papers concerning toxicological studies and effects of
223 EOs on non-target organisms are missing (see Outstanding Questions). Similarly, the
224 mechanisms of action have not been fully clarified, including mutual relationships among
225 individual substances in EOs and the effects of sublethal concentrations on target and non-
226 target organisms. However, despite those shortcomings, based on available toxicological
227 studies, we conclude that most EOs raise no concerns of their use in plant and livestock
228 protection and can be considered as safe for environment and human health in common
229 concentrations or doses (Box 2). Therefore, the existing legislation concerning authorization
230 should be simplified and better cooperation should be established between research and BP
231 manufacturers in order to put research results into practice. This is a key challenge, because
232 BPs based on EOs have the potential to provide a significant improvement in the quality and
233 safety of foods, including human health, which should be a priority for all food-producing
234 nations worldwide.

235

236 **Acknowledgments**

237 Roman Pavela is supported by grants from the Ministry of Agriculture of the Czech Republic
238 (Project NAZV No. QJ1510160). Giovanni Benelli is supported by PROAPI (PRAF 2015)
239 and University of Pisa, Department of Agriculture, Food and Environment (Grant ID:
240 COFIN2015_22). Funders had no role in the study design, data collection and analysis,
241 decision to publish, or preparation of the manuscript.

242

243 **Disclosure**

244

245 RP has been engaged in the development of botanical pesticides for more than 20
246 years. At present, he is the head of the team “Secondary Plant Metabolites in Crop
247 Protection“. RP cooperates with several BP manufacturers in the development of new
248 products, of which more than 10 are currently available on the market. RP has been awarded
249 many prizes for his research of BPs including the highest award of the Government of the
250 Czech Republic “Česká hlava” (“The Czech Head“).

251 GB is an entomologist focused on insect-plant interactions, and the development of
252 novel control tools in the fight against arthropod pests. GB is cooperating with more than 80
253 researches worldwide and has published more than 150 researches. He has been involved in
254 several international projects, including FP7 Project CoCoRo, GA 270382, ICMR Project
255 15200, and H2020 Project subCULTron, GA 640967FP7.

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257 **References**

258

- 259 1. Regnault-Roger, C. et al. (2012) Essential oils in insect control: low-risk products in
260 a high-stakes world. *Annu. Rev. Entomol.* 57, 405–424
- 261 2. Rodriguez, E. et al. (1984) *Biology and Chemistry of Plant Trichomes*. NewYork: Plenum

- 262 3. Nagegowda, D.A. (2010) Plant volatile terpenoid metabolism: Biosynthetic genes,
263 transcriptional regulation and subcellular compartmentation. *FEBS Letters* 584, 2965–2973
- 264 4. Dudareva, N. et al. (2006) Plant volatiles: recent advances and future perspectives. *Crit.*
265 *Rev. Plant Sci.* 25, 417–440
- 266 5. Bakkali, F. et al. (2008) Biological effects of essential oils—a review. *Food Chem. Toxicol.*
267 46, 446–475
- 268 6. Aharoni, A. et al. (2005) Volatile science? Metabolic engineering of terpenoids in plants.
269 *Trends Plant Sci.* 10 doi:10.1016/j.tplants.2005.10.005
- 270 7. Burt, S. (2004) Essential oils: their antibacterial properties and potential applications in
271 foods—a review. *Int. J. Food Microbiol.* 94, 223–253
- 272 8. Zabka, M. et al. (2004) Antifungal activity and chemical composition of twenty essential
273 oils against significant indoor and outdoor toxigenic and aeroallergenic fungi. *Chemosphere.*
274 112, 443–448
- 275 9. Pavela, R. (2015) Essential oils for the development of eco-friendly mosquito larvicides: A
276 review. *Ind. Crop. Prod.* 76, 174–187
- 277 10. Nerio, L.S. et al. (2010) Repellent activity of essential oils: a review. *Bioresour. Technol.*
278 101, 372–378
- 279 11. Isman, M.B. et al. (2011) Commercial opportunities for pesticides based on plant
280 essential oils in agriculture, industry and consumer products. *Phytochem. Rev.* 10, 197–
281 204
- 282 12. Isman, M.B. and Grieneisen, M.L. (2014) Botanical insecticide research: many
283 publications, limited useful data. *Trends Plant Sci.* 19, 140–145
- 284 13 Isman MB (2015) A renaissance for botanical insecticides? *Pest Manag. Sci.* 71, 1587–
285 1590

- 286 14. Naqqash MN, Gökçe A, Bakhsh A, Salim M (2016) Insecticide resistance and its
287 molecular basis in urban insect pests. *Parasitol. Res.* 115, 1363–1373
- 288 15. Beard, J. et al. (2003) Health impacts of pesticide exposure in a cohort of outdoor
289 workers. *Environ. Health Perspect.* 111, 724–730
- 290 16. Mei, C. et al. (2015) Residential Exposure to Pesticide During Childhood and Childhood
291 Cancers: A Meta-Analysis. *Pediatr.* 136, 719-729
- 292 17. Goulson, D. (2013) REVIEW: An overview of the environmental risks posed by
293 neonicotinoid insecticides. *J. Appl. Ecol.* 50, 977-987
- 294 18. McCaffery, A. and Nauen, R. (2006) The Insecticide Resistance Action Committee
295 (IRAC): public responsibility and enlightened industrial self interest. *Outlook Pest Manag*
296 17, 11–14
- 297 19. Marchand, P.A. (2015) Basic substances: an opportunity for approval of low-concern
298 substances under EU pesticide regulation. *Pest Manag. Sci.* 71, 1197–1200
- 299 20. Perez, S.G. et al. (2010) Activity of essential oils as a biorational alternative to control
300 coleopteran insects in stored grains. *J. Med. Pl. Res.* 4, 2827-2835
- 301 21. Pavela, R. (2011) Antifeedant and larvicidal effects of some phenolic components of
302 essential oils lasp lines of introduction against *Spodoptera littoralis* (Boisd.). *J. Essent. Oil*
303 *Brear. Pl.* 14, 266-273
- 304 22. Turgis, M. et al. (2009) Antimicrobial activity of mustard essential oil against *Escherichia*
305 *coli* O157:H7 and *Salmonella typhi*. *Food Control.* 20, 1073–1079
- 306 23. Devi, S.A. et al. (2010) Eugenol (an essential oil of clove) acts as an antibacterial agent
307 against *Salmonella typhi* by disrupting the cellular membrane. *J. Ethnopharm.* 130, 107–115
- 308 24. Bajpai, V.K. et al. (2013) Antibacterial mode of action of *Cudrania tricuspidata* fruit
309 essential oil, affecting membrane permeability and surface characteristics of food-borne
310 pathogens. *Food Control.* 32, 582–590

- 311 25. Lambert, R.J.W. et al. (2001) A study of the minimum inhibitory concentration and mode
312 of action of oregano essential oil, thymol and carvacrol. *J.Appl. Microbiol.* 91, 453–462
- 313 26. Tian, J. et al. (2012) The Mechanism of Antifungal Action of Essential Oil from Dill
314 (*Anethum graveolens* L.) on *Aspergillus flavus*. *PLoS ONE* 7(1), e30147
- 315 27. Pavela, R. (2014) Acute, synergistic and antagonistic effects of some aromatic compounds
316 on the *Spodoptera littoralis* Boisd. (Lep., Noctuidae) larvae. *Ind. Crop. Prod.* 60, 247-258
- 317 28. Pavela, R. (2015) Acute toxicity and synergistic and antagonistic effects of the aromatic
318 compounds of some essential oils against *Culex quinquefasciatus* Say larvae. *Parasitol. Res.*
319 114, 3835-3853
- 320 29. Tak, J.-H. et al. (2016) Comparative and synergistic activity of *Rosmarinus officinalis* L.
321 essential oil constituents against the larvae and an ovarian cell line of the cabbage looper,
322 *Trichoplusia ni* (Lepidoptera: Noctuidae). *Pest Manag. Sci.* 72, 474-480
- 323 30. Pavela, R. (2014) Insecticidal properties of *Pimpinella anisum* essential oils against the
324 *Culex quinquefasciatus* and the non-target organism *Daphnia magna*. *J. Asia Pacific Entomol.*
325 17, 287–293
- 326 31. Pavela, R. and Govindarajan, M (2016) The essential oil from *Zanthoxylum monophyllum*
327 a potential mosquito larvicide with low toxicity to the non-target fish *Gambusia affinis*. *J.*
328 *Pest Sci.* DOI 10.1007/s10340-016-0763-6
- 329 32. Benelli, G., Mehlhorn, H. (2016) Declining malaria, rising dengue and Zika virus: insights
330 for mosquito vector control. *Parasitol. Res.* 115, 1747-54
- 331 33. Pavela, R (2007) Lethal and sublethal effects of thyme oil (*Thymus vulgaris* L.) on the
332 house fly (*Musca domestica* Lin.). *J. Essent. Oil Bear Pl.* 10, 346–356
- 333 34. Pavela R (2014) Limitation of Plant Biopesticides. In *Advances in Plant Biopesticides*
334 (Dwijendra Singh, ed), pp. 347–359, Springer

- 335 35. Llana-Ruiz-Cabello, M., et al. (2015) In vitro toxicological evaluation of essential oils and
336 their main compounds used in active food packaging: A review. *Food Chem. Toxicol.* 81, 9-
337 27
- 338 36. Turek, C. and Stintzing, F.C. (2013) Stability of Essential Oils: A Review. *Compr. Rev.*
339 *Food Sci. Saf.* 12, 40–53
- 340 37. Scott, R.P.W. (2005) Essential oils. In: Worsfold P, Townshend A, Poole C, editors.
341 Encyclopedia of analytical science. 2nd ed. Amsterdam, London, New York: Elsevier. p 554–
342 561
- 343 38. Schweiggert U et al. (2007) Conventional and alternative processes for spice production –
344 a review. *Trends Food Sci. Technol.* 18, 260–268
- 345 39. De Barros Fernandes, R. V. et al. (2014). Effect of solids content and oil load on the
346 microencapsulation process of rosemary essential oil. *Ind. Crop Prod.* 58, 173–181
- 347 40. Rodríguez, J. et al. (2016) Current encapsulation strategies for bioactive oils: From
348 alimentary to pharmaceutical perspectives. *Food Res. Internat.* 83, 41–59
- 349 41. Jun-Xia, X. et al. (2011). Microencapsulation of sweet orange oil by complex
350 coacervation with soybean protein isolate/gum arabic. *Food Chem.* 125, 1267–1272.
- 351 42. Ciobanu, A. et al. (2013). Complexation efficiency of cyclodextrins for volatile flavor
352 compounds. *Food Res. Internat.* 53, 110–114
- 353 43. Benelli, G. (2015) Research in mosquito control: current challenges for a brighter future.
354 *Parasitol. Res.* 114, 2801–2805
- 355 44. Benelli, G. (2016) Plant-mediated biosynthesis of nanoparticles as an emerging tool
356 against mosquitoes of medical and veterinary importance: a review. *Parasitol Res.* 115, 23-34
- 357 45. Iannicelli, J. et al. (2016) Effect of polyploidization in the production of essential
358 oils in *Lippia integrifolia*. *Ind. Crop. Prod.* 81, 20–29

359 46. Clark, R.J. and Menary, R.C. (1981) Variations in composition of peppermint oil in
360 relation to production areas. *Econ. Bot.*35, 59–69

361 47. Hansted, L. et al. (1994) Influence of temperature on the rhythmic emission of volatiles
362 from *Ribes nigrum* flowers in situ. *Plant Cell Environ.*17,1069–1072

363 48. Pavela, R. et al. (2016) New knowledge for yield, composition and insecticidal activity of
364 essential oils obtained from the aerial parts or seeds of fennel (*Foeniculum vulgare* Mill.) *Ind.*
365 *Crop. Prod.* 83, 275-282

366 49. Mahmoud, SS and Croteau, RB (2002) Strategies for transgenic manipulation of
367 monoterpene biosynthesis in plants. *Trends Plant Sci.* 7, 366-373

368 50. El Asbahani, A. et al. (2015) Essential oils: From extraction to encapsulation. *Internat. J.*
369 *Pharmac.* 483, 220–243

370 51. Belzile, A.S. et al. (2000) Dillapiol derivatives as synergists: structure-activity
371 relationship analysis. *Pestic. Biochem. Physiol.* 66, 33–40

372 52. Priestley, C.M. et al. (2003) Thymol, a constituent of thyme essential oils, is a positive
373 modulator of human GABA and a homo-oligosteric GABA receptor from *Drosophila*
374 *melanogaster*. *Br. J. Pharmacol.* 140, 1363–1372

375 53 Mills, C. et al. (2004) Inhibition of acetylcholinesterase by tea tree oil. *J. Pharm.*
376 *Pharmacol.* 56:375–79

377 54. Lopez, M.D. and Pascual-Villalobos M.J. (2010) Mode of inhibition of
378 acetylcholinesterase by monoterpenoids and implications for pest control. *Ind. Crops Prod.*
379 31, 284–288

380 55. Enan, E.E. (2001) Insecticidal activity of essential oils: octopaminergic sites of action.
381 *Comp. Biochem. Physiol.* 130, 325–327

382 56. Enan, E.E. (2005) Molecular response of *Drosophila melanogaster* tyramine receptor
383 cascade to plant essential oils. *Insect Biochem. Mol. Biol.* 35, 309–321

384

385

386 **Table 1.** The most common mode of actions of essential oils.

Mode of action	Mechanism of inhibition	Examples of possible compounds	Refs.
Inhibitor of cytochromes P450 (CYPs)	Inhibitors of insect P450 cytochromes responsible for phase I metabolism of xenobiotics, including insecticides.	Dillapiole from <i>Anethum sowa</i> , piperamides from <i>Piper</i> spp.	51
GABA receptors	Compounds bind to GABA receptors associated with chloride channels located on the membrane of postsynaptic neurons and disrupts the functioning of GABA synapse.	Thymol from e.g. <i>Thymus vulgaris</i>	52
Inhibition of cholinergic system	Inhibition of acetylcholinesterase (AChE)	Fenchone from e.g. <i>Foeniculum vulgare</i> , S-carvone from e.g. <i>Mentha spicata</i> , linalool from e.g. <i>Citrus</i> spp.	53, 54
Modulators of ectopaminergic system	They act through the octopaminergic system by activating receptors for octopamine, which is a neuromodulator.	Eugenol from e.g. <i>Syzygium aromaticum</i> , α -terpineol from e.g. <i>Pinus sylvestris</i> .	55, 56

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390 **Table 2.** Acute toxicity of some essential oils on non-target vertebrates^a

Plant	LD ₅₀ mg/kg	
	Orally administered	Dermally administered
<i>Abies alba</i>	>5,000 (rat)	>5,000 (rabbit)
<i>Anethum graveolens</i>	4,040 (rat)	>5,000 (rabbit)
<i>Angelica archangelica</i>	>10,000 (rat)	>5,000 (rabbit)
<i>Apium graveolens</i>	>5,000 (rat)	>5,000 (rabbit)
<i>Cinnamomum camphora</i>	3,730 (rat)	>5,000 (rabbit)
<i>Citrus sinensis</i>	>5,000 (rat)	>5,000 (rabbit)
<i>Coriandrum sativum</i>	4,130 (rat)	N.I.
<i>Cymbopogon citratus</i>	>5,000 (rat)	>5,000 (rabbit)
<i>Elettaria cardamomum</i>	>5,000 (rat)	>5,000 (rabbit)
<i>Eugenia spp.</i>	2,650 (rat)	>5,000 (rabbit)
<i>Foeniculum vulgare</i>	3,120 (rat)	N.I.
<i>Lavandula angustifolia</i>	4,250 (rat)	>5,000 (rabbit)
<i>Melaleuca alternifolia</i>	1,900 (rat)	>5,000 (rabbit)
<i>Ocimum basilicum</i>	>5,000 (rat)	>5,000 (rabbit)
<i>Rosmarinus officinalis</i>	>5,000 (rat)	>10,000(rabbit)
<i>Thymus vulgaris</i>	2 840 (rat)	>5,000 (rabbit)
<i>Zingiber officinale</i>	3,400 (mouse)	N.I.

391 ^aAccording to Safety Data Sheets of Sigma Aldrich, N.I. = not indicated

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396 **Figure Legend**

397 **Figure I. MVA and MEP pathways.** Abbreviations: FPP, farnesyl diphosphate; DLG, D, L-glyceraldehyde; DMAPP,
398 dimethylallyl pyrophosphate; DXP, 1-deoxy-D-xylulose 5-phosphate; DXR, 1-deoxy-D-xylulose 5-phosphate
399 reductoisomerase; DXS, 1-deoxy-D-xylulose 5-phosphate synthase; GA-3P, glyceraldehyde 3-phosphate; GGPP,
400 Geranylgeranyl diphosphate; GPP, Geranyl diphosphate; GGPPS, Geranylgeranyl diphosphate synthase; HMGR,
401 3-hydroxy-3-methylglutaryl coenzyme A reductase; IPP, isopentenyl pyrophosphate; MEP, 2-C-methyl-d-
402 erythritol-4-phosphate.

403

404 **Box 1. Pathways of terpenoid biosynthesis in aromatic plants**

405 Monoterpenes are the most common group contained in essential oils. It is well known that the biosynthesis of
406 terpenoids in plants takes place via two main pathways (see Figure I): the mevalonate (MVA) pathway in the
407 cytosol and the methylerythritol phosphate (MEP) pathway in the plastids, which yields the 5-carbon
408 precursors isopentenyl pyrophosphate (IPP) and dimethylallyl pyrophosphate (DMAPP), which are condensed
409 via geranyl pyrophosphate synthase to give the 10- carbon monoterpenes (not shown). Although isopentenyl
410 pyrophosphate can move between compartments, the monoterpenes and diterpenes tend to be formed in the
411 plastid, where unique cyclases produce the ring structures. Monoterpenes present in essential oils may contain
412 terpenes that are hydrocarbons, alcohols, aldehydes, ketones, ethers, and lactones. The sesquiterpenes have a
413 wide variety of structures with more than 100 skeletons, because the elongation of the chain to 15 carbons
414 increases the number of possible cyclizations that are formed via the mevalonate pathway in the cytosol.
415 Aromatic compounds are less common and are derived mainly from the shikimate pathway, for example, the
416 phenylpropanoid dillapiole, but a few phenols, such as carvacrol and cuminaldehyde, are a rare group derived
417 from terpene biosynthesis by desaturation.

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420 **Box 2. Essential oils as botanical pesticides: advantages and future challenges**

421 Overall, the use of essential oils as botanical pesticides has shown a number of advantages including:

- 422 • High effectiveness against a wide number of pests and diseases of agricultural and medical
423 importance.
- 424 • Multiple mechanisms of actions: due to the large number of active ingredients in each blends, the
425 development of resistance is less likely
- 426 • Low toxicity against non-target organisms, including humans
- 427 • The production processes are relatively simple and cheap
- 428 • Low health risk during application due to low toxicity rates of residues

429 Key challenges for further research are:

- 430 • Simplification of the complex and costly authorization process to legitimize new botanical pesticides,
431 based on plant extracts with proven history of use in the food industry, cosmetics or medicine;
- 432 • Avoid loss of efficiency against target pests in the field, highlighting the needing of efficient
433 stabilization processes (e.g. encapsulation). Alternatively, the botanical-mediated synthesis of
434 effective nanoinsecticides could help to avoid high levels of degradation of active compounds from
435 essential oils.
- 436 • New production technologies that guarantee abundant quantities of raw essential oil sources with
437 homogeneous chemical composition.

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