

Manuscript Number: STOTEN-D-19-03844

Title: Exposure of coastal dune vegetation to plastic bag leachates: a neglected impact of plastic litter

Article Type: Research Paper

Keywords: Compostable bag, *Glaucium flavum*, Polyethylene bag, Seed germination, Seedling growth, *Thinopyrum junceum*

Corresponding Author: Dr. Elena Balestri, Ph.D.

Corresponding Author's Institution: University of Pisa

First Author: Virginia Menicagli

Order of Authors: Virginia Menicagli; Elena Balestri, Ph.D.; Claudio Lardicci

Abstract: The presence of plastic bags on coastal dunes worldwide is well documented. Plastic bags contain additives that during rainfall events can leach out from bags into sand dune and be absorbed by seeds and roots of plants. Dune plants play a fundamental role in dune system formation, yet the possible impact of bag leaching on their establishment and development has been neglected. We assessed in laboratory whether (i) not biodegradable bags (high-density polyethylene, PE) and new generation of compostable bags (Mater-bi®, MB) would influence via leaching water chemical/physical properties and (ii) leachates would affect germination and seedling growth using *Thinopyrum junceum* and *Glaucium flavum* as models. Leachates were obtained from different amounts of not-exposed and bags exposed to beach or marine conditions simulating various pollution degrees (none, low, intermediate and high pollution). All water variables were affected by leaching. The magnitude of these alterations depended on bag type and environmental exposure. Seeds of *T. junceum* treated with the high concentration of marine-exposed MB bag leachate germinated later than controls while those of *G. flavum* treated with the remaining leachates germinated earlier. For both species, leachates from the low concentration of PE and MB marine-exposed bags increased seed germinability. A short radicle was observed in *T. junceum* seedlings treated with not-exposed MB bag leachates. *Glaucium flavum* seedlings treated with beach- and marine-exposed PE bags and not-exposed MB bags leachates showed a greater below-aboveground length ratio and those grown with the low concentration of not-exposed PE bag leachate had a longer hypocotyl compared to controls. Leachates from the high concentration of PE and MB bag caused seedling anomalies in both species. These findings indicate that not biodegradable and compostable bags may interact with abiotic/biotic factors and affect via leaching germination phenology, seedling establishment and plant interactions with consequences on dune community structure.

Suggested Reviewers: John Griffin

Department of Biosciences Swansea, Wales, UK, United Kingdom
j.n.griffin@swansea.ac.uk

Roberto Simonini
Department of Life Sciences, University of Modena and Reggio Emilia,
Italy
roberto.simonini@unimore.it

Anna Traveset
Institut Mediterrani d'Estudis Avançats (CSIC/UIB), Spain
atraveset@imedea.uib-csic.es

Italo Castro
Universidade Federal de São Paulo
italobraga@gmail.com

Fei-Hai Yu
Taizhou University, Taizhou 318000, China
feihaiyu@126.com

Opposed Reviewers:

Dear Editor,

My co-authors (Virginia Menicagli and Claudio Lardicci) and I are pleased to submit the enclosed manuscript “Exposure of coastal dune vegetation to leachates from plastic bags: a neglected effect of plastic litter” as Research Article in *Science of the Total Environment*.

The paper focuses on plastic pollution, and in particular on the potential effects of plastic bags on dune plants via leaching, i.e. the release of chemicals from plastics in natural environments. Many previous studies on plastic litter have investigated the effects of bags on a variety of organisms via ingestion or entanglement, but only few studies have dealt with plastic bag leaching and its impact on plants. Recently, a study has demonstrated that when in contact with water, chemicals can be released from plastic bags and migrate into the water phase, contaminating water available to plants. However, the potential effects of this phenomenon on dune vegetation that strictly depend on the availability of water and grow in a habitat highly impacted by plastics are completely ignored. Therefore, in this study the effects of the leaching of conventional not biodegradable bags and biodegradable/compostable shopper bags on the most vulnerable life history stages of dune plants, germination and early seedling growth, were examined. Conventional bags are a consistent fraction of plastic litter found in coastal environments. Biodegradable/compostable bags made with Mater-bi[®] (a starch-based polymer) have been marketed in Europe to reduce the impact associated to plastic littering and disposal, but their presence in natural habitats has recently been observed. These bags are considered as an eco-friendly alternative to conventional ones, and they are specially designed to be disposed in industrial or home compost facilities at the end of their life. However, an increased number of studies indicate that once entered natural environments compostable bags degrade slowly and can affect marine organism and communities. Firstly, we performed a laboratory leaching experiment to examine whether (i) both types of bag would influence via leaching water chemical/physical variables relevant to dune plant development and (ii) the conditions experienced by bags before being deposited close to dune vegetation, i.e., seawater

immersion or sand burial would influence water variables. Then, we conducted a germination and seedling growth test to investigate whether bag leachates would affect seed germinability, germination phenology and seedling growth of *Thinopyrum junceum* (L.) Á. Löve and *Glaucium flavum* Crantz, two dune plant species belonging to different taxonomic groups.

The results of the leaching experiment demonstrate that conventional and compostable bags once deposited on sand dunes considerably altered via leaching chemical/physical characteristics of the water available to dune plants during rainfall events. Water quality depended on the type of bag material and the environmental conditions experienced by the bag. The results of the germination and early seedling growth test show that bag leachates interfered with factors controlling seed dormancy and seedling emergence. Both germination phenology and seedling growth of the two species are differentially affected by leachates, and the direction and the magnitude of these effects depended on type and amount of bag as well as on the environmental conditions experienced by the bag before being deposited close to seeds.

This manuscript fits the Aims and Scope of the journal, as it examines the impact of discarded plastics (anthroposphere) derived from land-based or marine-based sources (hydrosphere) via leaching by rainwater (atmosphere) on dune plants (biosphere) that play a pivotal role in maintaining and structuring coastal dune systems. Our results are based on experimental ecological data, and their discussion from an ecological management point of view could be useful to formulate new effective environmental policy to limit the impact of plastic pollution on natural habitats.

On behalf of all co-authors,

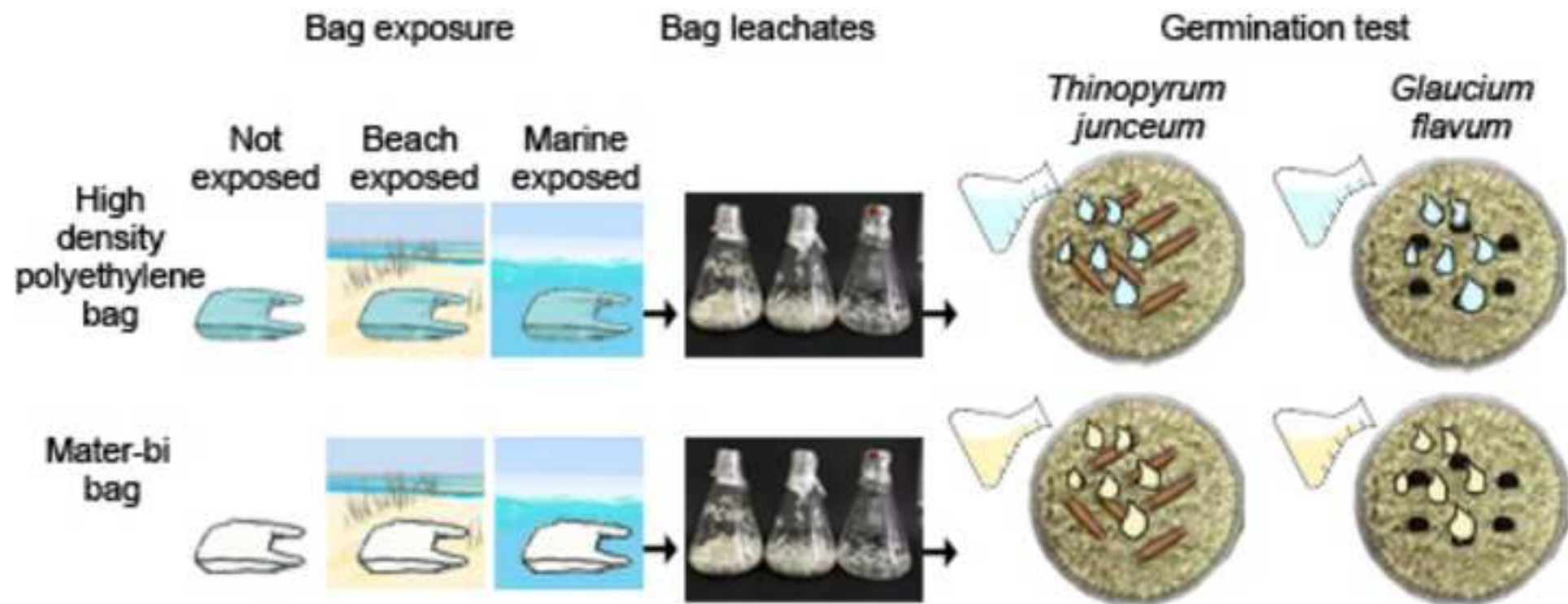
Yours sincerely,

Elena Balestri

Exposure of coastal dune vegetation to plastic bag leachates: a neglected impact of plastic

litter

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65



Highlights.

- Dune systems are hot-spots of accumulation of plastic bags.
- Effects of plastic bag leaching by water on dune plants are largely ignored.
- Leachates from not biodegradable and biodegradable bags alter water quality.
- Leachates differentially influence germination and early growth of two dune plants.
- Leachates can affect plant population and community dynamics of dune habitats.

1 **Exposure of coastal dune vegetation to plastic bag leachates: a neglected impact of plastic**
2 **litter**

3

4 Virginia Menicagli¹, Elena Balestri^{1*}, Claudio Lardicci¹

5

6 ¹Department of Biology, University of Pisa, via Derna 1, 56126, Pisa, Italy

7

8 *Corresponding author

9 elena.balestri@unipi.it

10 University of Pisa, via Derna 1, 56126 Pisa, Italy

11

12

13 Keywords:

14 Compostable bag; *Glaucium flavum*; Polyethylene bag; Seed germination; Seedling growth;

15 *Thinopyrum junceum*

16

17

18

19

20

21

22

23

24

25

26

27 **1. Introduction**

28

29 Coastal dunes consist of a narrow strip of sand located at the interface between the terrestrial and
30 the marine environment, and they result from the dynamic interaction among vegetation, sand and
31 aeolian processes (Fenu et al., 2013; Provoost et al., 2011). Coastal dunes are considered among the
32 most valuable habitats in the world providing key ecological functions and services, such as coastal
33 protection, erosion control and carbon sequestration (Barbier et al., 2011; Martínez et al., 2004).

34 However, they are currently threatened by global climate changes, like sea level rise and increase of
35 high intensity storms and anthropogenic activities, such as excessive resource exploitation and
36 intense coastal development (Balestri and Lardicci, 2013; Barbier et al., 2011; Frosini et al., 2012).
37 Moreover, sand beaches and dunes are hot-spots of plastic accumulation (Ceccarini et al., 2018;
38 Poeta et al., 2014; Poeta et al., 2017; Rangel-Buitrago et al., 2018).

39 Plastic pollution is a global environmental problem, and many studies have dealt with the adverse
40 effects of plastic debris on human health and terrestrial and aquatic organisms (Law, 2017; Moore,
41 2008; Rochman et al., 2015; Talsness et al., 2009; Wilcox et al., 2018). Recent studies have shown
42 that plastic items entering dune systems can physically interact with vegetation (Poeta et al., 2017)
43 and alter sand physical/chemical properties, such as temperature and permeability, and geochemical
44 cycling of elements (Carson et al., 2011). Plastic items can also adsorb chemicals from the natural
45 environment, like persistent organic pollutants, polycyclic aromatic hydrocarbon and metals (Mato
46 et al., 2001; Nakashima et al., 2012; Teuten et al., 2009), toxic for a variety of organisms (Balmer et
47 al., 2015; Minkina et al., 2018; Zhang et al., 2017). When in contact with water, adsorbed chemicals
48 and water-soluble additives included in the polymer matrix during the manufacturing process (Alam
49 et al., 2018; Hermabessiere et al., 2017; Nazareth et al., 2018) can be released from plastics and
50 migrate into the surrounding environment (Nakashima et al., 2012; Teuten et al., 2009). This
51 phenomenon can also occur in dune habitats, especially during rainfall events. Under this
52 circumstance, the water contaminated by plastics can leach into sand and eventually soak seeds and

53 plant roots. Results of a recent study have shown that leachates from plastic bags, both not-
54 biodegradable and biodegradable, affected the development of seedlings of a terrestrial species
55 considered as indicator of phytotoxicity of a variety of chemicals (Balestri et al., 2019). However,
56 no studies have investigated the impact of plastic leachates on dune vegetation. This information is
57 essential for advancing our understanding of the environmental effects of plastics on coastal dune
58 ecosystems.

59 In the present study, we focused on the effects of the leaching of shopper bags by water on the
60 most vulnerable life history stages of dune plants, germination and early seedling growth (Balestri
61 and Cinelli, 2004; Maun 2009; Rajaniemi and Barrett, 2018). Shopper bags made of conventional
62 not biodegradable polymers such as high-density polyethylene (HDPE) are among the most
63 common types of plastic item found along sandy shores and dunes (Alshawafi et al., 2017; Munari
64 et al., 2016; Pasternak et al., 2017; Schmuck et al., 2017; Šilc et al., 2018). Most of these bags
65 derives from land-based sources, such as runoff from rivers and wind-blown litter, or marine-based
66 sources (maritime traffic), but some bags are deliberately or unintentionally discarded directly on
67 beaches by beachgoers (Ryan et al., 2009). Recently, biodegradable and compostable bags have
68 also been found in coastal environments (Balestri et al., 2017). These latter bags are considered as a
69 valid eco-friendly alternative to conventional ones, but they are designed to be disposed in
70 industrial or home compost facilities at the end of their life (EN 13432, 2000). Increasing evidences
71 indicate that once entered natural environments compostable bags degrade slowly and can affect
72 marine organisms and communities (Accinelli et al., 2012; Balestri et al., 2017; Green et al., 2015;
73 Nazareth et al., 2018; Tosin et al., 2012). Predictions suggest that the number of compostable bags
74 entering in the environment will greatly increase and even reach similar levels to that of
75 conventional bags in the future (UNEP, 2015).

76 Here, we examined in laboratory whether (i) conventional and compostable bags would
77 influence via leaching water chemical/physical variables relevant to dune plant development and
78 (ii) the conditions experienced by bags before being deposited close to dune vegetation, i.e.,

79 seawater immersion (for those bags derived from land- and marine- based sources) or sand burial
80 (for those bags deposited on beaches) would influence leachate quality. We then assessed whether
81 bag leachates would differentially affect seed germinability, germination phenology and seedling
82 growth of dune plant species. To this end, we used as models two species belonging to different
83 taxonomic groups that often co-occur on coastal mobile dunes, the monocotyledon *Thinopyrum*
84 *junceum* (L.) Á. Löve, and the dicotyledon *Glaucium flavum* Crantz.

85

86 **2. Material and methods**

87

88 *2.1. Preparation and chemical/physical analysis of bag leachates*

89

90 This experiment tested whether plastic shopper bags would affect water quality via leaching and
91 natural weathering (beach or marine exposure condition) would influence leachate quality. Two
92 types of bags, a high-density polyethylene (PE) bag and a biodegradable and compostable bag made
93 of Mater-bi[®] (MB), purchased from Italian retailers were used to produce leachates. Mater-bi[®] is
94 constituted by starch and vinyl-alcohol copolymers (Sforzini et al., 2016) and certified for industrial
95 composting (EN13432) and “home” composting scheme (OK Compost Home). This certification
96 means that bags are capable to completely degrade in six months in industrial facilities and to be
97 converted in compost devoid of any phytotoxic effects (EN 13432, 2000). All the bags employed in
98 this study belong to the same lot of plastic bags used in a previous study on the effects of bag
99 leachate on the bioindicator species *Lepidium sativum* L. (Balestri et al., 2019).

100 Before the leaching experiment (March 2018), a group of bags (hereafter referred to not-exposed
101 bags, NE) was left in laboratory at a temperature of 22 °C (\pm 1 SE) to test the effects of bag material
102 itself on chemical/physical properties of the water. Other two groups of bags were exposed outdoor
103 over a period of 10 days for weathering (Balestri et al., 2019). The first bag group (hereafter
104 referred to as beach-exposed bags, BE) was placed in an upper beach area (average daily

105 temperature of 7.1 ± 1.6 °C, total amount of precipitations of 60.8 mm and mean daylength of 11
106 hours) at Rosignano Solvay (43°23'N 10°26' E, Italy). Bags were carefully laid out on the sandy
107 substrate and fixed with pebbles to avoid dispersion. The second group of bags (hereafter referred
108 to as marine-exposed bags, ME) was immersed in seawater in a shallow (0.5 m depth) coastal zone
109 at Rosignano Solvay. Each bag was secured with cotton wires to a sample holder anchored to the
110 bottom and left to move freely in the water column (average seawater temperature of 13.7 ± 0.1
111 °C). At the end of the exposure period, the bags were collected and transported to the laboratory for
112 the leachate preparation.

113 Each plastic bag was cut into pieces of approximately 1 cm^2 . For each type of bag, different
114 amounts of pieces were placed into clean glass flasks to obtain liquid-to-solid (water-to-plastic)
115 ratios of 100, 10 and 5, corresponding respectively to approximately 8.3×10^{-4} , 8.3×10^{-3} and $1.6 \times$
116 10^{-2} bag/mL, hereafter referred to as low (L), intermediate (I) and high (H) pollution degree. These
117 ratios were chosen to mimic various degrees of bag pollution occurring in natural sandy shores
118 (Alshawafi et al., 2017; Munari et al., 2016; Pasternak et al., 2017; Schmuck et al., 2017). To obtain
119 bag leachates, sterilized deionized water with a pH value (6.40 ± 0.03) similar to that of the
120 rainwater of the coastal Mediterranean basin (Loye-Pilot et al., 1986) was used. For each bag type,
121 deionized water with no plastic material was used as control solution (no pollution). Flasks were
122 placed in a culture chamber in darkness on an orbital shaker (95 rotations per minute) for 72 h
123 (Bejgarn et al., 2015) at 24 ± 1 °C. The experiment followed a full factorial design with three
124 factors, plastic (fixed, two levels: PE and MB), exposure (fixed, three levels: not-exposed bags,
125 beach-exposed bags and marine-exposed bags) and pollution (fixed, four levels: control or no
126 pollution, low, intermediate and high). There were three replicates per each treatment combination.

127 After the incubation period, plastic fragments in each flask were removed from the liquid phase
128 by filtration using a nylon mesh (200 μm). To investigate whether the leaching from plastic bags
129 altered the quality of water, a sample (30 mL) of filtered leachate obtained from each MB and PE
130 flask was collected, and water chemical/physical variables (pH, oxidation-reduction potential, total

131 dissolved solids and salinity) relevant to plant development (Husson, 2013; Isermann, 2005; Sykes
132 and Wilson, 1989) were measured by a multiparameter meter (HI98194, Hanna Instruments). The
133 remaining filtered leachate of each flask was used for the seed germination and seedling growth
134 test.

135

136 2.2. Seed germination and seedling growth test

137

138 This experiment evaluated the potential effect of bag leachates on the seed germination process
139 and early growth of *Thinopyrum junceum* and *Glaucium flavum* (yellow horned-poppy). Seeds were
140 harvested from plants inhabiting Rosignano Solvay dune system in July 2017 and separately stored
141 outdoor in clean glass jars in darkness until the setup of the germination test. In March 2018, seeds
142 were visually examined under a stereomicroscope (Wild M3C, Leica) and subjected to pressure test
143 with forceps. This test allowed us to distinguish between non-viable seeds, as those that collapse
144 under gentle pressure, and viable seeds as those that remain firm after the pressure (Borza et al.,
145 2007). Non-viable seeds were discarded.

146 Viable seeds were rinsed with sterile deionized water and sown in 9-cm Petri dishes (6 seeds per
147 dish) containing sterilized natural sand (0.5 - 1 mm, < 0.01% organic matter content) previously
148 moistened with 3 mL of one of the filtered leachates. Each dish was sealed with parafilm to prevent
149 desiccation, and the dishes were randomly placed in a culture chamber in darkness at 15 ± 1 °C to
150 simulate favorable natural condition for germination (Sykes and Wilson, 1989; Thanos et al., 1989).
151 During the incubation period, seeds were daily checked by using low green light intensity to record
152 the number of germinated seeds in each dish. A seed was considered to have germinated when the
153 radicle length had reached at least 2 mm (Balestri and Cinelli, 2004; Luo et al., 2017). A visual
154 evaluation of seedling developmental abnormalities (Chandler, 2008; ISTA, 2003) was also carried
155 out. Germinated seeds were removed from each dish five days after their germination and stored in
156 70% ethanol for morphological measurements. The test was considered finished when no additional

157 seeds germinated for at least three consecutive weeks. The experiment followed a full factorial
158 design with four factors, plant species (fixed, two levels: *T. junceum* and *G. flavum*), plastic (fixed,
159 two levels: PE and MB), exposure (fixed, three levels: not-exposed bags, beach-exposed bags and
160 marine-exposed bags) and pollution (fixed, four levels: control or no pollution, low, intermediate
161 and high). Four replicates were used for each treatment combination.

162 At the end of the experiment, for each dish the percentage of seed germination and the mean
163 germination time, i.e., the time in days elapsed between seed sowing and germination, was
164 calculated as

$$165 \quad t = \frac{\sum_{i=1}^k n_i t_i}{\sum_{i=1}^k n_i},$$

166 where t_i was the time elapsed from the start of the experiment to the i^{th} day of observation; n_i was
167 the number of germinated seeds in the day i , and k was the day in which the last germination event
168 was observed (Ranal and De Santana, 2006). The percentage of abnormal seedlings was also
169 calculated for each dish. For each treatment, a sample of the normal seedlings ($n = 6$) stored in
170 ethanol was randomly chosen and each seedling was carefully placed on squared paper and
171 photographed. The length of the radicle and the length of aboveground organ (hypocotyl for *G.*
172 *flavum* and coleoptile for *T. junceum*) of each seedling was measured with an image analysis
173 software (ImageJ 2, Rueden et al., 2017). The below- to aboveground organ length ratio for each
174 seedling was also computed as considered as a valuable indicator of relative resource allocation
175 (Yang et al., 2018).

176

177 2.3. Data analysis

178

179 Non-metric multidimensional scaling (MDS) based on the Euclidean distance was separately
180 conducted for each bag type on chemical/physical variables of leachates (pH, oxidation-reduction
181 potential, total dissolved solids and salinity) to visualize differences among samples of each
182 treatment (exposure and pollution). Three-way permutational multivariate analysis of variance

183 (PERMANOVA) was performed on chemical/physical data to examine the overall effect of
184 treatments (plastic, exposure and pollution), followed by univariate three-way PERMANOVAs to
185 test for differences between treatments for each individual variable.

186 Four-way multivariate PERMANOVAs were separately conducted on $\log(x+1)$ transformed
187 germination data (percentage of germination and mean germination time) and on seedling growth
188 data (radicle length, aboveground organ length and below- to aboveground organ length ratio) to
189 investigate the effects of the different type of plastic bag, exposures and pollution on the
190 performance of *T. junceum* and *G. flavum*. Then, separate analysis of variance (ANOVA) were
191 conducted on each variable and on the percentage of abnormal seedlings. A Student-Newman-Keuls
192 (SNK) test was applied to perform post hoc comparisons among levels of significant terms.

193 PERMANOVA analyses were carried out on the Euclidean distance of previously normalized
194 data using 9999 permutations of the residuals under a reduced model, and when significant effects
195 were detected posteriori pair-wise comparisons using 9999 random permutations were conducted.
196 In posteriori pairwise comparisons of chemical/physical variables there were not enough
197 permutable units to get a reasonable test by permutation for some terms, thus p-values were
198 obtained using a Monte Carlo random sample from the asymptotic permutation distribution
199 (Anderson et al., 2008). Statistically significant terms ($p < 0.05$) were checked for differences in
200 multivariate group dispersion through permutational analysis of multivariate dispersion
201 (PERMDISP). Prior to ANOVA analyses, data were checked for normality and homogeneity of
202 variance with Shapiro-Wilk test and Cochran's *C* test, respectively. Data of mean germination time
203 ($\sqrt{x+1}$), radicle length ($\ln x$) and below- to aboveground length ratio ($\ln(x+1)$) were
204 transformed to meet ANOVA assumptions. Since the transformations applied to the percentage of
205 germination and the percentage of abnormalities failed to remove the heterogeneity of variances,
206 untransformed data were analyzed, and the results were considered robust if not significant (at
207 $p > 0.05$) or significant at $p < 0.01$ to compensate for increased probability of type I error
208 (Underwood, 1997). PERMANOVA analyses were conducted using PERMANOVA + for PRIMER

209 6 statistical software (Anderson et al., 2008; Clarke and Gorley, 2006) while ANOVA analyses
210 were performed using GMAV version 5.0 for Windows (Underwood and Chapman, 1998).

211

212 **3. Results**

213

214 *3.1. Chemical/physical analysis of leachates*

215

216 Non-metric multidimensional scaling plots relative to chemical/physical variables of leachates
217 obtained from PE and MB bags showed that control samples (no pollution) were largely
218 overlapped, and they substantially differed from samples belonging to the plastic pollution
219 treatments (Fig. 1). Significant effects of all the main factors and of their interactions were detected
220 by multivariate and univariate PERMANOVA (Table 1). The pH of control solutions (no pollution)
221 was significantly lower than that of PE leachates irrespectively of the type of exposure, and it was
222 also lower than that of MB leachates from marine-exposed bags. Instead, the pH of controls was
223 significantly higher than that of leachates from not-exposed and beach-exposed MB bags (Fig. 2
224 and Table 2). The impact of each type of bag on pH leachates increased with increasing pollution
225 (Fig. 2 and Table 2). For the pH of leachates obtained from the intermediate and the high amount of
226 MB bags, as well as from the low and the intermediate amount of PE bags, there were significant
227 differences among exposure conditions, and the lowest pH values were measured in the leachates
228 from not-exposed bags (Fig. 2 and Table 2). The oxidation-reduction potential (ORP) of PE and
229 MB bag leachates was lower compared to that of the control, except that for those from not-exposed
230 MB bags, and it decreased with increasing pollution irrespectively of bag type (Fig. 2 and Table 2).
231 A significant greater amount of total dissolved solids (TDS) was measured in plastic leachates
232 compared to controls (Fig. 2 and Table 2). Except that for leachates from not-exposed MB bags, the
233 amount of TDS significantly increased with increasing pollution, regardless of the type of bag and
234 exposure (Fig. 2 and Table 2). The salinity of control solutions was similar to that of leachate from

235 not-exposed MB bags but significantly lower compared to that of all the other leachates produced
236 with the intermediate and the high amount of bag material (Fig. 2 and Table 2). Significant higher
237 values of TDS and salinity were measured in leachates from marine-exposed bag than that from
238 not-exposed and beach-exposed bags, irrespectively of plastic type and pollution level. For the
239 leachates obtained from the intermediate and the high amount of bag material, there were
240 significant differences in TDS and salinity values among exposure conditions, and the lower values
241 were measured in the leachates from not-exposed bags (Fig. 2 and Table 2).

242

243 3.2. Seed germination and seeding growth test

244

245 Results of PERMANOVA on germination variables and of ANOVA on germination time
246 revealed a significant effect of the main factors, species and pollution, and of the interaction among
247 species, plastic, exposure and pollution (Tables 3 and 4). Seeds of *T. junceum* treated with leachates
248 from the high amount of marine-exposed MB bags geminated later than control seeds (Figs. 3 and 4
249 and Table 5). Instead, seeds of *G. flavum* treated with bag leachates germinated earlier than control
250 seeds irrespectively of bag type and exposure conditions, except that those soaked with the highest
251 amount of not-exposed MB bags (Figs. 3 and 4 and Table 5). Results of ANOVAs also detected a
252 significant effect of the interactions between the factors, species and exposure, and exposure and
253 pollution, on the germination percentage (Table 4). A higher seed germination percentage was
254 observed for *T. junceum* compared to *G. flavum* (Table 5). For both species, seeds treated with
255 leachates from the low concentration of marine-exposed bags showed a higher germination
256 percentage compared to control seeds regardless of plastic type (Fig. 3 and Table 5).

257 Some seedlings failed to produce the hypocotyl (for *G. flavum*) or the coleoptile (for *T.*
258 *junceum*), and the percentage of these abnormalities was significantly higher in seedlings grown
259 with leachates from the high amount of bag material, irrespectively of plastic type and exposure
260 (Figs. 3 and 5 and Table 5). Multivariate PERMANOVA analysis on seedling growth variables

261 detected a significant effect of the main factors, species, plastic and pollution, and of the interaction
262 among all factors (Table 3). ANOVA analysis on radicle length revealed a significant effect of
263 factors, species and pollution, and of the interaction among all the main factors (Table 6).
264 *Thinopyrum junceum* seedlings grown with the leachate from the high amount of not-exposed MB
265 bags showed a significant shorter radicle compared to control seedlings (Fig. 6 and Table 7). A
266 shorter radicle length was also found in *G. flavum* seedlings grown with the leachate from the low
267 amount of beach-exposed PE bags compared to control seedlings (Fig. 6 and Table 7). ANOVA
268 performed on the length of aboveground organs detected a significant effect of all the main factors
269 and of their interaction (Table 6). Seedlings of *G. flavum* grown with the leachate from the low
270 amount of not-exposed PE bag had a significant longer hypocotyl compared to those belonging to
271 control (Fig. 6 and Table 7). For below- to aboveground organ length ratio, a significant effect of
272 the factors, species and pollution, and of the interaction among all the main factors was detected
273 (Table 6). *Glaucium flavum* seedlings grown with the leachate from the intermediate amount of
274 beach-exposed PE bags or with that from the high amount of marine-exposed PE bags showed a
275 significantly higher radicle to hypocotyl ratio than that of control seedlings as a result of relatively
276 greater radicle elongation (Fig. 6 and Table 7). A significantly higher radicle to hypocotyl ratio was
277 also found for *G. flavum* seedlings grown with the intermediate or high amount of not-exposed MB
278 leachates compared to control seedlings, but this was mainly due to relatively lower hypocotyl
279 elongation (Fig. 6 and Table 7).

280

281 **4. Discussion**

282

283 Coastal dune vegetation establishment and development are strongly influenced by the
284 availability and amount of rainwater (Maun, 2009). Results of our experiment simulating the
285 leaching of conventional and compostable bags by rainwater on sand dunes demonstrate that both
286 types of bag altered water chemical/physical characteristics, and the magnitude of these alterations

287 varied with bag concentration and the environmental conditions experienced by bags. Generally, PE
288 bags enhanced water pH while MB bags, except that those immersed in seawater, reduced it,
289 confirming results of a previous studies on the characteristics of the leachates obtained with the
290 same lots of plastic bags (Balestri et al., 2019). The basic pH of PE leachates could be related to the
291 release of linear long-chain alkanes and alkenes from bags, while the acid pH of MB leachates
292 could be due to the release of free butane 1,4-diol from bags and other compounds (Balestri et al.,
293 2019) employed with corn starch for the preparation of Mater-bi[®] (Canellas et al., 2015). Instead,
294 the slightly basic and neutral pH values of leachates obtained respectively from marine- and beach-
295 exposed MB bags could be a result of the buffering effect of salts deposited on bags during
296 weathering. The ORP values of the leachates from not-exposed PE and MB bags could be related to
297 the changes of pH induced by these bags (Liu et al., 2009), while those of the leachates from
298 exposed bags could be mainly a result of biological activities of microorganisms grown on bags
299 during weathering. Lastly, the greater amount of total dissolved solids and the higher salinity in
300 leachates from all exposed PE and MB bags compared to not-exposed ones could depend on the
301 embrittlement of bags and the deposition of salts due to weathering. In nature, alterations of
302 chemical/physical characteristics of soil water can have considerable effects on plant development.
303 Indeed, the pH and the ORP values of water in sandy soils can influence the mobility and the
304 availability of nutrients and carbonate, while soil salinity is the major selective force in seed
305 germination and seedling growth of dune plants (Husson, 2013; Maun, 2009; Sykes and Wilson,
306 1989).

307 Results of our seed germination experiment indicate that both the types of bag affected via
308 leaching seed germination processes and seedling growth. As observed for water chemical/physical
309 variables, the magnitude and direction of bag leachate effects on the germination phenology varied
310 between the species and depended on type and amount of bag, as well as on the environmental
311 conditions experienced by the bag before being deposited close to seeds. In general, bag leachates
312 advanced the timing of seed germination in *G. flavum*. This was likely due to the migration of

313 chemical compounds from bags into the water phase (Balestri et al., 2019) and to the variations of
314 water chemical/physical characteristic that are known to interact with factors controlling
315 germination and dormancy release (El-Maarouf-Bouteau and Bailly, 2008). Instead, the bag
316 leachate produced from the high amount of marine-exposed MB bags delayed the timing of seed
317 germination in *T. junceum*. This could be related to the release of high amounts of salts in water, as
318 previous studies have shown an increase of mean germination timing of seeds this species exposed
319 to high salt concentrations (El-Katony et al., 2015). On the other hand, the germinability of seeds of
320 *G. flavum* and *T. junceum* was positively affected by leachates from low amount of marine-exposed
321 PE and MB bags. This stimulatory effect was possibly due to the release in water of low amount of
322 salts deposited on the bags during seawater immersion. In fact, moderate salt concentrations have
323 been shown to increase seed germination percentage of some dune species (Ungar, 1978; Woodell,
324 1983).

325 As concerning seedling growth, *T. junceum* and *G. flavum* responded differentially to leachates
326 from PE and MB bags. Indeed, *T. junceum* seedlings were affected only by the leachate produced
327 from the high concentration of not-exposed MB bags, and the radicle was the most sensitive organ.
328 Instead, *G. flavum* seedlings were influenced by leachates from PE bags, showing a relatively
329 greater elongation of the radicle than hypocotyl with leachates from beach- and marine-exposed
330 bags and a relative increase of hypocotyl length with the leachate from the low concentration of
331 not-exposed bags compared to controls. These alterations could be due to the basic water pH values,
332 that are known to promote *G. flavum* growth (Scott 1963), and to the biological activity of BPA
333 present in the water phase (Balestri et al., 2019). Indeed, this additive at low concentration has been
334 shown to promote plant growth (Li et al., 2018; Pan et al., 2013; Staples et al., 2010). Seedlings of
335 *G. flavum* were also sensitive to the leachate from the high concentration not-exposed MB bags,
336 showing a lower investment in hypocotyl growth relative to radicle compared to controls. This
337 inhibitory effect could be attributed to the activity of chemical compounds migrated from bags as
338 well as to water acid pH values that are known to adversely affect plant growth (Turner et al.,

339 1988). However, regardless of bag type and exposure a consistent number of seedlings of both the
340 species grown with leachates from the high concentration of bags exhibited a developmental
341 abnormality. The occurrence of seedling abnormalities in sensitive plants is an indicator of
342 phytotoxicity of a certain substance (Chandler, 2008; De Barro, 2008; ISTA, 2003; Mitchell et al.,
343 1988). Here, the observed abnormality could be related to the presence in the water phase of plastic
344 additives and non-intentionally added compounds, such as for example bisphenol A (BPA) and 1,6-
345 dioxacyclododecane-7,12-dione, that are toxic to a variety of organisms and terrestrial plants
346 (Balestri et al., 2019; Jyothi et al., 2014; Kennedy, 2002).

347 In dune environments, shifts in germination phenology and alterations in biomass allocation to
348 above-belowground organs can have a considerable impact on plant population dynamics and
349 structure of communities. For example, an advance or a delay of seed germination timing of a
350 species could lead to an overlap of its germination spread (i.e. the time elapsed between the first and
351 the last seed germination event) with that of a neighbor species and hence increase the intensity of
352 interspecific interactions (Maun, 2009). On the other hand, a reduction of the germination spread of
353 a species could increase the intensity of intraspecific interactions among co-specific seedlings. In
354 addition, the production of short radicles could not guarantee an adequate uptake of nutrients and
355 water to seedlings while short hypocotyls could make seedlings more vulnerable to sand burial
356 (Balestri et al., 2012; Maun, 2009).

357 On the basis of the findings of the present study, different plant population establishment
358 scenarios could be hypothesized. For example, in sand dune areas not impacted by plastic bags,
359 seeds of *T. junceum* are expected to germinate earlier than those of *G. flavum*, and the germination
360 spreads of these species would never overlap. In areas polluted by PE bags, irrespectively of their
361 exposure, seeds of *G. flavum* buried close to bags would germinate earlier and their germination
362 spread would be reduced as compared to that of seeds located far away from bags (Fig. 4a,b,c).
363 Under this scenario, the chance of interspecific competition among *G. flavum* seedlings near to the
364 bags would increase, and emerging seedlings would morphologically differ from those grown in

365 non-polluted areas. In areas polluted by MB bags, directly deposited or aged on beaches, the
366 germination behavior *G. flavum* would be similar to that described in the above scenario, but the
367 germination spread of this species would overlap to that of *T. junceum* (Fig. 4d,e). Under this
368 scenario, *G. flavum* seedlings would greatly differ in developmental stage and the chance of both
369 intra- and interspecific competition among seedlings would be enhanced. In addition, many *G.*
370 *flavum* seedlings emerging close to the bags directly abandoned in dunes would differ in the pattern
371 of allocation of biomass from those emerged in not polluted areas. Lastly, in dune areas highly
372 polluted by MB bags transported by the sea, *T. junceum* seeds buried close to bags would germinate
373 later than those located far away from bags (Fig. 4f), and this would lead to a seedling population
374 more heterogeneous in size. Delayed germination timing would also cause an overlap of the seed
375 germination spreads of *T. junceum* and *G. flavum* and increase the chance of interspecific
376 competition among seedlings (Fig. 4f). Irrespectively of the type of scenario, in highly polluted
377 dune areas a significant number of germinated seeds of both study species would not emerge and
378 survive lacking of the above-ground organ.

379

380 **5. Conclusion**

381

382 The present study provides the first experimental evidence of the impact of plastic litter, and
383 specifically of shopper bags, on dune plant establishment and early development. Our findings
384 demonstrated that both conventional and compostable bags can interact with abiotic/biotic factors
385 before entering vegetated dunes, and their leachates can interfere in complex ways with
386 mechanisms that regulate germination, dormancy release and early growth. The presence of both
387 types of bag on mobile dunes could locally affect sexual recruitment and the intensity of intra- and
388 interspecific seedling interactions, and hence could have a consistent impact on dune community
389 dynamics and structure. Overall, these findings indicate that the leaching of plastic bags should be
390 considered as a further threat to coastal environments and associated ecosystems. Importantly, they

391 suggest that more efforts should be paid in the future to avoid or limit the accidental or intentional
392 dispersion of biodegradable and compostable bags in natural environments.

393

394 **Acknowledgements**

395 We sincerely thank Flavia Vallerini, Viviana Ligorini and Andrea Torre for their support in the
396 laboratory experiment. This work is part of the PhD research project of Virginia Menicagli funded
397 by the University of Pisa.

398

399 **References**

400

401 Accinelli, C., Saccà, M.L., Mencarelli, M., Vicari, A., 2012. Deterioration of bioplastic carrier bags
402 in the environment and assessment of a new recycling alternative. *Chemosphere*. 89, 136-143.

403 <https://doi.org/10.1016/j.chemosphere.2012.05.028>.

404 Alam, O., Billah, M., Yajie, D., 2018. Characteristics of plastic bags and their potential
405 environmental hazards. *Resour Conserv Recycl*. 132, 121-129.

406 <https://doi.org/10.1016/j.resconrec.2018.01.037>

407 Alshawafi, A., Analla, M., Alwashali, E., Aksissou, M., 2017. Assessment of marine debris on the
408 coastal wetland of Martil in the North-East of Morocco. *Mar Pollut Bull*. 117, 302-310.

409 <https://doi.org/10.1016/j.marpolbul.2017.01.079>

410 Anderson, M.J., Gorley, R.N., Clarke, K.R., 2008. PERMANOVA+ for PRIMER: Guide to
411 software and statistical methods. PRIMER-E: Plymouth, UK.

412 Balestri, E., Cinelli, F., 2004. Germination and early-seedling establishment capacity of *Panocratium*
413 *maritimum* L. (Amaryllidaceae) on coastal dunes in the north-western Mediterranean. *J Coast*

414 *Res*. 20, 761-770. [https://doi.org/10.2112/1551-5036\(2004\)20\[761:GAEECO\]2.0.CO;2](https://doi.org/10.2112/1551-5036(2004)20[761:GAEECO]2.0.CO;2)

415 Balestri, E., Lardicci, C., 2013. The impact of physical disturbance and increased sand burial on
416 clonal growth and spatial colonization of *Sporobolus virginicus* in a coastal dune system. PLoS
417 ONE. 8(8): e72598. <https://doi:10.1371/journal.pone.0072598>

418 Balestri, E., Menicagli, V., Vallerini, F., Lardicci C., 2017. Biodegradable plastic bags on the
419 seafloor: A future threat for seagrass meadows? Sci Total Environ. 605-606, 755-763.
420 <https://doi.org/10.1016/j.scitotenv.2017.06.249>

421 Balestri, E., Vallerini, F., Castelli, A., Lardicci, C., 2012. Application of plant growth regulators, a
422 simple technique for improving the establishment success of plant cuttings in coastal dune
423 restoration. Estuar Coast Shelf Sci. 99, 74-84. <https://doi.org/10.1016/j.ecss.2011.12.017>

424 Balestri, E., Menicagli, V., Ligorini, V., Fulignati, S., Raspolli Galletti, A.M., Lardicci, C., 2019.
425 Phytotoxicity assessment of conventional and biodegradable plastic bags using seed
426 germination test. Ecol Indic. 102, 569-580. <https://doi.org/10.1016/j.ecolind.2019.03.005>

427 Balmer, B.C., Ylitalo, G.M., McGeorge, L.E., Baugh, K.A., Boyd, D., Mullin, K.D., Rosel, P.E.,
428 Sinclair, Wells, R.S., Zolman, E.S., Schwacke, L.H., 2015. Persistent organic pollutants (POPs)
429 in blubber of common bottlenose dolphins (*Tursiops truncatus*) along the northern Gulf of
430 Mexico coast, USA. Sci Tot Environ. 527-528, 306-312.
431 <http://dx.doi.org/10.1016/j.scitotenv.2015.05.016>

432 Barbier, E. B., Hacker, S. D., Kennedy, C., Koch, E. W., Stier, A. C., Silliman, B., 2011. The value
433 of estuarine and coastal ecosystem services. Ecol Monogr. 81, 169-193.
434 <https://doi.org/10.1890/10-1510.1>

435 Bejgarn, S., MacLeod, M., Bogdal, C., Breitholtz, M., 2015. Toxicity of leachate from weathering
436 plastics: an exploratory screening study with *Nitocra spinipes*. Chemosphere. 132, 114-119.
437 <https://doi.org/10.1016/j.chemosphere.2015.03.010>

438 Borza, J.K., Westerman, P.R., Liebman, M., 2007. Comparing estimates of seed viability in three
439 foxtail (*Setaria*) species using the imbibed seed crush test with and without additional
440 tetrazolium testing. Weed Technology. 21, 518-522. <http://dx.doi.org/10.1614/WT-06-110>

441 Canellas, E., Vera, P., Nerin, C., 2015. UPLC-ESI-Q-TOF-MS(E) and GC-MS identification and
442 quantification of non-intentionally added substances coming from biodegradable food
443 packaging. *Anal. Bioanal. Chem.* 407, 6781–6790. <https://doi.org/10.1007/s00216-015-8848-2>

444 Carson, H.S, Colbert, S.L., Kaylor, M., McDermid, K.J., 2011. Small plastic debris changes water
445 movement and heat transfer through beach sediments. *Mar Pollut Bull.* 62, 1708-1713.
446 <https://doi.org/10.1016/j.marpolbul.2011.05.032>

447 Ceccarini, A., Corti, A., Erba, F., Modugno, F., La Nasa, J., Bianchi, S., Castelvetro, V., 2018. The
448 hidden microplastics: new insights and figures from the thorough separation and
449 characterization of microplastics and of their degradation by-products in coastal sediments.
450 *Environ Sci Technol.* 52, 5634-5643. <https://doi.org/10.1021/acs.est.8b01487>

451 Chandler, J.W., 2008. Cotyledon organogenesis. *J Exp Bot.* 59, 2917-2931.
452 <https://doi.org/10.1093/jxb/ern167>

453 Clarke, K.R., Gorley, R.N., 2006. *PRIMER v6: User manual/tutorial*. PRIMER-E: Plymouth.

454 De Barro, J., 2008. Understanding and managing the causes of abnormal seedlings in Lucerne.
455 Rural Industries Research and Development Corporation.

456 El-Maarouf-Bouteau, H., Bailly, C., 2008. Oxidative signaling in seed germination and dormancy.
457 *Plant Signal Behav.* 3, 175–182.

458 El-Katony, T. M., Khedr, A. A., Soliman, N. G., 2015. Nutrients alleviate the deleterious effect of
459 salinity on germination and early seedling growth of the psammophytic grass *Elymus farctus*.
460 *Botany.* 93, 559-571. <https://doi.org/10.1139/cjb-2015-0096>

461 EN 13432, 2000. Packaging - Requirements for packaging recoverable through composting and
462 biodegradation - Test scheme and evaluation criteria for the final acceptance of packaging.

463 Fenu, G., Carboni, M., Acosta, A.T.R., Baccetta, G., 2013. Environmental factors influencing
464 coastal vegetation pattern: new insights from the mediterranean basin. *Folia Geobot.* 48: 493-
465 508. <https://doi.org/10.1007/s12224-012-9141-1>

466 Frosini, S., Lardicci, C., Balestri, E., 2012. Global change and response of coastal dune plants to the
467 combined effects of increased sand accretion (burial) and nutrient availability. PLoS ONE. 7:
468 e47561. <https://doi.org/10.1371/journal.pone.0047561>

469 Green, D.S., Boots, B., Blockley, D.J., Rocha, C., Thompson, R.C., 2015. Impacts of discarded
470 plastic bags on marine assemblages and ecosystem functioning. Environ Sci Technol. 49, 5380-
471 5389. <https://doi.org/10.1021/acs.est.5b00277>.

472 Hermabessiere, L., Dehaut, A., Paul-Pont, I., Lacroix, C., Jezequel, R., Soudant, P., Duflos, G.,
473 2017. Occurrence and effects of plastic additives on marine environments and organisms: a
474 review. Chemosphere. 182, 781-793. <https://doi.org/10.1016/j.chemosphere.2017.05.096>

475 Husson, O., 2013. Redox potential (Eh) and pH as drivers of soil/plant/microorganism systems: a
476 transdisciplinary overview pointing to integrative opportunities for agronomy. Plant Soil. 362,
477 389–417. <https://doi.org/10.1007/s11104-012-1429-7>

478 Isermann, M., 2005. Soil pH and species diversity in coastal dunes. Plant Ecol. 178, 111–120.
479 <https://doi.org/10.1007/s11258-004-2558-8>

480 ISTA 2003. International rules for seed testing. Edition 2003, International seed testing association,
481 Zurich, Switzerland.

482 Jyothi, G., Reddy, K.R.K., Reddy, K.R.N., 2014. Oleic acid: a mycoherbicide from *Cochliobolus*
483 *lunatus* for management of *Echinochloa crusgalli* in paddy. JEBAS. 2, 405-409.

484 Kennedy, G.L. Jr., 2002. Toxicity of adipic acid. Drug Chem Toxicol. 25, 191-202.

485 Law, K.L., 2017. Plastics in the marine environment. Annu Rev Mar Sci. 9, 205–29.
486 <https://doi.org/10.1146/annurev-marine-010816-060409>

487 Li, X., Wang, L., Wang, S., Yang, Q., Zhou, Q., Huang, X., 2018. A preliminary analysis of the
488 effects of bisphenol A on the plant root growth via changes in endogenous plant hormones.
489 Ecotoxicol Environ Saf. 150, 152-158. <https://doi.org/10.1016/j.ecoenv.2017.12.031>

490 Liu, X., Wang, J., Zhang, D., Li, Y., 2009. Grey relational analysis on the relation between marine
491 environmental factors and oxidation-reduction potential. *Chinese J Oceanol Limnol.* 27, 583-
492 586, <https://doi.org/10.1007/s00343-009-9152-9>

493 Loye-Pilot, M.D., Martin, J.M., Morelli, J., 1986. Influence of Saharan dust on the rain acidity and
494 atmospheric input to the Mediterranean. *Nature.* 321, 427- 428.

495 Luo, Y., Liang, J., Zeng, G., Chen, M., Mo, D., Li, G., Zhang, D., 2018. Seed germination test for
496 toxicity evaluation of compost: Its roles, problems and prospects. *Waste Manag.* 71, 109-114.
497 <https://doi.org/10.1016/j.wasman.2017.09.023>

498 Martinez M.L., Psuty, N.P., Lubke, R.A., 2004. A Perspective on Coastal Dunes, in: Martínez,
499 M.L., Psuty, N.P. (Eds.), *Coastal dune - ecology and conservation.* Springer, pp. 3-10.

500 Mato, Y., Isobe, T., Takada, H., Kanehiro, H., Ohtake, C., Kaminuma, T., 2001. Plastic resin pellets
501 as a transport medium for toxic chemicals in the marine environment. *Environ Sci Technol.* 35,
502 318–324. <https://doi.org/10.1021/es0010498>

503 Maun, M. A., 2009. *The biology of coastal sand dunes.* Oxford University Press, New York.

504 Minkina, T., Fedorenko, g., Nevidomskaya, D., Fedorenko, A., Chaplygin, V., Mandzhieva, S.,
505 2018. Morphological and anatomical changes of *Phragmites australis* Cav. due to
506 the uptake and accumulation of heavy metals from polluted soils. *Sci Tot Environ.* 636, 392–
507 401. <https://doi.org/10.1016/j.scitotenv.2018.04.306>

508 Mitchell, R.L., Burchett, M.D., Pulkownik, A., McCluskey, L., 1988. Effects of environmentally
509 hazardous chemicals on the emergence and early growth of selected Australian plants. *Plant*
510 *Soil.* 112, 195-199. <https://doi.org/10.1007/BF02139995>

511 Moore, C.J., 2008. Synthetic polymers in the marine environment: a rapidly increasing, long-term
512 threat. *Environ Res.* 108, 131-9. <https://doi.org/10.1016/j.envres.2008.07.025>

513 Munari, C., Corbau, C., Simeoni, U., Mistri, M., 2016. Marine litter on Mediterranean shores:
514 analysis of composition, spatial distribution and sources in north-western Adriatic beaches.
515 *Waste Manag.* 49, 483-490. <http://dx.doi.org/10.1016/j.wasman.2015.12.010>

516 Nakashima, E., Isobe, A., Kako, S., Itai, T., Takahashi, S., 2012. Quantification of toxic metals
517 derived from macroplastic litter on Ookushi beach, Japan. *Environ Sci Technol.* 46, 10099-
518 10105. <https://doi.org/10.1021/es301362g>

519 Nazareth, M., Marques, M. R. C., Leite, M. C. A., Braga Castro, I., 2019. Commercial plastics
520 claiming biodegradable status: Is this also accurate for marine environment? *J Hazard Mater.*
521 366, 714-722. <https://doi.org/10.1016/j.jhazmat.2018.12.052>

522 Pan, W.J., Xiong, C., Wu, Q.P., Liu, J.X., Liao, H.M., Chen, W., Liu, Y.S., Zheng, L., 2013. Effect
523 of BPA on the germination, root development, seedling growth and leaf differentiation under
524 different light conditions in *Arabidopsis thaliana*. *Chemosphere.* 93, 2585-2592.
525 <https://doi.org/10.1016/j.chemosphere.2013.09.081>

526 Pasternak, G., Zviely, D., Ribic, C.A., Ariel, A., Spanier, E., 2017. Sources, composition and spatial
527 distribution of marine debris along the Mediterranean coast of Israel. *Mar Pollut Bull.* 114,
528 1036-1045. <https://doi.org/10.1016/j.marpolbul.2016.11.023>

529 Poeta, G., Battisti, C., Acosta, A. T. R., 2014. Marine litter in Mediterranean sandy littoral: spatial
530 distribution patterns along central Italy coastal dunes. *Mar Pollut Bull.* 89, 168-173.
531 <http://dx.doi.org/10.1016/j.marpolbul.2014.10.011>

532 Poeta, G., Fanelli, G., Pietrelli, L., Acosta, A. T. R., Battisti, C., 2017. Plasticsphere in action:
533 evidence for an interaction between expanded polystyrene and dunal plants. *Environ Sci Pollut*
534 *Res.* 24, 11856-11859. <https://doi.org/10.1007/s11356-017-8887-7>

535 Provoost, S., Laurence, M., Jones, M., Edmondson, S. E., 2011. Changes in landscape and
536 vegetation of coastal dunes in northwest Europe: a review. *J Coast Conserv.* 15, 207-226.
537 <https://doi.org/10.1007/s11852-009-0068-5>

538 Rajaniemi, T. K., Barrett, D. T., 2018. Germination responses to abiotic stress shape species
539 distributions on coastal dunes. *Plant Ecol.* 219, 1271-1282. [https://doi.org/10.1007/s11258-018-](https://doi.org/10.1007/s11258-018-0877-4(0123456789().,-volV()0123456789().,-volV))
540 [0877-4\(0123456789\(\).,-volV\(\)0123456789\(\).,-volV\)](https://doi.org/10.1007/s11258-018-0877-4(0123456789().,-volV()0123456789().,-volV))

541 Ranal, M. A., De Santana, D. C., 2006. How and why to measure the germination process? Revista
542 Brasil Bot. 29, 1. <https://dx.doi.org/10.1590/S0100-84042006000100002>

543 Rangel-Buitrago, N., Castro-Barros, J. D., Gracia, A., Villamil Villadiego, J. D., Williams, A. T.,
544 2018. Litter impacts on beach/dune systems along the Atlantico Department, the Caribbean
545 coastline of Colombia. Mar Pollut Bull. 137, 35-44.
546 <https://doi.org/10.1016/j.marpolbul.2018.10.009>

547 Rochman, C.M., Tahir, A., Williams, W.L., Baxa, D.V., Lam, R., Miller, J.T., Then, F.,
548 Werorilangi, S., Teh, S.J., 2015. Anthropogenic debris in seafood: Plastic debris and fibers
549 from textiles in fish and bivalves sold for human consumption. Sci Rep. 5, 14340.
550 <https://doi.org/10.1038/srep14340>

551 Rueden, C.T., Schindelin, J., Hiner, M.C., DeZonia, B.E., Walter, A.E., Arena, E.T., Eliceiri, K.W.,
552 2017. ImageJ2: ImageJ for the next generation of scientific image data. BMC
553 Bioinformatics 18:529, <https://doi.org/10.1186/s12859-017-1934-z>

554 Ryan, P.G., Moore, C.J., van Franeker, J.A., Moloney, C.L., 2009. Monitoring the abundance of
555 plastic debris in the marine environment. Phil Trans R Soc B. 364, 1999–2012.
556 <https://doi.org/10.1098/rstb.2008.0207>

557 Schmuck, A.M., Lavers, J.L., Stuckenbrock, S., Sharp, P.B., Bond, A.L., 2017. Geophysical
558 features influence the accumulation of beach debris on Caribbean islands. Mar Pollut Bull. 121,
559 45-51. <https://doi.org/10.1016/j.marpolbul.2017.05.043>

560 Scott, G. A. M., 1963. *Glaucium flavum* Crantz. J Ecol. 51, 743-754.

561 Sforzini, S., Oliveri, L., Chinaglia, S., Viarengo, A., 2016. Application of biotest for the
562 determination of soil ecotoxicity after exposure to biodegradable plastics. Front Env Sci. 4: 68.
563 <https://doi.org/10.3389/fenvs.2016.00068>

564 Šilc, U., Kuzmic, F., Cakovic, D., Stesevic, D., 2018. Beach litter along various sand dune habitats
565 in the southern Adriatic (E Mediterranean). Mar Pollut Bull. 128, 353-360.
566 <https://doi.org/10.1016/j.marpolbul.2018.01.045>

567 Staples, C., Friederich, U., Hall, T., Klečka, G., Mihaich, E., Ortego, L., Caspers, N., Hentges, S.,
568 2010. Estimating potential risks to terrestrial invertebrates and plants exposed to bisphenol A
569 in soil amended with activated sludge biosolids. *Environ Toxicol Chem.* 29, 467-475.
570 <https://doi.org/10.1002/etc.49>

571 Sykes, M. T., Wilson, J. B., 1989. The effect of salinity on the growth of some New Zealand sand
572 dune species. *Acta Bot Neerl.* 38, 173-182. <https://doi.org/10.1111/j.1438-8677.1989.tb02040.x>

573 Talsness, C.E., Anderson Andrade, J. M., Kuriyama, S.N., Taylor, J.A., vom Saal, F.S., 2009.
574 Components of plastic: experimental studies in animals and relevance for human health. *Phil*
575 *Trans R Soc B.* 364, 2079–2096. <https://doi.org/10.1098/rstb.2008.0281>

576 Teuten, E. L., Saquing, J. M., Knappe, D. R. U., Barlaz, M. A., Jonsson, S., Bjorn, ... Takada, H.,
577 2009. Transport and release of chemicals from plastics to the environment and to wildlife. *Phil*
578 *Trans R Soc B.* 364, 2027-2045. <https://doi.org/10.1098/rstb.2008.0284>

579 Thanos, C. A., Georghiou, K., Skarou, F., 1989. *Glaucium flavum* seed germination- an
580 ecophysiological approach. *Ann Bot.* 63, 121-130.
581 <https://doi.org/10.1093/oxfordjournals.aob.a087714>

582 Tosin, M., Weber, M., Siotto, M., Lott, C., Degli Innocenti, F., 2012. Laboratory test methods to
583 determine the degradation of plastics in marine environmental conditions. *Front Microbiol.* 3,
584 225. <https://doi.org/10.3389/fmicb.2012.00225>

585 Turner, G.D., Lau, R.R., Young, D.R., 1988. Effect of acidity on germination and seedling growth
586 of *Paulownia tomentosa*. *J Appl Ecol.* 25, 561-567. <https://doi.org/10.2307/2403844>

587 Underwood, A.J., 1997. Experiments in ecology: their logical design and interpretation using
588 analysis of variance. Cambridge University press. <https://doi.org/10.1017/CBO9780511806407>

589 Underwood, A. J., Chapman, M. G., 1998. GMAV, Version 5.0 for Windows.

590 UNEP, 2015. Biodegradable plastics and marine litter. Misconceptions, concerns and impacts on
591 marine environments. United Nations Environment Programme (UNEP), Nairobi (2015).

592 Ungar, I.A., 1978. Halophyte seed germination. *Botanical Review*. 44, 233–264. *JSTOR*,
593 www.jstor.org/stable/4353933.

594 Wilcox, C., Puckridge, M., Schuyler, Q.A., Townsend, K., Hardesty, B.D., 2018. A quantitative
595 analysis linking sea turtle mortality and plastic debris ingestion. *Sci Rep*. 8, 12536.
596 <https://doi.org/10.1038/s41598-018-30038-z>

597 Woodell, S. R. J., 1983. Salinity and seed germination patterns in coastal plants. *Vegetatio*. 61, 223-
598 229.

599 Yang, Y., Dou, Y., An, S., Zhu, Z., 2018. Abiotic and biotic factors modulate plant biomass and
600 root/shoot (R/S) ratios in grassland on the Loess Plateau, China. *Sci Tot Environ*. 636, 621–
601 631. <https://doi.org/10.1016/j.scitotenv.2018.04.260>

602 Zhang, C., Feng, Y., Liu, Y., Chang, H., Li, Z., Xue, J., 2017. Uptake and translocation of organic
603 pollutants in plants: a review. *J Integr Agric*. 16, 1659-1668. [https://doi.org/10.1016/S2095-](https://doi.org/10.1016/S2095-3119(16)61590-3)
604 [3119\(16\)61590-3](https://doi.org/10.1016/S2095-3119(16)61590-3)

605

606

607

608

609

610

611

612

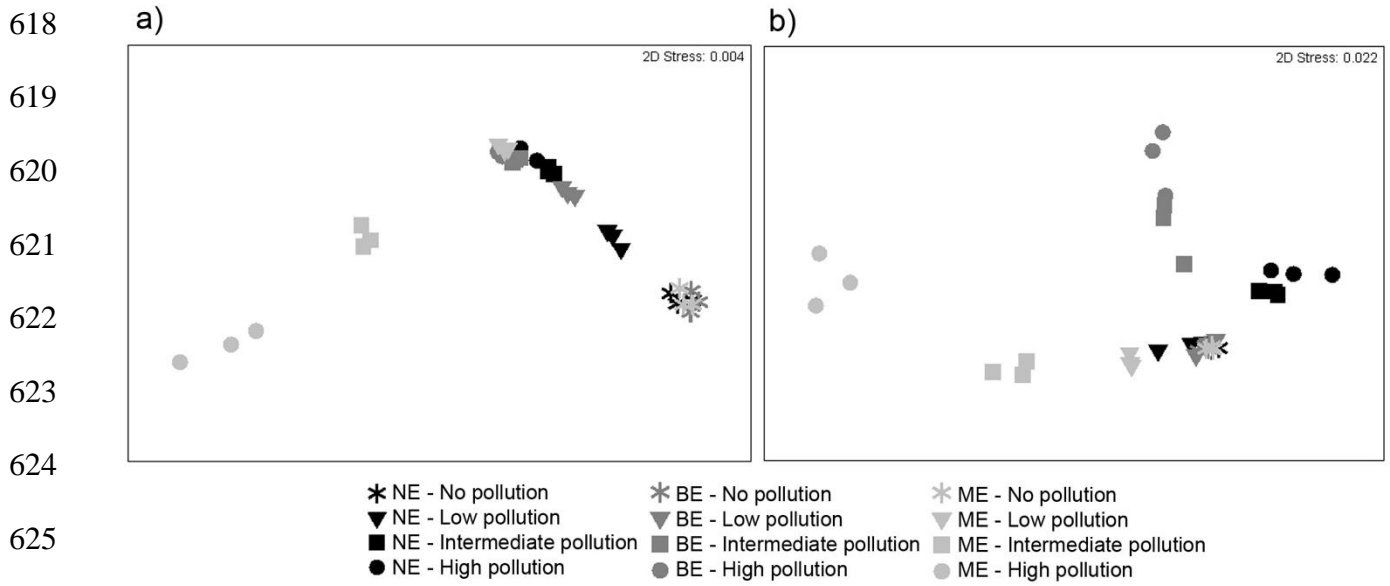
613

614

615

616

617



627 **Fig. 1.** Non-metric multidimensional scaling ordination (MDS) of chemical/physical characteristics
 628 of leachates from obtained from different amounts of not-exposed (NE), beach-exposed (BE) and
 629 marine-exposed (ME) PE (a) and MB (b) bags simulating different degrees of pollution.

630 2-column fitting image

631

632

633

634

635

636

637

638

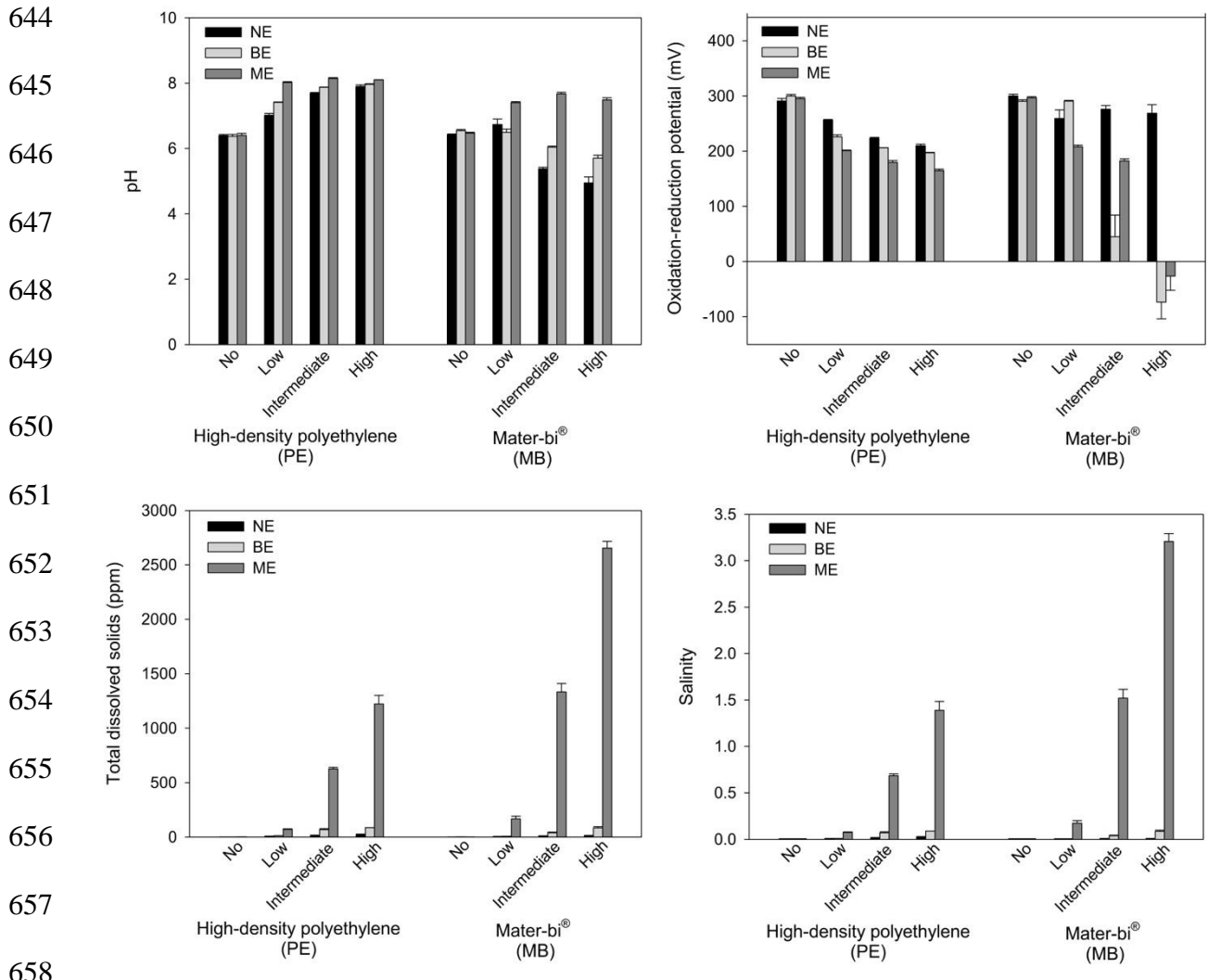
639

640

641

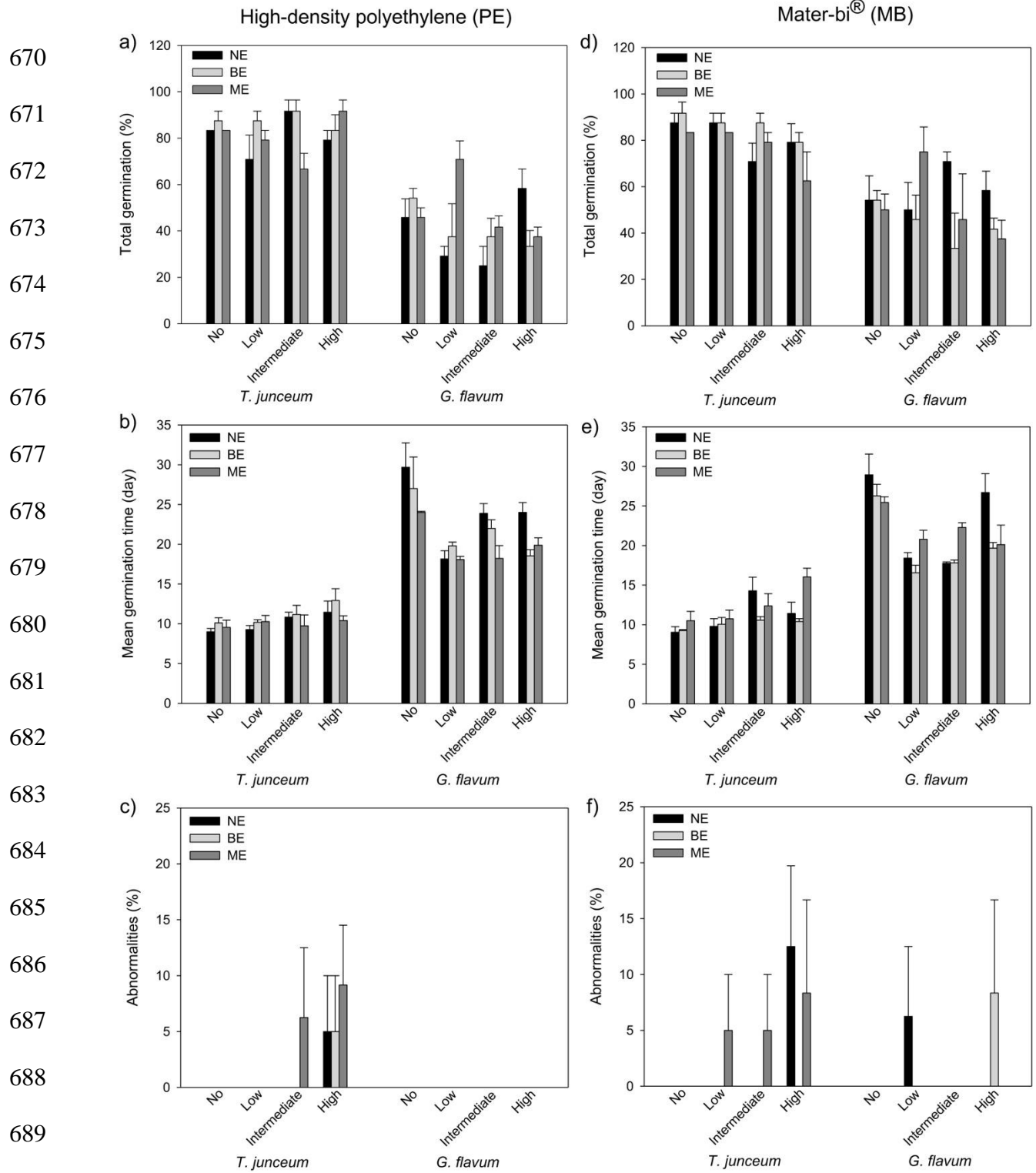
642

643



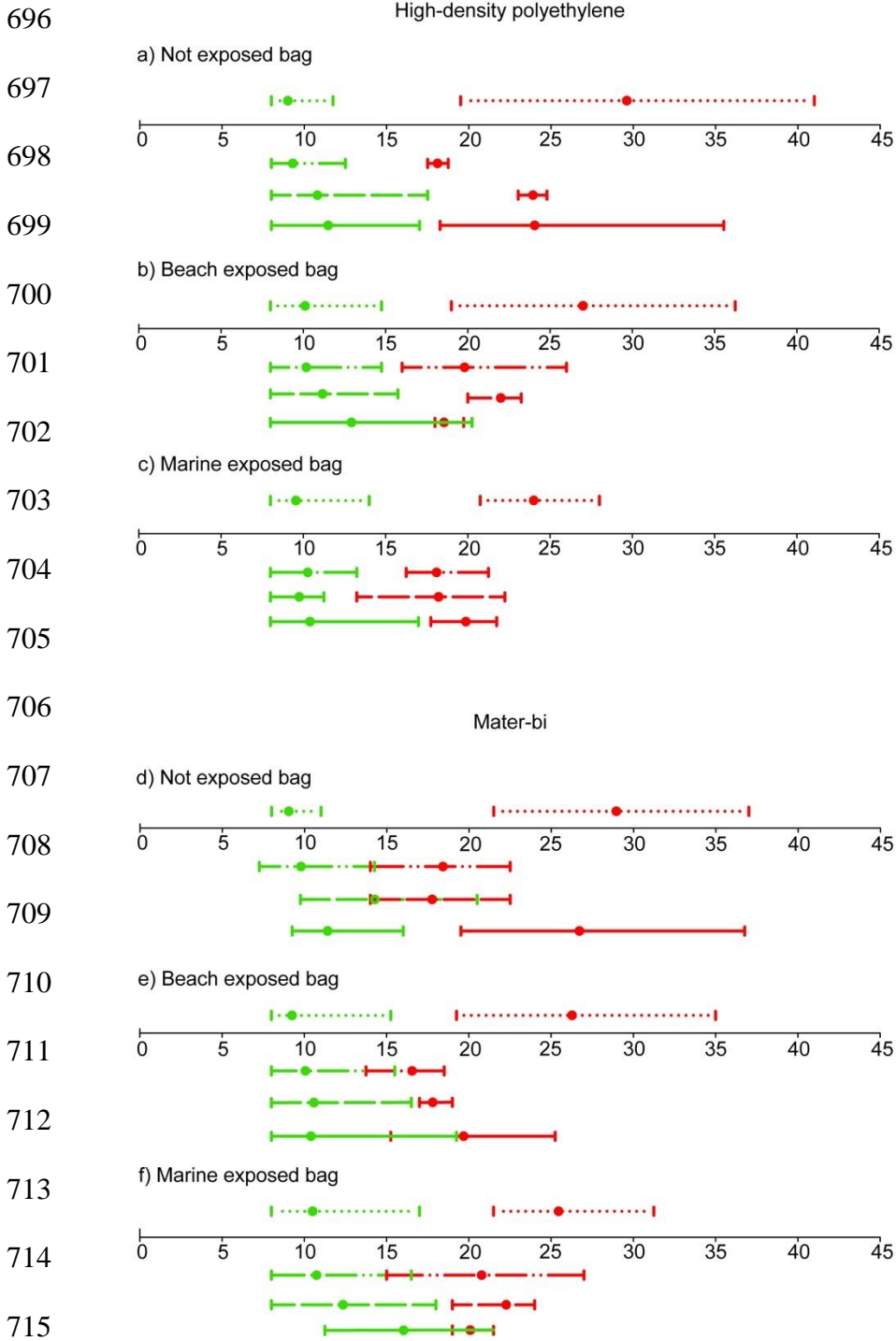
659 **Fig. 2.** Chemical/physical variables (pH, oxidation-reduction potential, total dissolved solids and
 660 salinity) of leachates obtained from different amounts of not-exposed (NE), beach-exposed (BE)
 661 and marine-exposed (ME) PE and MB bags simulating different degrees of pollution. Data are
 662 mean \pm SE, n = 3.

663 2- column fitting image



691 **Fig. 3.** Percentage of germination and mean germination time of seeds and percentage of abnormal
 692 seedlings of *T. junceum* and *G. flavum* grown with leachates obtained from different amounts of
 693 not-exposed (NE), beach-exposed (BE) and marine-exposed (ME) PE (a, b, c) and MB (d, e, f) bags
 694 simulating different degrees of pollution. Data are mean \pm SE, n = 4.

695 2-column fitting image

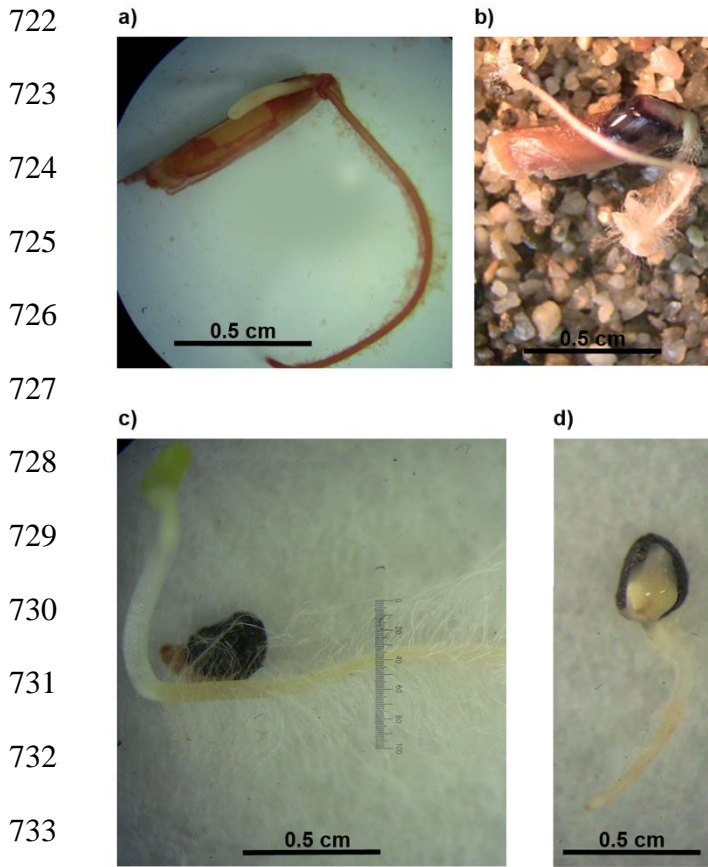


716 **Fig. 4.** The time spread of germination and the mean germination time of *T. junceum* (green) and *G.*
 717 *flavum* (red) seeds treated with bag leachates at various pollution degrees (— high, — —
 718 intermediate, — · — low and ······ no pollution).

719 2-column fitting image

720 Color image in online version and grayscale image in printed version

721



735 **Fig. 5.** Normal and abnormal seedlings lacking
736 aboveground organs of *T. junceum* (a, b) and *G.*
737 *flavum* (c, d) grown with bag leachates.

738 Single column-fitting image

739 Color image in online version and grayscale

740 image in printed version

741

742

743

744

745

746

747

748
749
750
751
752
753
754
755
756
757
758
759
760
761
762
763
764
765
766
767
768
769
770
771
772
773

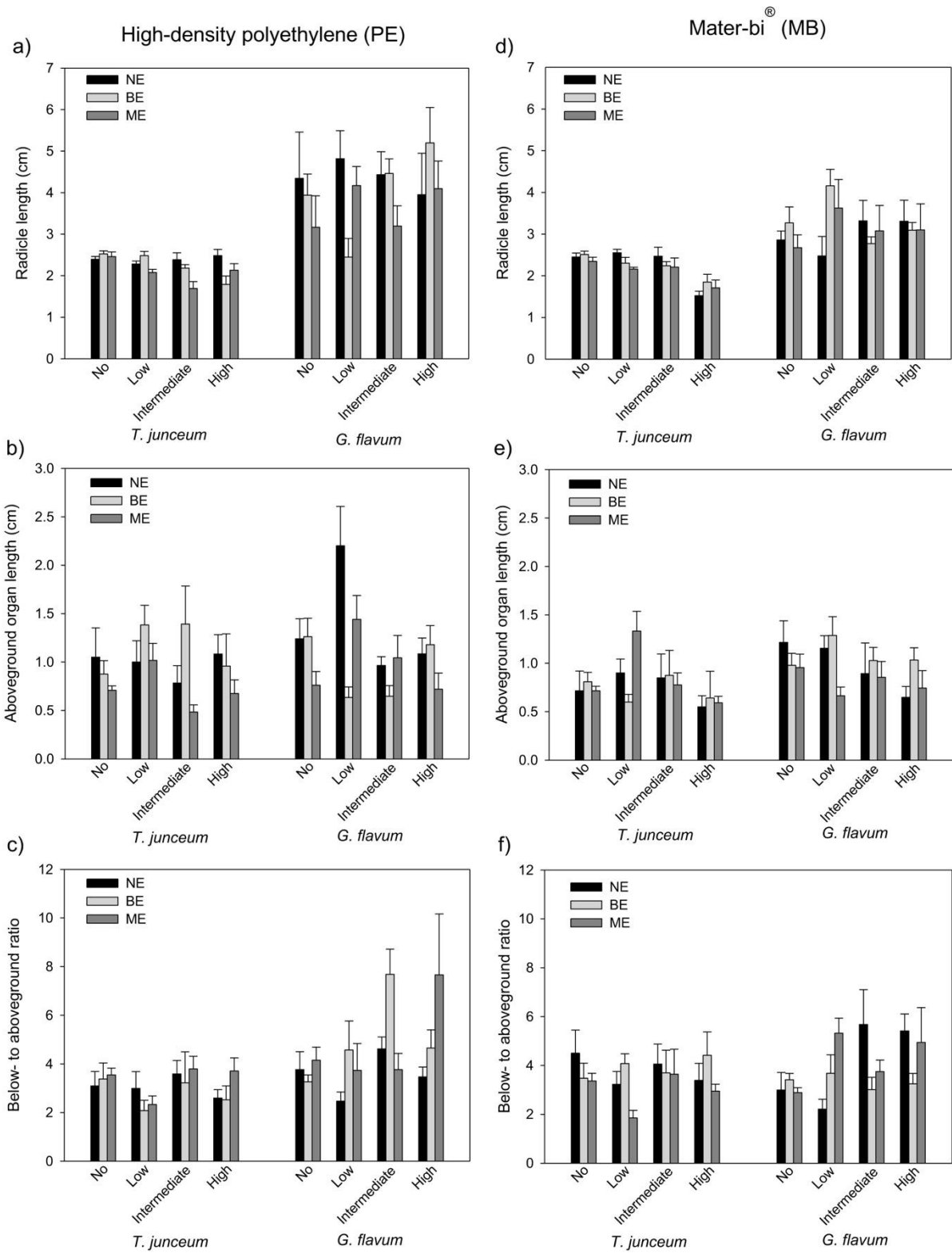


Fig. 6. Radicle length, aboveground organ length, and below- to aboveground ratio of seedlings grown with leachates obtained from different amounts of not-exposed (NE), beach-exposed (BE) and marine-exposed (ME) PE (a, b, c) and MB (d, e, f) bags simulating different degrees of pollution. Data are mean \pm SE, n = 6.

2-column fitting image

1 **Table 2**

2 Results of *a posteriori* pair-wise comparison test performed on
 3 the statistically significant term Plastic x Exposure x Pollution in
 4 the PERMANOVA on pH, oxidation-reduction potential (ORP),
 5 total dissolved solids (TDS) and salinity data measured in bag
 6 leachates.

7	pH	MB ≠ PE:	NE (I, H) - BE, ME (L, I, H)	
8		NE ≠ BE ≠ ME:	PE (L, I) - MB (I, H)	NE = BE ≠ ME: PE (H) - MB (L)
9		C ≠ H ≠ I ≠ L:	PE (NE, BE)	C ≠ L ≠ I = H: PE (ME)
10		C = L ≠ I = H:	MB (NE)	C = L ≠ I = H: MB (BE)
11		C ≠ L = I = H:	MB (ME)	
12	ORP	MB ≠ PE:	NE (I, H) - BE (L, I, H) - ME (H)	
13		NE ≠ BE ≠ ME:	PE (L, I, H) - MB (I)	NE = BE ≠ ME: MB (L)
14		NE ≠ BE = ME:	MB (H)	
15		C ≠ L ≠ I ≠ H:	PE (NE, BE, ME) - MB (ME)	C ≠ L ≠ I = H: MB (BE)
16	TDS	MB ≠ PE:	NE (I) - BE (L) - ME (L, I, H)	
17		NE ≠ BE ≠ ME:	PE, MB (I, H)	NE = BE ≠ ME: PE, MB (L)
18		C ≠ L ≠ I ≠ H:	PE (ME) - MB (BE, ME)	C ≠ L ≠ I = H: PE (NE, BE)
19		C ≠ L = I = H:	MB (NE)	
20	Salinity	MB ≠ PE:	NE (H) - ME (L, I, H)	
21		NE ≠ BE ≠ ME:	PE, MB (I, H)	NE = BE ≠ ME: PE, MB (L)
22		C ≠ L ≠ I ≠ H:	PE, MB (ME)	C = L ≠ I = H: PE (NE, BE)
23		C = L ≠ I = H:	MB (BE)	

24 H: high pollution, I: intermediate pollution, L: low pollution, C: control or no
 25 pollution, NE: not-exposed bag, BE: beach-exposed bag, ME: marine-exposed (ME),
 26 MB: Mater-bi[®], PE: high-density polyethylene (PE).

27

1 **Table 3**

2 Results of multivariate PERMANOVA analyses performed on (a) germination (percentage of
 3 germination and mean germination time) and (b) growth variables (radicle length, aboveground
 4 organ length and below- to aboveground ratio) of *T. junceum* and *G. flavum* seedlings grown with
 5 leachates obtained from different amounts of not-exposed, beach- or marine-exposed MB or PE
 6 bags simulating different degrees of pollution. Significant results are in bold.

7	a) Germination variables				b) Growth variables		
	8 Source	d.f.	Pseudo-F	p	d.f.	Pseudo-F	p
9	Species (S)	1	257.82	<0.001	1	48.00	<0.001
10	Plastic (P)	1	1.00	0.347	1	5.87	0.004
11	Exposure (E)	2	1.12	0.336	2	1.94	0.108
12	Pollution (Po)	3	3.53	0.003	3	4.21	0.001
13	S x P	1	1.80	0.164	1	4.77	0.010
14	S x E	2	3.66	0.009	2	1.09	0.348
15	S x Po	3	5.28	0.003	3	2.27	0.039
16	P x E	2	2.64	0.049	2	1.88	0.114
17	P x Po	3	0.82	0.530	3	1.31	0.250
18	E x Po	6	1.54	0.132	6	1.19	0.282
19	S x P x E	2	0.78	0.495	2	4.45	0.002
20	S x P x Po	3	0.48	0.786	3	0.36	0.896
21	S x E x Po	6	1.58	0.113	6	2.04	0.025
22	P x E x Po	6	0.92	0.508	6	1.15	0.311
23	S x P x E x Po	6	2.05	0.034	6	3.21	<0.001
24	Residual	144			240		

25

26

Table 4

[Click here to download Table: Table 4.docx](#)

1 **Table 4**

2 Results of ANOVA analyses performed on the percentage of germination, mean germination time
3 and percentage of abnormal seedlings of *T. junceum* and *G. flavum* treated with leachates obtained
4 from different amounts of not-exposed, beach- or marine-exposed MB or PE bags simulating
5 different degrees of pollution. Significant results are in bold.

Source	d.f.	<u>Total germination (%)</u>		<u>Mean germination time</u>		<u>Abnormalities (%)</u>	
		F	p	F	p	F	p
Species (S)	1	252.90	<0.001	980.21	<0.001	4.37	0.038
Plastic (P)	1	2.48	0.117	0.92	0.339	1.01	0.317
Exposure (E)	2	0.02	0.975	2.81	0.063	0.79	0.457
Pollution (Po)	3	2.45	0.065	14.88	<0.001	4.48	0.004
S x P	1	4.86	0.029	2.33	0.129	0.21	0.646
S x E	2	5.16	0.006	4.86	0.009	2.63	0.075
S x Po	3	0.50	0.680	28.61	<0.001	2.34	0.076
P x E	2	1.81	0.167	9.10	<0.001	0.28	0.752
P x Po	3	1.60	0.191	0.49	0.692	0.43	0.732
E x Po	6	3.20	0.005	1.48	0.190	0.33	0.920
S x P x E	2	1.07	0.347	0.44	0.642	0.60	0.548
S x P x Po	3	0.91	0.438	1.78	0.154	0.09	0.965
S x E x Po	6	2.46	0.026	2.58	0.021	1.32	0.251
P x E x Po	6	0.74	0.621	0.56	0.764	0.20	0.977
S x P x E x Po	6	2.42	0.029	3.15	0.006	1.23	0.294
Residual	144						

24

25

26

1 **Table 5**

2 Results of SNK post-hoc test of the significant
 3 terms in the ANOVA on the percentage of seed
 4 germination, the mean germination time (MGT)
 5 and the percentage of abnormal seedlings of *T.*
 6 *junceum* (T) and *G. flavum* (G) treated with bag
 7 leachates.

8	Total germination	<i>Species (S) x Exposure (E)</i>
9		G < T : NE, BE, ME
10		<i>E x Pollution (Po)</i> :
11		NE = BE < ME : L
12		C = I = H < L : ME
13	MGT	<i>S x Plastic (P) x E x Po</i>
14		T < G
15		PE < MB : T (NE (I), ME (H)) - G (ME (I))
16		MB < PE : G (NE, BE (I))
17		NE = BE < ME : T (MB(H))
18		NE = BE > ME : G (PE (I))
19		NE > BE = ME : G (PE, MB (H))
20		C = L = I < H : T (MB(ME))
21		C > I = H > L : G (PE(NE))
22		C > L = I = H : G (PE (BE, ME) -MB (BE))
23		C = H > I = L : G (MB(NE))
24	Abnormalities	<i>Po</i>
25		C = L = I < H

26 H: high pollution, I: intermediate pollution, L: low pollution, C:
 27 control or no pollution, NE: not-exposed bag, BE: beach-exposed
 28 bag, ME: marine-exposed (ME), MB: Mater-bi[®], PE: high-density
 29 polyethylene (PE).

30

31

1 **Table 6**

2 Results of ANOVA analyses performed on radicle length, aboveground organ length and below- to
 3 aboveground ratio of *T. junceum* and *G. flavum* seedlings grown with leachates obtained from
 4 different amounts of not-exposed, beach- or marine-exposed MB or PE bags simulating different
 5 degrees of pollution. Significant results are in bold.

Source	d.f.	Radicle		Aboveground		Below- to aboveground	
		F	p	F	p	F	p
Species (S)	1	138.10	<0.001	7.71	0.005	16.85	<0.001
Plastic (P)	1	12.23	<0.001	8.70	0.003	0.01	0.904
Exposure (E)	2	2.94	0.054	3.69	0.026	0.34	0.709
Pollution (Po)	3	1.35	0.259	5.77	<0.001	4.90	0.002
S x P	1	7.25	0.007	0.04	0.842	6.75	0.009
S x E	2	0.30	0.742	2.01	0.136	1.53	0.217
S x Po	3	4.81	0.002	0.72	0.540	2.60	0.053
P x E	2	1.92	0.148	2.34	0.098	1.44	0.238
P x Po	3	1.91	0.128	1.41	0.239	0.95	0.417
E x Po	6	1.00	0.428	1.55	0.162	1.30	0.257
S x P x E	2	0.41	0.664	5.73	0.003	5.64	0.004
S x P x Po	3	0.93	0.428	0.42	0.741	0.24	0.870
S x E x Po	6	1.20	0.306	2.41	0.027	2.49	0.023
P x E x Po	6	1.89	0.084	0.58	0.747	1.45	0.196
S x P x E x Po	6	4.29	<0.001	3.68	0.001	2.27	0.037
Residual	240						

25

26

27

1 **Table 7**

2 Results of SNK post-hoc test on the statistically significant term

3 Plastic x Exposure x Pollution in the ANOVA on radicle length,

4 aboveground organ length and below- to aboveground ratio of *T.*5 *junceum* (T) and *G. flavum* (G) seedlings grown with bag leachates.

6 Radicle length	T < G: PE (NE (L, I), BE (I, H), ME (L, I, H)) - MB (NE (H), BE, ME (L, H))
7	MB < PE: T (NE (H)) - G (NE (L), BE (I, H), ME (H))
8	PE < MB: G (BE (L))
9	NE = ME > BE: G (PE (L))
10	NE < BE = ME: G (MB (L))
11	H < I = L = C: T (MB (NE))
12	L < C = I = H: G (PE (BE))
13 Aboveground	T < G: PE (NE (L), ME (I)) - MB (BE (L))
14 organ length	G < T: PE (BE (L, I)) - MB (ME (L))
15	MB < PE: T (BE (L)), G (NE, ME (L))
16	PE < MB: G (BE (L))
17	NE = ME < BE: T (PE (I))
18	NE > ME > BE: G (PE (L))
19	L > C = I = H: G (PE (NE))
20 Below- to	T < G: PE (BE (L, I, H), ME (H)) - MB (NE (H), ME (L))
21 aboveground ratio	MB < PE: G (BE (I))
22	PE < MB: T (BE (H, L))
23	NE = BE > ME: T (MB (L))
24	I > H = L = C: G (PE (BE))
25	H > I = L = C: G (PE (ME))
26	H = I > L = C: G (MB(NE))

27 H: high pollution, I: intermediate pollution, L: low pollution, C: control or no pollution,

28 NE: not-exposed bag, BE: beach-exposed bag, ME: marine-exposed (ME), MB: Mater-

29 bi[®], PE: high-density polyethylene (PE).

30