1	Pistacia lentiscus essential oil has repellent effect against three major insect pest
2	of pasta
3	
4	Hind Houria Bougherra ^a , Stefano Bedini ^b , Guido Flamini ^c , Francesca Cosci ^b ,
5	Kamel Belhamel ^a , Barbara Conti ^b *
6	
7	^a Laboratory of Organic Materials, Department of Process Engineering , Faculty of
8	Technology, University of Bejaia, DZ 06000, Algeria.
9	^b Department of Agriculture, Food and Environment, University of Pisa, Italy.
10	^c Department of Pharmaceutical Sciences, University of Pisa, Italy.
11	
12	
13	
14	
15	
16	
10	
17	
18	
19	
20	
21	
2223	
24	
∠ ¬r	

ABSTRACT

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

Rhyzopertha dominica, Sitophilus zeamais, and Tribolium confusum are three of the major food-stuff pests who cause important economic losses of shelved products with special reference to pasta. Due to its long shelf life, pasta is highly exposed to insects that can penetrate into the packaging with consequences economically severe. Ecofriendly strategies to prevent such insect attacks to the final packaged product are therefore highly foreseen by pasta companies. Due to their repellent properties, essential oils, extracted from aromatic plants, could represent a valid, eco-friendly alternative to chemical repellents. In this study, we evaluated the repellent activity of Pistacia lentiscus essential oil (PEO) and its main chemical components by two different bioassay with and without the presence of pasta. Results showed that the whole PEO exerts a broad-range aspecific repellency among the target pests with RD₅₀ values ranging from 0.010 to 0.037 µL cm⁻². On the contrary, the repellence of PEO components resulted to vary depending on the compound and on the pest species. Among the PEO chemical components, relative median potency analyses indicated that β -caryophyllene was able to exert the highest repellency rates against S. zeamais (RD₅₀ 0.046 µM cm⁻²). The comparison between the two bioassays, with and without pasta, indicated that the two methodologies gave consistent results. Overall, our research firstly showed that, because of their effectiveness as repellents, PEO and its major constituents could represent valid and safe tools against pasta pests.

45

46

47

48

Keywords: Rhyzopertha dominica, Sitophilus zeamais, Tribolium confusum, area

49 preference bioassay, pitfall bioassay, repellence

1. Introduction

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

About 13.6 million tons of pasta are produced worldwide (International Pasta Organization, 2012) with a market value that, for the only Italy, the main pasta producer, has been estimated as about 4.6 million Euro (UNIPI, 2012). Due to such a large production, and the particularly long shelf life (about 2-3 years), the control of pests is an important aspect of pasta post-production management. Stored-food insect pests cause severe quantitative and qualitative losses in stored raw materials, such as cereals or stored grains as well as in semi-processed and in final food products, such as pasta (Hou et al., 2004; Germinara et al., 2010; Bachrouch et al., 2010; Trematerra and Süss, 2006). The Coleoptera Rhyzopertha dominica (F.) (Bostrychidae), Sitophilus zeamais Motsch. (Dryophthoridae), and Tribolium confusum Du Val (Tenebrionidae) are three of the major pasta pests who cause important economic losses of shelved products (Trematerra and Süss, 2006). Actually, due to the long pasta shelf life, insects can easily penetrate into the packaging and reproduce many generations (Locatelli and Süss, 2002). Unfortunately, a single insect occurring in a pasta package, although non compromising the quality of the product, is enough to affect seriously the image of the company manufacturing or distributing the goods (Kim et al., 2010) with consequences that can be economically severe (Hou et al., 2004; Licciardello et al., 2013). For these reasons, strategies to prevent insect attacks to the final packaged product such as the development of new repellent packaging methods are highly foreseen by pasta companies (Cagri et al., 2004; Hou et al., 2004; Germinara et al., 2010). However, because the concerns about their toxicity, chemical insecticides and repellents are not well accepted by costumers. Therefore, alternatives approaches to chemical fumigants that play a major role in insect pest control in stored food are

75 needed.

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

In recent years, essential oils (EOs) of aromatic plants, characterized by low toxicity to mammalians and already extensively used in the food industries as supplements, and flavouring compounds, received a great attention as pest control agents due to their insecticidal, repellent, and/or antifeedant properties (Isman, 2006; Nerio et al., 2010; Conti et al., 2010; 2011; Benelli et al., 2012). As a consequence, aromatic plants are studied as potential sources of repellents and insecticides (Nerio et al., 2010, Shaaya and Kostyukovysky, 2006; Conti et al., 2010; Caballero-Gallardo et al., 2012; Olivero-Verbel et al., 2013). Pistacia lentiscus L. is an aromatic evergreen shrub belonging to the Anacardiaceae family, largely distributed in the Mediterranean basin (Abdelwahed et al., 2007). Although P. lentiscus essential oil (PEO) has been showed to have good insecticidal activity (Lamiri et al., 2001; Traboulsi et al., 2002), to the best of our knowledge, no information are available about its properties as insect repellent. In this research the EO extracted from Algerian P. lentiscus plants was analyzed by gas chromatography (GC) and by gas chromatography/electron impact mass spectroscopy (GC-EIMS) and then its repellent activity against adults of R. dominica, S. zeamais, T. confusum was evaluated. Since EOs bioactivity is due to the combined action of their chemical compounds (Hummelbrunner and Isman, 2001), in this research we also tested the specific activity of α -terpineol, α -pinene, and β caryophyllene, three main terpene constituents of PEO. In order to evaluate the repellency also in the presence of pasta, we compared the area preference method (Tapondjiou et al., 2005), a commonly used method to assess the repellence with a two choice pitfall method (Germinara et al., 2007), in which the repellent activity of the PEO main constituents was counterbalanced by the attractiveness of the pasta.

100 101 2. Materials and methods 102 103 2.1. Plant material 104 105 Pistacia lentiscus L. leaves were collected from wild plants, at flowering stage, in 106 the locality of Fenaïa, Béjaia (211 km East of Algiers, 481 m above sea level: 107 36°40'28.08" N; 4°49'53.97" E) in June 2012. 108 109 2.2. Essential oil extraction and GC-MS analyses 110 111 The harvested leaves were dried in the shade, at room temperature (20-25°C) until 112 constant weight, and then coarsely ground and hydro-distilled in a Clevenger-type 113 apparatus for 4 h. The resulting essential oil was dried over anhydrous sodium 114 sulphate and stored in a glass vial at 4°C until use. 115 Gas chromatography (GC) analyses were carried out with an HP-5890 Series II 116 instrument equipped with HP-WAX and HP-5 capillary columns (30 m × 0.25 mm, 117 0.25 µm film thickness), working with the following temperature program: 60°C for 118 10 min, ramp of 3°C min⁻¹ up to 220°C; injector and detector temperatures 250°C; 119 carrier gas helium (2 ml min⁻¹); detector dual FID; split ratio 1:30; injection of 0.5 μl 120 (10% hexane solution). Components identification was carried out, for both columns, 121 by comparing their retention times with those of pure authentic samples and by means 122 of their linear retention index (LRI), relative to the series of *n*-hydrocarbons. Gas 123 chromatography-electron impact mass spectroscopy (GC-EIMS) analyses were 124 performed with a Varian CP-3800 gas chromatograph, equipped with a HP-5 capillary column (30 m × 0.25 mm; coating thickness 0.25 μm) and a Varian Saturn 2000 ion trap mass detector with the following analytical conditions: injector and transfer line temperatures 220°C and 240°C respectively; oven temperature programmed from 60°C to 240°C at 3°C min⁻¹; carrier gas helium at 1 ml min⁻¹; injection of 0.2 μl (10% hexane solution); split ratio 1:30. Constituents identification was based on the comparison of retention times with those of authentic samples, comparing their LRIs with the series of *n*-hydrocarbons and using computer matching against commercial (Adams, 1995) and home-made library mass spectra (built up from pure substances and components of known oils and MS literature data (Davies, 1990; Adams, 1995). Moreover, molecular weights of all identified substances were confirmed by gas chromatography-chemical ionization mass spectrometry (GC-CIMS), using methanol as the chemical ionizing gas.

2.3. Insect cultures and rearing conditions

Strains of *R. dominica, S. zeamais*, and *T. confusum* were reared at the Department of Agriculture, Food and Environment of the University of Pisa, since 2000. Insects were reared at room temperature, 65% R.H., natural photoperiod, in $(20\times25\times15 \text{ cm})$ plastic boxes containing grains of maize and wheat and covered by a nylon net allowing air exchange. Since the adults remain until three days into the grain, homogeneous adults (0-3 days old) were obtained by removing adults from the box and the daily newly emerged insects were used for the bioassays.

2.4. Area preference bioassay

The bioassays were conducted following the method described by Tapondjiou et al. (2005). Half filter paper disks (8 cm Ø) were treated with 500 μL of PEO as ethanolic solution at doses corresponding from 0.01 to 5 µL cm². Single PEO constituents (purchased from Sigma-Aldrich®) were tested as ethanolic solutions at doses ranging from 0.02 to 4 µM cm⁻². The treated filter paper disks were dried under a fan. Each Petri dish's bottom (8 cm Ø) was half-covered with half filter paper treated with the PEO or chemical solutions, while the other half, was covered with a half filter paper disk treated with 500 µl of ethanol (control). Twenty unsexed adults were introduced in each Petri dish, and the lid was sealed with Parafilm®. The Petri dishes were maintained at 25 ± 1 °C, 65% R.H., in the dark. Five replicates were performed for each assay, and insects were used only once. The number of insects on the two half of the Petri dish was recorded after 1, 3, and 24 h from the beginning of the test. The percent repellence (PR) of PEO and of each volatile compound was calculated by the formula: PR (%) = $[(Nc-Nt)/(Nc+Nt)] \times 100$ where Nc is the number of insects present in the control half paper and Nt the number of insects present in the treated one.

166

167

165

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

2.5. Two-choice pitfall bioassay

168

169

170

171

172

173

174

The repellent activity of the volatile compounds was evaluated against R. dominica, S. zeamais, and T. confusum adults, using the bioassay described by Germinara et al. (2007). The bioassay was conducted in a steel arena (32 cm $\emptyset \times 12$ cm high) with, in the bottom, two diametrically opposed holes (3 cm \emptyset) located 3 cm from the sidewall. 10 μ l of chemicals or ethanol (control) were adsorbed onto a filter paper disk (1 cm \emptyset) suspended at the center of each hole by a cotton thread taped to

the lower surface of the arena. Glass flasks (500 ml) filled with 100 gr of pasta (spaghetti n. 5, [©]Barilla G. e R. Fratelli S.p.A.) were positioned under each hole, and the inside surface of their necks were coated with paraffin oil to prevent insects, that have previously chosen, from returning to the arena. Preliminary trials allowed us to exclude any repellent or attractant effect of paraffin oil. The floor of the arena was covered with filter paper to provide an uniform surface and to facilitate insect movements. Sixty insects, deprived of food for at least 4 hours, were placed under an inverted Petri dish (3 cm $\emptyset \times 1.3$ cm high) at the center of the arena and allowed to acclimate for 30 min. The arenas were covered with steel lids and sealed with Parafilm[®] to prevent insects from escaping and were left for 24 h in the dark at 25 \pm 1°C and 65% R. H. Five replicates were performed for each assay, and insects were used only once. The number of insects in the flasks was recorded 24 h from the beginning of the test. The percent repellence (PR) of each volatile was then calculated after 24 h using the formula: PR (%) = $[(Nc-Nt)/(Nc+Nt)] \times 100$ where Nc was the number of insects present in the control flask and Nt the number of insects present in the treated flask. The number of non-choosing insects (Nn), that remained in the arena without choosing any of the two chambers with the food, were recorded.

192

193

191

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

2.6. Statistics and data analyses

194

195

196

197

198

199

Data were analyzed by one-way ANOVA (insect species or essential oil component as factor) or two-way ANOVA (insect species and essential oil component as fixed factors, essential oil concentration as covariate). Data were arcsine-transformed when needed to fulfill the assumptions of the ANOVA. Means and standard errors (S.E.) given in tables and figures are for untransformed data. In Petri

200	repellence tests on filter paper the median repellent dose (RD50) was calculated by
201	Log-probit regressions (Finney, 1971). Significant differences between RD50 values
202	were determined by estimation of confidence intervals of the relative median potency.
203	Differences among RD50 were judged to be statistically significant when values of the
204	95% confidence interval of relative median potency were \neq 1.0.
205	All the analyses were performed by the SPSS 22.0 software (SPSS Inc., Chicago, IL,
206	USA).
207	
208	3. Results
209	
210	3.1. Essential oil extraction and GC-MS analysis
211	
212	In the essential oil obtained from the flowering aerial parts 52 constituents were
213	identified, accounting for 97.8% of the whole oil. The main components were $\alpha\text{-}$
214	pinene (15.9%), β -caryophyllene (7.0%), α -terpineol (6.9%), and myrcene (6.4%).
215	Other important volatiles were 4-terpineol, β -pinene, and germacrene D (5.4, 5.1 and
216	5.1%, respectively) (Table 1).
217	The main represented chemical classes were monoterpene and sesquiterpene
218	hydrocarbons (45.9 and 22.8%, respectively). The corresponding oxygenated
219	derivatives were less represented in this oil (14.9 and 13.3%, respectively) (Table 2).
220	
221	3.2. Area preference bioassay
222	
223	The area preference method showed that <i>P. lentiscus</i> essential oil (PEO),
224	exerted a clear repellent activity against R. dominica, S. zeamais, and T. confusum

(Table 3). Probit regressions of repellence values showed that S. zeamais was the most susceptible species to PEO (RD₅₀ = $0.010 \,\mu L \, cm^{-2}$) while, the less sensitive species was R. dominica (RD₅₀ = $0.037 \mu L \text{ cm}^{-2}$) (Table 3). However, the relative median potency analyses of probits do not detected significant differences among species (Table 4). Such a broad-range activity of PEO was confirmed also by ANOVA that showed no significant differences among species at the end of the bioassays (24 h) ($F_{2,15} = 1.686$, P = 0.230). On the contrary, significant differences were detected among species when single compounds were utilized as repellents (Table 5). Relative median potency analyses indicated that β-caryophyllene was the most effective compound against S. zeamais and also significantly more effective than α -terpineol against R. dominica, while, α -terpineol resulted more effective than α pinene against *T. confusum* (Table 6). The relative median potency analysis showed also a significant different responsiveness of the three pest species to β -caryophyllene. Such responsiveness was: S. zeamais > T. confusum > R. dominica. Rhyzophertha dominica was also the less responsive species to α -terpineol and α -pinene (Table 4). Consistently, two ways ANOVA showed that the repellence after 24 h of PEO main components was significantly different as a function of the species ($F_{2,122} = 9.718$, P< 0.001), the compound ($F_{2,122} = 5.928$, P = 0.004) and that there was an interaction between the species and the repellent compound ($F_{4,122} = 2.632$, P = 0.038).

244

245

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

3.3. Two-choice pitfall bioassay

246

247

248

249

The effect of the PEO main components in the presence of pasta, was tested by the two-choice pitfall bioassay. The repellency of the PEO main components against *R. dominica* ranged from 4 to 37.4% for α -pinene and β -caryophyllene, respectively,

while the activity of the PEO main components against *T. confusum* ranged from 22.2 and 36.0% for α -pinene and α -terpineol, respectively (Fig. 1A). Consistently with the area preference bioassay, the most overall responsive species resulted to be S. zeamais with PR values from 23.6 for α -pinene and 58.2% for α -terpineol (Fig. 1A), while the less responsive was R. dominica with PR values from 4 for α-pinene and 37.4% for αterpineol (Fig. 1A). However, no statistically significant differences in repellence rates were found, as a function of species ($F_{2,45} = 2.298$, P = 0.115), compound ($F_{2,45}$ = 2.562, P = 0.091) and no interaction ($F_{4,45} = 1.609, P = 0.193$). In the two-choice pitfall we also observed a different percentage of individuals that at the end of the test did no choice and remained in the arena (non-choosing individuals). Such nonchoosing individuals ranged in average from 77.9 \pm 3.11 to 11.4 \pm 3.39% for R. dominica and S. zeamais, respectively (Fig. 1B). Two-way ANOVA indicates that the differences among the percentages of non-choosing individuals of the three species were statistically significant ($F_{2,45} = 127.856$; P < 0.001), while there was no significant effect of the PEO main components ($F_{2,45} = 1.703$; P = 0.196) and no interaction ($F_{4,45} = 2.246$; P = 0.083). A statistically significant moderate negative relationship between the percentage of non-choosing individuals and the PR (r =0.334; $F_{1,45} = 5.391$; P = 0.025) was also found by regression analysis.

268

269

267

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

4. Discussion

270

271

272

273

274

Results highlighted that PEO and its main chemical components have a clear repellent activity against *R. dominica*, *S. zeamais*, and *T. confusum*, the three pasta pest species tested.

To our knowledge this is the first assessment of *P. lentiscus* essential oil

repellency against insects. Overall, our data on the PEO are in accordance with previous studies showing the repellent effect of several plant essential oils on *R*. *dominica* (Jilani and Malik, 1973), *T. confusum* (Akou-Edi, 1983), and *S. zeamais* (Akou-Edi, 1983; Nerio et al., 2009; Conti et al., 2010). On the base of our data, PEO resulted able to exert an aspecific broad-range repellent activity with no statistically significant differences, in term of effectiveness, among the three insect species. In fact, PEO resulted about 3 to 27 fold more effective against *S. zeamais* than the essential oils extracted from several Colombian aromatic plants and about 30 fold the commercial insect repellent IR3535 (Nerio et al., 2009).

The chemical analysis of the essential oil extracted from Algerian P. lentiscus plants indicated that its composition, although consistent, presents some differences, with the composition of other essential oils extracted from P. lentiscus plants. Actually, in a previous study about Algerian P. lentiscus, Benyoussef and coworkers (2005) found 4-terpineol, α -terpineol, and germacrene D among the principal chemicals. However, such a differences in the chemical composition of P. lentiscus essential oil may be due to the different geographical area of growth and period of harvesting of the plant material (Zrira et al., 2003; Aouinti et al., 2013).

Contrary to what observed for the whole PEO, α -terpineol, α -pinene, and β -caryophyllene, the main components of PEO, when tested as single compounds, showed different effectiveness against the three species. These results confirm that the repellent activity of EOs is due to the combined action of its chemical components (Hummelbrunner and Isman, 2001).

Among the tree species studied, *S. zeamais* resulted the overall most sensitive species, while *R. dominica* was the less affected one. Differences in the bioactivity of EOs compounds against stored-product pest species were also observed by Rozman et

al. (2007) who found that 1,8-cineole and linalool were insecticidal to *R. dominica* but not effective against *T. castaneum*.

Accordingly to our data, the overall most active compound against the three pasta pests species, was β -caryophyllene. This result is consistent with previous observations by Chaubey (2012) who found that β -caryophyllene was more toxic and with higher anti-feeding activity than α -pinene against T. castaneum and S. oryzae. On the contrary, Zeng et al. (2010) reported no repellent activity of β -caryophyllene, against R. dominica and S. oryzae.

For what concern α -pinene and α -terpineol, our results are in agreement with previous reports indicating that α -pinene is effective against T. confusum (Ojimelukwe and Alder, 1999), S. oryzae (Lee et al., 2001), and S. zeamais (Karemu et al., 2013). Actually, in our experiment, the RD50 of α -pinene and α -terpineol resulted about five and two fold, respectively, the RD50 that can be deduced from the data by Ojimelukwe & Alder (1999) against T. confusum while, on the contrary, a much higher repellent activity of α -pinene against T. confusum was found by Kim et al. (2010).

In this experiment, to evaluate the bioactivity of the compounds we compared the commonly used area preference method with the pitfall bioassay that is, in theory, more close to a real situation because insects are never in direct contact with the tested compound and because the larger volume of the pitfall chamber allows a better gradient of concentration of the volatiles, whose repulsiveness is counterbalanced by the attractive presence of pasta.

The repellent activities of the PEO compounds assessed by the pitfall bioassay confirmed, overall, the results obtained by the area preference method. However, the pitfall bioassay allowed the detection of a part of the population remaining in the

arena at the end of the experiment (non-choosing individuals). Interestingly, the amount of such non-choosing individuals was significantly different among the species and resulted to be negatively correlated to the repellent activity of the tested compounds. Therefore, the possibility that in the area preference method, different species behaviors may introduce undetected biases into the estimates of repellency can not to be excluded.

5. Conclusions

The results obtained in this experiment show that P. lentiscus essential oil, is rich in chemical components such as α -terpineol, α -pinene, and β -caryophyllene that resulted very effective as repellents against insect pasta pests. Since, aromatic plants essential oils present a very low toxicity to human beings and pose no significant environmental risk, P. lentiscus essential oil represent an appealing sustainable alternative to chemical insects repellents that could be usefully utilized in the reduction of losses caused to stored pasta and other food-stuff by insect pests.

Acknowledgements

We would like to thanks Dr. Adjaoud Abdenour (Faculté des Sciences de la Nature et de la Vie, University of Bejaia, DZ 06000, Algeria) for the species identification of the collected plants, Mr. Paolo Giannotti for the skilled assistance during the set-up of the experiment and Dr. Riccardo Antonelli for the photographs of the insects.

350	References
351	
352	Abdelwahed, A., Bouhlela, I., Skandrania, I., Valentic, K., Kadrid, M., Guiraudd, P.,
353	Steimand, R., Mariotteb, A.M., Ghediraa, K., Laportec, F., Dijoux-Francab,
354	M.G., Chekir-Ghedira, L., 2007. Study of antimutagenic and antioxidant
355	activities of Gallic acid and 1,2,3,4,6-pentagalloylglucose from <i>Pistacia lentiscus</i> :
356	Confirmation by microarray expression profiling. Chemico-Biological Int. 165,
357	1-13.
358	Adams, R.P., 1995. Identification of Essential Oil Components by Gas
359	Chromatography-Mass-spectrometry. Allured Pub. Co., Carol Stream IL USA.
360	Aouinti, F., Imelouane, B., Tahri, M., Wathelet, J.P., Amhamdi, H., Elbachiri, A.,
361	2013. New study of the essential oil, mineral composition and antibacterial
362	activity of Pistacia lentiscus L. from Eastern Morocco. Research on Chemical
363	Intermediates, in press, doi: 10.1007/s11164-013-1134-z.
364	Akou-Edi, D., 1983. Effects of neem seed powder and oil on Tribolium confusum and
365	Sitophilus zeamais. Proceedings of 2 nd Int. Neem Conference,
366	Rauischholzhausen, Germany, pp. 445-451.
367	Bachrouch, O., Mediouni-Ben Jemâa, J., Chaieb, I., Talou, T., Marzouk, B.,
368	Abderraba, M., 2010. Insecticidal activity of Pistacia lentiscus essential oil on
369	Tribolium castaneum as alternative to chemical control in storage. Tunis. J. Plant
370	Prot. 5, 63-70.
371	Benelli, G., Flamini, G., Canale, A., Cioni, P.L., Molfetta, I., Conti, B., 2012.
372	Repellence of Hyptis suaveolens whole essential oil and major constituents
373	against adults of the granary weevil Sitophilus granarius. Bull. Insectol. 65, 177-
374	183.

- Benyoussef, E.H., Charchari, S., Nacer-Bey, N., Yahiaoui, N, Chakou, A.,
- Bellatreche, M., 2005. The essential oil of *Pistacia lentiscus* L. from Algeria. J.
- 377 Essent. Oil Res. 17, 642-644.
- 378 Caballero-Gallardo, K., Olivero-Verbel, J., Stashenko, E.E., 2012. Repellency and
- toxicity of essential oils from Cymbopogon martinii, Cymbopogon flexuosus and
- 380 Lippia origanoides cultivated in Colombia against Tribolium castaneum. J.
- 381 Stored Prod. Res. 50, 62-65.
- Cagri, A., Ustunol, Z., Ryser, E. T., 2004. Antimicrobial edible films and coatings. J.
- 383 Food Prot. 67, 833-848.
- Chaubey, M.K., 2012. Responses of *Tribolium castaneum* (Coleoptera:
- Tenebrionidae) and *Sitophilus oryzae* (Coleoptera: Curculionidae) against
- essential oils and pure compounds. Herba Polonica 58, 33-45.
- Conti, B., Canale, A., Cioni, P. L., Flamini, G., 2010. Repellence of essential oils
- from tropical and Mediterranean Lamiaceae against *Sitophilus zeamais*. Bull.
- 389 Insectol. 63, 197-202.
- 390 Conti, B., Canale, A., Cioni, P. L., Flamini, G., Rifici, A., 2011. Hyptis suaveolens
- and *Hyptis spicigera* (Lamiaceae) essential oils: qualitative analysis, contact
- toxicity and repellent activity against *Sitophilus granarius* (L.) (Coleoptera:
- 393 Dryophthoridae). J. Pest. Sci. 84, 219-228.
- 394 Davies, N.W., 1990. Gas chromatographic retention indices of monoterpenes and
- sesquiterpenes on methyl silicon and carbowax 20 M phases. J. Chromatography
- 396 503, 1-24.
- 397 Finney, D.J., 1971. Probit Analysis. Third ed. Cambridge University Press,
- 398 Cambridge, 32E. 57th St. NY 10022: XV + 333 pp.
- 399 Germinara, G.S., Rotundo, G., De Cristofaro, A., 2007. Repellence and fumigant

- 400 toxicity of propionic acid against adults of Sitophilus granarius (L.) and S. oryzae
- 401 (L.). J. Stored Prod. Res. 43, 229-233.
- 402 Germinara, G.S., Conte, A., Lecce, L., Di Palma, A., Del Nobile, M.A., 2010.
- 403 Propionic acid in bio-based packaging to prevent *Sitophilus granarius* (L.)
- 404 (Coleoptera, Dryophthoridae) infestation in cereal products. Innovat. Food Sci.
- 405 Emerg.Tech. 11, 498-502.
- Hou, X., Fields, P., Taylor, W., 2004. The effect of repellents on penetration into
- packaging by stored-product insects. J. Stored Prod. Res. 40, 47-54.
- Hummelbrunner, L.A., Isman, M.B., 2001. Acute, sublethal, antifeedant and
- synergistic effects of monoterpenoid, essential oil compounds on the tobacco cut
- worm Spodoptera litura (Lepidoptera: Noctuidae). J. Agric.Food Chem. 49, 715-
- 411 720.
- International Pasta Organization, 2012. Survey. The world of pasta industry in 2011.
- 413 http://www.internationalpasta.org.
- Isman, M.B., 2006. Botanical insecticides, deterrents and repellents in modern
- agriculture and an increasingly regulated world. Annu. Rev. of Entomol. 51, 45-
- 416 66.
- Karemu, C.K., Ndung'u, M.W., Githua, M., 2013. Evaluation of repellant effects of
- oils from selected *Eucalyptus* species against *Sitophilus zeamais* (Motschulsky).
- Proceedings of the First International Conference on Pesticidal Plants. 21-23
- January, Nairobi, Kenya, p. 86.
- 421 Kim, S.L, Yoon, J.S., Jung, J.W., Hong, K.B., Ahn, Y.J., Kwon, H.W., 2010. Toxicity
- and repellency of *Origanum* essential oil and its components against *Tribolium*
- 423 castaneum (Coleoptera: Tenebrionidae) adults. J. Asia-Pacific Entomol. 13, 369-
- 424 373.

- Jilani, G., Malik, M.M., 1973. Studies on neem plant as repellent against stored grain
- 426 insects. Pakistan J. Sci. Ind. R. 16, 251-254.
- 427 Lamiri, A., Lhaloui, S., Benjilali, B., Berrada, M., 2001. Insecticidal effects of
- 428 essential oils against hessian fly, *Mayeticola destructor* (Say). Field Crop Res.
- 429 71, 9-15.
- Lee, B.H., Choi, W.S., Lee, S.E., Park, B.S., 2001. Fumigant toxicity of essential oils
- and their constituent compounds towards the rice weevil, *Sitophilus oryzae* (L.).
- 432 Crop. Prot. 20, 317-320.
- Licciardello, F., Muratore, G., Suma, P., Russo, A., Nerín, C., 2013. Effectiveness of
- a novel insect-repellent food packaging incorporating essential oils against the
- red flour beetle (*Tribolium castaneum*). Innovat. Food Sci. Emerg. Tech. 19, 173-
- 436 180.
- 437 Locatelli, D.P., Süss, L., 2002. Capacità attrattiva di pasta di semola nei confronti di
- larve di *Plodia interpunctella* (Hbn.) e di adulti di *Sitophilus oryzae* (L.). Atti del
- 7° Simposio "La difesa antiparassitaria nelle industrie alimentari e la protezione
- degli alimenti", Piacenza, pp. 163-168.
- Nerio, L.S., Olivero-Verbel, J., Stashenko, E.E., 2009. Repellent activity of essential
- oils from seven aromatics plants grown in Colombia against *Sitophilus zeamais*
- Motschulsky (Coleoptera). J. Stored Prod. Res. 45, 212-214.
- Nerio, L.S., Olivero-Verbel, J., Stashenko, E., 2010. Repellent activity of essential
- oils: a review. Bioresource Technol. 101, 372-378.
- Olivero-Verbel, J., Tirado-Ballestas, I., Caballero-Gallardo, K., Stashenko, E.E.,
- 2013. Essential oils applied to the food act as repellents toward *Tribolium*
- 448 *castaneum.* J. Stored Prod. Res. 55, 145-147.
- Ojimelukwe, P.C., Alder C., 1999. Potential of zimtaldehyd, 4-allyl-anisol, linalool,

- terpineol and other phytochemicals for the control of confused flour beetle
- 451 (Tribolium confusum J. d. V.) (Coleoptera: Tenebrionidae). J. Pestic. Sci. 72, 81-
- 452 86.
- 453 Rozman, V., Kalinovic, I., Korunic, Z., 2007. Toxicity of naturally occurring
- compounds of Lamiaceae and Lauraceae to three stored product insects. J. Stored
- 455 Prod. Res. 43, 349-355.
- Shaaya, E., Kostyukovysky, M., 2006. Essential oils: potency against stored product
- insects and mode of action. Stewart Post Harvest Rev. 4, 5.
- Tapondjou, A., Adler, C., Fontem, D., Bouda, H., Reichmuth, C., 2005. Bioactivities
- of cymol and essential oils of *Cupressus sempervirens* and *Eucalyptus saligna*
- against Sitophilus zeamais Motschulsky and Tribolium confusum du Val. J.
- 461 Stored Prod. Res. 41, 91-102.
- Traboulsi, A.F., Taoubi, K., El Haj, S., Bessiere, J.M., Rammal, S., 2002. Insecticidal
- proprieties of essential plant oils against the mosquito *Culex pipens molestus*
- (Diptera: Culicidae). Pest Manag. Sci. 58, 491-495.
- Trematerra, P., Süss, L., 2006. Integrated pest management in Italian pasta factories.
- In: Proceedings of the 9th International Working Conference of Stored-product
- 467 Protection, October 2006, Campinas, San Paolo, Brazil. Brazilian Post-harvest
- 468 Association, Brazil, pp. 747-753.
- 469 UNIPI, 2012. Unione Pastai Italiani. <u>WWW.unipi-pasta.it</u>
- 470 Zeng, L., Lao, C.Z., Chen, Y.J., Liang, G.V., 2010. Study on the insecticidal activity
- 471 compounds of the essential oil from Syzygium aromaticum against stored grain
- insect pests. Proceeding of the 10th International Working Conference on Stored
- 473 Product Protection, 27 June-2 July 2010, Estoril, Portugal, pp 766-771.
- 474 Zrira, S., Elamrani, A., Benjilali, B., 2003. Chemical composition of the essential oil

4/5	of <i>Pistacia lentiscus</i> L. from Morocco - a seasonal variation. Flavour Frag. J. 18
476	475-480.
477	
478	
479	Figure legends
480	
481	Fig. 1. Repellent activity of <i>Pistacia lentiscus</i> essential oil main components (α-
482	pinene, β-caryophyllene, and α-terpineol) against Rhyzopertha dominica, Sitophilus
483	zeamais, and Tribolium confusum assessed by the two-choice pitfall bioassay. A)
484	Repellency rates. B) Percentages of individuals who remained in the arena (Non-
485	choosing Individuals). Bars indicate standard error.
486	
487	
488	
489	
490	
491	
492	
493	
494	
495	
496	
497	
498	
499	

Table 1Chemical composition of the essential oil from *Pistacia lentiscus* aerial parts used in the repellency assays

Constituents^a LRI % 941 15.9 α-pinene β-caryophyllene 1419 7 6.9 α-terpineol 1191 993 myrcene 6.4 4-terpineol 1179 5.4 β-pinene 982 5.1 germacrene D 1482 5.1 limonene 1032 4.1 α-cadinol 1654 4 δ-cadinene 1524 3.8 camphene 954 3.6 T-cadinol 1641 3.6 1028 2.8 *p*-cymene γ-terpinene 1063 1.8 tricyclene 928 1.6 sabinene 978 1.6 bornyl acetate 1287 1.6 terpinolene 1090 1.5 1-*epi*-cubenol 1628 1.4 α-humulene 1455 1.3 γ-muurolene 1476 1.2 α -muurolol 1.2 1646 α-muurolene 1499 1.1 1582 caryophyllene oxide 1.1 1020 α-terpinene 1 α-bisabolol 0.9 1684 (E,E)- α -farnesene 1507 0.7 linalool 1101 0.6 α-copaene 1377 0.5 (Z)-3-hexenyl benzoate 1570 0.5 borneol 1168 0.4 2-undecanone 1293 0.4 β-elemene 1392 0.4 *trans*-γ-cadinene 1514 0.4 spathulenol 1577 0.4 guaiol 1597 0.4 (E)-β-ocimene 1052 0.3 alloaromadendrene 1461 0.3 epi-bicyclosesquiphellandrene 1476 0.3 valencene 1492 0.3

γ-eudesmol	1631	0.3
α -phellandrene	1006	0.2
δ-elemene	1340	0.2
germacrene B	1557	0.2

^a Chemical constituents ≥ 0.1%

LRI, linear retention index on DB-5 column

Table 2Principal chemical classes in the essential oil from *Pistacia lentiscus* aerial parts used in the repellency assays

1	
Chemical classes	%
Monoterpene hydrocarbons	45.9
Oxygenated monoterpenes	14.9
Sesquiterpene hydrocarbons	22.2
Oxygenated sesquiterpene	13.3
Non-terpene derivatives	0.9
Total identified	97.8

Table 3 Repellency of the *Pistacia lentiscus* essential oil (EO) and of its main components (α -pinene, β -caryophyllene, and α -terpineol) against the pasta pests *Rhyzopertha dominica*, *Sitophilus zeamais*, and *Tribolium confusum*.

Repellent	Pest target	RD ₅₀	95 % CI	Slope ^a	Intercepta	χ2 (df)
	S. zeamais	0.010	0.022-0.920	0.677 ± 0.151	0.882 ± 0.189	2.02* (2)
P. lentiscus EO	R. dominica	0.037	0.029-0.046	1.874 ± 0.204	2.677 ± 0.285	3.63* (4)
	T. confusum	0.025	0.291-1.049	1.887 ± 0.202	0.390 ± 0.099	5.49* (3)
	S. zeamais	0.262	0.216-0.324	2.168 ± 0.239	1.260 ± 0.181	1.06* (3)
α-pinene	R. dominica	0.706	0.546-0.934	1.375 ± 0.176	0.208 ± 0.087	5.65* (4)
	T. confusum	0.225	0.170-0.337	1.908 ± 0.361	1.236 ± 0.317	2.00* (3)
	S. zeamais	0.046	0.029-0.063	1.277 ± 0.199	1.714 ± 0.232	3.70* (3)
β-caryophyllene	R. dominica	0.390	0.274-0.612	1.193 ± 0.255	0.489 ± 0.160	1.81* (2)
	T. confusum	0.153	0.090-0.282	0.746 ± 0.142	0.608 ± 0.157	0.11* (3)
	S. zeamais	0.099	0.081-0.125	2.289 ± 0.313	2.297 ± 0.356	2.07* (2)
α -terpineol	R. dominica	0.949	0.446-4.945	0.945 ± 0.140	0.021 ± 0.103	7.31* (4)
	T. confusum	0.097	0.073-0.128	1.712 ± 0.210	1.739 ± 0.236	3.43* (4)

RD₅₀, dose of repellent that repel 50 % of the exposed insects. Data for *P. lentiscus* EO are expressed as μ L cm⁻². Data for *P. lentiscus* EO components are expressed as μ M cm⁻²;

CI, Confidence Intervall;

^aValues ± standard error

χ2, chi-square; (df), degrees of freedom.

^{*} indicate P > 0.05

Table 4 Comparison of *Rhyzopertha dominica*, *Sitophilus zeamais*, and *Tribolium confusum* susceptibilities to *Pistacia lentiscus* essential oil (PEO) and its main components (α-pinene, β-caryophyllene, and α-terpineol) assessed by the area preference bioassay.

EOo	Coories	DMD	959	95% CI	
EOc	Species	RMP	from	to	
	R. dominica vs S. zeamais	2.668	0.963	12.346	
PEO	R. dominica vs T. confusum	1.649	0.639	4.982	
	S. zeamais vs T. confusum	0.618	0.157	1704	
	R. dominica vs S. zeamais	2.507*	1.691	4.052	
α-pinene	R. dominica vs T. confusum	3.317*	2.002	6.218	
	S. zeamais vs T. confusum	1.323	0.887	2.048	
	R. dominica vs S. zeamais	10.870*	4.727	35.296	
β-cariophyllene	R. dominica vs T. confusum	2.845*	1.486	6.571	
	S. zeamais vs T. confusum	0.262*	0.122	0.480	
	R. dominica vs S. zeamais	6.147*	1.972	43.387	
α-terpineol	R. dominica vs T. confusum	7.354*	2.337	52.130	
	S. zeamais vs T. confusum	1.196	0.455	3.129	

CI, Confidence Intervall

^{*} indicates values statistically significative

Table 5 Pistacia lentiscus essential oil main components repellent activity against *Rhyzopertha dominica, Sitophilus zeamais,* and *Tribolium confusum.* Adults exposed to different concentrations of α -pinene, β -cariophyllene, and α -terpineol for different exposure time (1, 3, and 24 h) in the area preference bioassay.

Species	μM cm ⁻²	Exposure (h)		% Repellency	
Species	μivi cili -	Exposure (ii)	α-pinene	β-caryophyllene	α-terpineol
		1	$30.00^a \pm 13.04$	20.00 ± 13.04	6.00 ± 4.00
	0.2	3	32.00 ± 16.25	16.00 ± 13.64	12.00 ± 5.83
		24	18.00 ± 13.19	44.00 ± 13.27	28.00 ± 9.69
		1	48.00 ± 12.81	42.00 ± 14.28	44.00 ± 10.77
R. dominica	0.4	3	46.00 ± 13.27	46.00 ± 9.27	36.00 ± 12.08
		24	44.00 ± 13.27	48.00 ± 14.97	40.00 ± 15.16
		1	28.00 ± 10.68	60.00 ± 13.78	28.00 ± 9.70
	1	3	$36.00 \pm 11.22 \text{ AB}$	$64.00 \pm 9.27 \text{ B}$	$22.00 \pm 8.00 \text{ A}$
		24	46.00 ± 12.49	68.00 ± 8.00	42.00 ± 15.62
		1	$0.00 \pm 0.00 \text{ A}$	$56.00 \pm 6.78 \; \mathrm{B}$	$30.00 \pm 11.40 \text{ B}$
	0.05	3	$0.00 \pm 0.00 \text{ A}$	$60.00 \pm 9.49 \text{ B}$	$30.00 \pm 17.61 \text{ AB}$
		24	$8.00 \pm 8.00 \text{ A}$	$60.00 \pm 5.48 \; \mathrm{B}$	$18.00 \pm 15.62 \text{ A}$
	0.1	1	32.00 ± 13.93	48.00 ± 8.00	56.00 ± 8.72
S. zeamais		3	48.00 ± 20.59	56.00 ± 14.35	64.00 ± 5.10
		24	$18.00 \pm 9.70 \text{ A}$	$72.00 \pm 6.63 \text{ B}$	$54.00 \pm 2.45 \text{ B}$
	0.2	1	28.00 ± 13.19	18.00 ± 8.00	47.00 ± 6.63
		3	46.00 ± 25.42	36.00 ± 15.68	56.00 ± 12.88
		24	36.00 ± 17.49	74.00 ± 10.30	76.00 ± 8.71
		1	$16.67 \pm 12.02 \text{ AB}$	$6.00 \pm 6.00 \text{ A}$	$46.67 \pm 8.82 \text{ B}$
	0.1	3	$26.67 \pm 14.53 \text{ AB}$	$10.00 \pm 10.00 \text{ A}$	$66.67 \pm 13.33 \text{ B}$
		24	26.67 ± 13.33	44.00 ± 19.13	63.33 ± 8.82
		1	$26.67 \pm 21.86 \text{ A}$	$4.00 \pm 2.45 \text{ A}$	$83.33 \pm 8.82 \text{ B}$
T. confusum	0.2	3	$0.00\pm0.00~A$	$0.00\pm0.00~A$	$86.67 \pm 8.82 \text{ B}$
		24	53.33 ± 13.33	40.00 ± 11.40	73.33 ± 8.82
		1	56.67 ± 18.56	63.33 ± 14.53	58.00 ± 8.00
	0.4	3	76.67 ± 23.33	63.33 ± 23.33	76.00 ± 9.27
		24	66.67 ± 3.33	60.00 ± 23.09	82.00 ± 9.70

^a Values are means \pm standard error. Values within each species and exposure time followed by different letters are significantly different by Tukey HSD test ($P \le 0.05$).

Table 6 Comparison of the repellency of *Pistacia lentiscus* essential oil main components (EOCs), α -pinene, β -caryophyllene, and α -terpineol, against the pasta pests *Rhyzopertha dominica*, *Sitophilus zeamais*, and *Tribolium confusum* assessed by the area preference bioassay.

Species	EOCs	RMP	95% CI	
Species	Locs	KWII	from	to
	α-terpineol vs α-pinene	1.105	0.546	2.126
R. dominica	α-terpineol vs β-caryophyllene	2.051*	1.005	4.764
	α-pinene vs β-caryophyllene	1.857	0.917	4.494
	α -terpineol vs α -pinene	0.389*	0.166	0.721
S. zeamais	α-terpineol vs β-caryophyllene	1.950*	1.153	3.552
	α-pinene vs β-caryophyllene	5.019*	2.544	13.417
	α -terpineol vs α -pinene	0.333*	0.097	0.856
T. confusum	α-terpineol vs β-caryophyllene	0.702	0.273	1.603
	α-pinene vs β-caryophyllene	2.109	0.865	6.106

CI, Confidence Intervall

^{*} indicates values statistically significative

Fig. 1.



