Short title: Grain legumes response to N fertilisation

NITROGEN FIXATION OF GRAIN LEGUMES DIFFERS IN RESPONSE TO NITROGEN FERTILISATION

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SUMMARY

Legume crops are not usually fertilised with mineral N. However there are at least two agronomic cases when it would be advantageous to distribute N fertiliser to legume crops: at sowing, before the onset of nodule functioning, and when a legume is intercropped with a cereal. We highlight the impact of various levels of fertiliser nitrogen on grain yield, nodulation capacity, and biological nitrogen fixation in the four most common grain legume crops grown in central Italy. Chickpea (*Cicer arietinum* L.), field bean (*Vicia faba* L. var. minor), pea (*Pisum sativum* L.), and white lupin (*Lupinus albus* L.) were grown in soil inside growth boxes for two cropping seasons with five nitrogen fertilisation rates: 0, 40, 80, 120, and 160 kg ha^{-1}. In both years experimental treatments (five crops and five levels of N) were arranged in a randomized block design. We found that unfertilised plants overall yielded grain, total biomass, and nitrogen at a similar level to plants supplied with 80-120 kg ha^{-1} of mineral nitrogen. However, above those N rates the production of chickpea, pea, and white lupin decreased, thus indicating that the high supply of N fertiliser decreased the level of N_2 fixed to such an extent that the full N_2-fixing potential might not be achieved. In all four grain legumes the amount of N_2 fixed was positively related to nodule biomass, which was inversely related to the rate of the N fertiliser applied. The four grain legumes studied responded differently to N fertilisation: in white lupin and chickpea the amount of nitrogen derived from N_2 fixation linearly decreased with increasing N supply as a result of a reduction in nodulation and N_2 fixed per unit mass of nodules. Conversely, in field bean and pea, the decrease in N_2 fixation was only due to a reduction in nodule biomass since nodule fixation activity increased with N supply. Our results suggest that the legume species and the N rate are critical factors in determining symbiotic N_2-fixation responses to N fertilisation.

Keywords: BNF, grain legumes, N fertilisation, nodule, root.

INTRODUCTION

Legumes are key components of sustainable cropping systems. This is because they may access atmospheric N_2 through a symbiotic relationship in their root system with a group of soil-borne bacteria collectively called rhizobia, which results in the development of specialized organs called nodules. The symbiotic microorganisms in the root nodules take up gaseous di-nitrogen from the air and fix nitrogen into ammonia or amino acids. The N_2 can then be assimilated by the host plant, which in turn provides carbon resources to the rhizobia. Symbiotic N_2 fixation in legumes is not obligatory for the survival of the host plant as they can use mineral N in soil (Namvar and Sharifi, 2011; Voisin *et al*. 2002a, 2002b).

A number of reviews have been published on biological nitrogen fixation (BNF) in legumes (Cheema and Ahmad, 2000; Salvagiotti *et al*., 2008; Van Kessel and Hartley, 2000). Most authors agree that an increase in the concentration of combined N in soil decreases nodule establishment, legume nodulation activity, and N_2 fixation; thus legume crops are not usually fertilised with mineral N. However, there are at least two agronomic cases when it would be advantageous to distribute N fertiliser to legume crops. The first is at sowing, before the onset of nodule functioning, when young legume plants require N from external sources in order to achieve proper vegetative growth and the N_2-fixing symbiosis. In this situation amounts of N fertiliser of about 50 kg ha^{-1}, defined as "starter N", have been proved to be beneficial to plant development and subsequent nodulation (Namvar and Sharifi, 2011; Van Kessel and Hartley, 2000). The second case is when a legume is intercropped with a cereal. Given that the N transfer from the legume crop to companion intercropped species is very low (Mariotti *et al*., 2012; Pirhofer-Walzl *et al*., 2012), a rate of N fertiliser of about 80 kg ha^{-1} is required for the intercropping in order to sustain the cereal's high yield (Ghaley *et al*., 2005).

Only a few studies have attempted to establish a quantitative relationship between grain yield or N_2 fixation and the N fertiliser rate in grain legume crops grown in soil (Salon *et al*., 2001; Voisin *et al*., 2002b). According to Streeter (1988) the vast majority of studies have been carried out in controlled environments with nutrient solutions supplied to inert solids, and most experiments have been conducted to verify whether soil N can inhibit the formation and development of nodules or nitrogen fixation. Moreover many experiments only evaluated the starter N effect, thus scheduling very low N rates and making observations only for a short time after sowing (Jensen, 1986, 1987; Voisin *et al*., 2002b). Nitrogen rates higher than 50 kg ha^{-1} applied at sowing, usually decreased the BNF of grain legume crops (Voisin *et al*., 2002b) while rarely affected grain yield (Clayton *et al*., 2004; Voisin *et al*., 2002a, 2002b). The stimulating effect on legume BNF at relatively low levels of soil mineral N at sowing should be distinguished from the inhibition of legume BNF by high levels of soil mineral N, and declines in BNF should also be distinguished from grain yield reduction.

In this research we hypothesized that in grain legume crops: i) BNF is positively related to nodule mass, ii) nodule mass is negatively related to N fertilisation, and iii) previous relationships differ among legume crops. Thus, we highlight the impact of various levels of fertiliser nitrogen applied at sowing time on grain yield, nodulation capacity, and biological nitrogen fixation in the four most common grain legume crops grown in central Italy: chickpea, field bean, pea, and white lupin. The experiment was carried out in growth boxes in order to measure the entire root system and nodule biomass.

MATERIALS AND METHODS

Site characteristics and experimental design

The research was carried out in two consecutive years, 2011 and 2012, at the Research Centre of the Department of Agriculture, Food and Environment of the University of Pisa, Italy, which is located at a distance of approximately 5 km from the sea (43°40′ N, 10°19′ E) and 1 m above sea level. The climate of the area is hot-summer Mediterranean (Csa) with mean annual maximum and minimum daily air temperatures of 20.2°C and 9.5°C respectively, and a mean rainfall of 971 mm per year.

In both years, experimental treatments consisted of five crops (four legume crops plus durum wheat) and five levels of mineral nitrogen fertilisation, arranged in a randomized block design. Three replications were used. The four legumes were chickpea (*Cicer arietinum* L. cv. Pascia), field bean (*Vicia faba* L. var. minor cv. Chiaro di Torrelama), pea (*Pisum sativum* L. cv. Iceberg), and white lupin (*Lupinus albus* L. cv. Multitalia). Durum wheat (*Triticum durum* L. cv. Claudio) was used as non N_2-fixing reference crop in order to determine plant-available soil nitrogen and estimate BNF. Applied N rates were 0 kg ha^{-1} (N0), 40 kg ha^{-1} (N40), 80 kg ha^{-1} (N80), 120 kg ha^{-1} (N120), and 160 kg ha^{-1} (N160). Nitrogen was applied pre-planting as urea and deep placed at 10 cm. Legume crops and durum wheat were supplied with the same amounts of fertiliser and were grown exactly under the same conditions.

Experimental equipment and crop management

In each year, the open-air facility consisted of 75 growth boxes (15 per species) of 200-L volume (0.25 m^2 area and 0.8 m depth), spaced 20 cm apart, and embedded in expanded clay to avoid daily fluctuations in soil temperature. In both growing seasons, approximately six months before seeding, growth boxes were filled with soil collected from a field previously cultivated with rapeseed (*Brassica napus* L.). The main properties of the soil before N fertiliser application were similar in the two years and were approximately: 71.0% sand, 23.7% silt, 5.3% clay (USDA method), 8.1 pH, 1.5% organic matter (Walkley and Black method), 0.6 g kg^{-1} total nitrogen (Kjeldahl method), 11.9 mg kg^{-1} available P (Olsen method), 122.1 mg kg^{-1} available K ($BaCl_2$-TEA method), 1.9 mg kg^{-1} soil mineral N (NO_3-N and NH_4-N) concentration (potentiometric method after extraction with 2M KCI and filtration). The soil pH was in the range of basic tolerance of all four legumes (Jayasundara *et al*., 1998) and durum wheat (Westerman, 1987).

Both legumes and durum wheat were grown following a standard technique for central Italy, with the exception of nitrogen fertilisation. Phosphorus was applied pre-planting as triple superphosphate at the rate of 150 kg ha^{-1} of P_2O_5 for all the crops. Potassium was also applied pre-planting as potassium sulphate at the rate of 150 kg $ha^{-1}$of K_2O and 54 kg ha^{-1} of S for all the crops. The legumes and durum wheat were sown on 11 February 2011 and on 14 February 2012, within the optimum planting time for spring legume production in central Italy. Legume seeds were inoculated just prior to sowing with a specific commercial rhizobial inoculant using *Rhizobium leguminosarum* bv. *viciae* for field bean and pea, *Mesorhizobium ciceri* for chickpea, and *Bradyrhizobium sp. (Lupinus)* for white lupin. In both years, three weeks after sowing, chickpea was thinned to 32 plants m^{-2}, field bean and pea to 56 plants m^{-2}, and white lupin to 40 plants m^{-2}. Row spacing was 30 cm for all the crops. Durum wheat was sown at a rate of 400 germinable seeds m^{-2} with a 15-cm row spacing and was not thinned. In both years all crops were irrigated from flowering to maturity (May to June). In this period 100 mm of irrigation water was applied and 40 mm in 2011 and 60 mm in 2012 came from rainfall. Weed control was performed throughout the two crop cycles by hand hoeing.

Sampling procedures and measurements

All five crops were harvested at physiological maturity: 24 June for field bean and pea, 5 July for chickpea, white lupin and durum wheat in 2011; and 22 June for pea, 26 June for field bean, 4 July for chickpea and durum wheat and 9 July for white lupin in 2012. Plants were cut at ground level and partitioned into seeds, pod-walls or chaff, stems+leaves, taproots, rootlets, and nodules. Roots were separated from the soil by gently washing to minimise loss or damage by a low flow from sprinklers. One sample of roots was stored in a refrigerator until the length of the roots was measured, which was estimated with the line intersection method (Tennant, 1975). Dry weight of all plant parts was determined by oven-drying at 60° C to constant weight. The number of pods or spikes was recorded and mean seed weight, harvest index and shoot/root ratio were determined. All plant parts were analysed for N concentration by the microKjeldahl method. Nitrogen content was obtained by multiplying N concentrations by dry matter of different plant parts.

The amount of N fixed was estimated with the improved N difference method, as proposed by Evans and Taylor (1987): [total N content in legume crop – total N content in reference crop] + [soil mineral N in legume crop at harvest – soil mineral N in reference crop at harvest]. Durum wheat was grown as the non N_2-fixing reference crop. The non N_2-fixing reference crop should be: i) a non-legume; ii) a non-nodulating legume of the same species as the N_2-fixing plant; or iii) an uninoculated legume in a system without a background population of compatible rhizobia. Ideally, the non N_2-fixing and N_2-fixing plants would be of the same species. In practice it is difficult to prevent contamination with rhizobia and infection of plants, especially in soils, and so non N_2-fixing species are more commonly used (Danso, 1995, Unkovich, 2008; Peoples *et al*., 2009; Ashworth *et al*., 2015). In order to estimate N_2-fixation in cool season grain legumes, the non-legume species barley and wheat are the more suitable reference crops (Henson, 1993; Kadiata *et al*., 2012; López-Bellido *et al*., 2006 and 2011; Neugschwandtner *et al*., 2015; Unkovich, 2008).

The nodule fixation activity (NFA) is the amount of N_2 fixed per unit mass of nodules and was calculated at harvest as: N_2 fixed (g m^{-2}) / nodule dry weight (g m^{-2}).

Weather conditions

Daily minimum and maximum temperatures, rainfall, and reference evapotranspiration during both growing seasons were obtained from a meteorological station located within 100 m from the trial site. Accumulated growth season rainfall in 2011 and 2012 was 283 mm and 284 mm respectively, both below the 20-year average of 322 mm. Rainfall was concentrated in February-March in 2011 and in April in 2012. The average maximum and minimum temperatures for the growing seasons were 21.6°C and 8.7°C in 2011 and 21.5°C and 7.9°C in 2012. Maximum and minimum temperatures did not differ from the 20-year average for the area and were similar in the two years, the only exception being the lower temperatures in February 2012. Accumulated reference evapotranspiration was similar in the two years (425 mm in 2011 and 396 mm in 2012) and did not differ from the 20-year average.

Statistical analysis

Results were subjected to analysis of variance. The effect of year, crop, and N rate, and their interactions were analysed using a split-split-plot design with year designed as whole plots, crop as sub-plots, and N rate as sub-sub-plots. Significantly different means were separated at the 0.05 probability level by the least significant difference test (Steel *et al*., 1997).

RESULTS

Analysis of variance revealed non-significant effects of years or “Year x Crop x N rate” interaction, “Year x N rate” interaction, “Year x Crop” interaction for all the parameters measured. Accordingly, the following results are averaged over the two years.

Above ground biomass

Biomass differed greatly among the four legume crops owing to their morphological and physiological features. Without N fertilisation, grain yield of field bean was 16% higher than pea, 64% higher than chickpea, and 102% higher than white lupin, while straw of field bean was 38% higher than pea and 19% higher than chickpea but 4% lower than white lupin (Table 1).

Nitrogen fertilisation did not modify the grain yield of field bean, while the highest N rate decreased the grain yields of white lupin (-27%), chickpea (-16%), and pea (-22%). The grain yield reduction was due to a lower number of seeds per square meter in chickpea, to a lower mean seed weight in white lupin and to both in pea (Table 1). Nitrogen fertilisation did not modify the straw of chickpea and pea, while the highest N rate decreased the straw of field bean (-12%) and white lupin (-23%) (Table 1). The harvest index (Table 1) was unaffected by N supply in field bean and white lupin, and was reduced by the highest N rate in chickpea (-17%) and in pea (-11%).

Nitrogen fertilisation progressively increased grain yield and straw of wheat and at the highest N rate grain yield and straw were 74% and 127% respectively higher than control (Table 1). The grain yield increase in wheat was mainly due to increased seed number (Table 1).

Root system

Without N fertilisation, the dry weight of field bean roots (347 g m^{-2}) was 169% higher than pea, 95% higher than chickpea, and 54% higher than white lupin (Figure 1). Nitrogen fertilisation did not modify root biomass in pea and increased that of field bean, chickpea, and white lupin up to N80, and thereafter values decreased. With the highest N rate, root biomass was 25% lower than the control in field bean and in white lupin and equal in chickpea. Overall differences were due to the rootlets, since taproots were not affected by N supply in any of the crops. Taproot biomass was negligible in pea and chickpea, and accounted for 11% and 21% of the total root biomass respectively in field bean and white lupin, irrespective of the N supply.

Without N fertilisation, roots were 21% of the total plant biomass in field bean, 18% in white lupin, 16% in chickpea, and 11% in pea. The root/shoot ratio was not modified by N fertilisation in chickpea and pea, while in the other two crops it increased up to N80 and then decreased.

When no N fertiliser was added, the length of field bean roots (4.2 km m^{-2}) was by 75% higher than pea, 253% higher than chickpea, and 268% higher than white lupin (Figure 1). Roots were lengthened by N fertilisation up to N40 in pea and white lupin (+35% and

+58% respectively) and up to N80 in chickpea and in field bean (+42% and +86% respectively). At higher N supply root length decreased, so that with the highest N rate it was slightly lower than the unfertilised control in field bean and pea (-9% and -15%) and was unchanged in chickpea and white lupin.

Root biomass and length of wheat were unchanged by N fertilisation (Figure 1).

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Nodule biomass

When no N fertiliser was added, nodule biomass of pea (18.6 g m^{-2}) was 33% higher than chickpea, 52% higher than field bean, and 86% higher than white lupin. The nodule mass of the four legumes was inversely related to the levels of the N fertiliser applied (Figure 2). The reduction rate differed among legume crops and each kg of applied N decreased the nodule biomass by 30 mg m^{-2} in white lupin, 40 mg m^{-2} in chickpea, 50 mg m^{-2} in field bean, and 90 mg m^{-2} in pea. Accordingly, with 160 kg N ha^{-1} nodule biomass of chickpea was approximately twice that of field bean, pea, and white lupin. When no N fertiliser was added, nodule biomass accounted for 4% of total root biomass in field bean and white lupin, for 8% in chickpea, and for 14% in pea. With the highest N rate nodule biomass declined to less than 4% in all the four crops.

Nitrogen concentration and content

Nitrogen concentrations of grain and straw were not affected by N fertilisation. Considering averages over the N rates, grain N concentration of field bean and white lupin (4.5 g kg^{-1}) was higher than that of chickpea and pea (3.5 g kg^{-1}), while straw N concentration of field bean and pea (1.6 g kg^{-1}) was higher than that of chickpea and white lupin (1.0 g kg^{-1}). The N concentration of roots and nodules was not affected by N rates and was similar among crops averaging 1.2% and 3.4%, respectively.

Nitrogen fertilisation did not modify the grain N content of field bean, while the highest N rate decreased the grain N content of chickpea, pea, and white lupin by approximately 20% (Figure 3). Straw N content of chickpea, pea, and white lupin was unchanged by N fertilisation, while that of field bean decreased with all N rates applied. The N content of roots was the highest with N80 in field bean, pea, and white lupin, while in chickpea the N content was not modified by N supply. When no N fertiliser was added, the nitrogen content of nodules was less than 0.6 g m^{-2} with slight differences among crops and among N rates, and depending on dry matter variations, decreased with increasing N rates (Figure 3). The amount of N uptake by durum wheat (reference crop) progressively increased with N supply from 7.5 to 13.7 g m^{-2} (Figure 3).

Without N fertilisation, total N content of field bean was 58% higher than pea, 72% higher than chickpea, and 79% higher than white lupin. Nitrogen fertilisation did not statistically change total N content of field bean while it decreased that of chickpea and white lupin with N rates higher than 80 kg ha^{-1} and that of pea with the highest supply. As N rate increased from 0 to 160 kg ha^{-1}, the total N content of chickpea, pea and white lupin decreased by approximately 15%.

Nitrogen fertilisation progressively increased grain and straw N content of wheat and at the highest N rate grain yield was 81% higher than control and straw was 133% (Table 1). Nitrogen content of roots was unchanged by N fertilisation.

Nitrogen fixation

When no N fertiliser was added, the amount of N_2 fixation in field bean reached 31.1 g m^{-2} and was approximately twice that of the other three legume crops. Nitrogen fertilisation significantly influenced the amounts of N_2 fixed by all legume crops, and a negative linear relationship was observed between N fertiliser rate and N_2 fixation (Figure 4). However the reduction rate differed among crops and each kg of applied N decreased N_2 fixed by 50 mg N m^{-2} in field bean and by approximately 60 mg N m^{-2} in chickpea, pea, and white lupin. Because of the linear decline, the increasing N supply from 0 to 160 kg ha^{-1} reduced the amount of N_2 fixed by only 27% in field bean but up to 60-69% in chickpea, pea, and white lupin.

For each crop, the amount of N_2 fixed was highly correlated with nodule mass (Figure 5). The increase in N_2 fixed per gram of nodule dry weight was 1.8 g m^{-2} in white lupin, 1.3 g m^{-2} in chickpea, 1.0 g m^{-2} in field bean, and 0.8 g m^{-2} in pea.

When no N fertiliser was added, N_2 fixation accounted for 81% of total N in field bean and approximately 67% in chickpea, pea, and white lupin. In all the four crops, the proportion of fixed N also linearly decreased with increasing N-fertiliser additions (Figure 6). However, once again, the decrease differed among species. In chickpea, pea, and white lupin (about 0.25% per kg of N applied) the proportion was twice as high as in field bean. With 160 kg N ha^{-1} N_2 fixation accounted for 62% in field bean, but only for 33% in pea, 29% in chickpea, and 24% in white lupin.

Regression analysis for NFA against N rate indicated highly significant relation in all the four legume crops (Figure 7). However, NFA increased in field bean and pea with the increasing N supply, while it decreased in chickpea and white lupin. Each kg of N applied with fertilisation increased the amount of N_2 fixed per gram nodule by 16 mg in field bean and by 4 mg in pea, and decreased those of chickpea and white lupin respectively by 3 and 7 mg.

DISCUSSION

Aerial biomass

We found that well-nodulated legumes (non-fertilised controls) overall grew and yielded grain, total biomass, and nitrogen at a similar level to plants supplied with 80-120 kg ha^{-1} of mineral nitrogen. These findings highlighted that symbiotic nitrogen fixation and root mineral N absorption are complementary up to a certain N supply, and within this range of N levels plants substituted with N from the fertiliser the amount of nitrogen they ordinarily would have derived from biological fixation. Similar results were previously reported by Deibert *et al*. (1979), Sagan *et al*. (1993), and Voisin *et al*. (2002a, 2002b), who found that biomass, nitrogen accumulation and seed yield were not affected by mineral N applications. However, above these N rates (80-120 kg N ha^{-1}) the biomass and N yield of chickpea, pea, and white lupin decreased, thus indicating that high N fertiliser supply decreased the level of N_2 fixed to such an extent that the full N_2-fixing potential might not have been achieved. Our results partially support the hypothesis of Lemaire *et al*. (1997), who stated that relationships between N and growth would be unchanged by the N nutrition regime.

Root biomass

In greenhouse trials Arrese-Igor *et al*. (1997) and Schulze *et al*. (1999) showed that legumes grown with mineral N usually have a more developed root system than strictly fixing plants. They hypothesized that the nitrate supply can have a considerable impact on carbohydrate partitioning, leading to enhanced root development thus providing an increased absorption surface. Thus, differences in carbon costs between symbiotic nitrogen fixation and nitrate absorption could be incurred by growth and/or maintenance of the nodulated roots. As such, the presence of mineral N in the soil can lead to higher root biomass through the limitation of BNF and its associated high C costs. However, under field conditions, Gunawardena *et al*. (1998) and Jensen (1987) did not find any difference in pea roots due to N fertilisation and Jensen (1986) and Voisin *et al*. (2002a) reported a positive effect of mineral N on root growth but only with N rates lower than 100 kg ha^{-1}, while no variation was found with higher rates. We found that the dry weight of pea roots progressively increased with the increase in N rate, while those of chickpea, field bean, and white lupin increased up to 80 kg ha^{-1} and thereafter decreased. Similarly, in all four crops root length increased up to N40 - N80, and thereafter decreased. Thus, upon a certain N concentration in the soil, plants may not have needed to lengthen their roots to absorb nitrogen.

Nodule biomass

Nodule mass of all four crops was inversely related to the levels of the N fertiliser applied. However, nitrogen fertilisation reduced the nodule mass of field bean and pea more than that of chickpea and white lupin (two fold). Voisin *et al*. (2003) reported that nodule growth was not affected by N source and the negative effect of nitrate on nodule mass might only result from the delayed onset of nodules. Unfortunately, we did not measure the number of nodules per plant thus it is not possible to know whether the reduced nodule mass was due to a delay in nodule initiation; however, we assumed that when soil N was sufficiently depleted by plant uptake, nodule formation, development or function could be reinstated. Thus, the high N uptake of field bean depleted the N soil content in less time than other legumes, and *Rhizobia* were able to restart their infection at an early stage and nodule growth and N_2 fixation were thus able to start again.

Nitrogen fixation

The two most commonly used methods for estimating N_2 fixation across the growing season are ^{15}N-isotope dilution and N difference. Reviewing the literature on BNF determination, Unkovich and Pate (2000) noted that the N difference method is less accurate than the ^{15}N-isotope method. However, according to Herridge *et al*. (2008) and Müller and Thorup-Kristensen (2002), the two methods deliver the same results when comparing BNF with different treatments. Ashworth *et al*. (2015) concluded that the N-difference method could be used instead of the ^{15}N-isotope method when precise values are not necessary. Therefore we think that for our research purposes the N-difference method would be profitably utilized.

Nitrogen fertilisation linearly decreased the amount of N_2 fixed by all four grain legumes with a slope ranging from 50 mg N m^{-2} per kg of applied N for field bean to 70 mg N m^{-2} for the other three crops. Thus, N supply affected N_2 fixation of the four legume crops differently, and was more damaging for chickpea, pea, and white lupin than for field bean, indicating that field bean rhizobia were the most tolerant to high soil mineral N concentrations. Similarly, Evans *et al.* (1989), Rennie and Dubez (1986) and Turpin *et al.* (2002) found a decrease in N_2 fixation due to N fertilisation and the advantage of field bean in N fertilisation reactions. Nitrogen fertilisation also linearly decreased the plant dependence on bacterial N_2 fixation but did not completely inhibit it. All four species continued to fix N_2 even when the N rate was up to 160 kg ha^{-1}, although with this N supply, N_2 fixation accounted for almost two thirds of total N uptake in field bean, but only for a quarter in chickpea, pea, and white lupin. At harvest, soil mineral N concentration was approximately 1.7 mg kg^{-1} without appreciable differences among wheat and legume crops and N rates. This confirms that both legume crops and durum wheat, whether fertilized or not with N, take up practically all the available soil N irrespectively of N fertiliser supply (Jensen, 1997).

Estimates of N_2 fixation have usually been based solely on measurements of above-ground plant biomass, thus both N uptake and N_2 fixation have often been underestimated since N in roots and nodules were not taken into account (Salvagiotti *et al*., 2008; Unkovich and Pate, 2000). However, we found that only slightly more than 10% of total N in chickpea, field bean, and white lupin, and slightly less than 10% in pea were stored in roots and nodules at maturity, irrespective of N supply. These values are lower than those reported by Unkovich and Pate (2000) for chickpea and white lupin (28-40%), which were measured at mid-flowering stage and therefore without the grain supply to total N content.

In all four grain legumes the amount of N_2-fixed was positively related to nodule mass which was inversely related to the levels of the N fertiliser applied. Thus, in all four crops N_2 fixation was reduced by depression of nodulation growth resulting from increasing in N fertilisation. Streeter (1988) proposed that N_2 fixed per unit nodule mass decreases progressively with the increase in medium nitrate concentration. In our research the amount of N_2 fixed per unit of nodule mass was linearly related to the N rate in all four legume crops. However, with an increasing N supply, nodules of field bean and pea appeared to intensify their NFA, while those of chickpea and white lupin appeared to reduce their activity. To the best of our knowledge no research was carried out to compare nodule fixation activity among *Rhizobium* types, an issue that would explain the differential NFA response to N supply among species.

All summarizing, we found that N fertilisation reduced N_2 fixation of field bean and pea by reducing nodule mass, and reduced N_2 fixation of chickpea and white lupin by reducing both dry matter and nitrogen fixation activity of the nodules. These findings were in accordance with Streeter (1988), who reported that N fertilisation can reduce N_2 fixation by i) inhibiting the infection and depression of nodulation growth, which results in a reduction in nodule mass per plant, or ii) inhibiting the nitrogenase activity per unit mass of nodule, corresponding to the amount of N_2-fixed per unit mass of nodules. *Rhizobium leguminosarum* bv. *viciae,* used for field bean and pea, seems to be more tolerant to high levels of combined N than *Mesorhizobium ciceri* and *Bradyrhizobium sp. (Lupinus)*. In addition, each kg of applied N reduced nodule biomass and N_2 fixed of pea by 1.3 fold and 1.7 fold respectively compared to field bean. Accordingly, different *Rhizobium* strains differ in their ability to induce nodulation and fix nitrogen and crop species differ in their susceptibility to nodulation. Thus, the nitrate inhibition would seem to be primarily host plant dependent as hypothesized by Cheema and Ahmad (2000) and Ohyama *et al*. (2011).

CONCLUSIONS

We found a negative relationship between N fertilisation rate and nodulation as well as N_2 fixation in grain legume crops. However with N rates lower than 120 kg ha^{-1} reductions in nodulation and N_2 fixation had no effect on above ground growth and grain yield. Above this N rate biomass production decreased, thus indicating that the high rates of N fertiliser decreased the level of N_2 fixed to such an extent that the full N_2-fixing potential might not have been achieved. We assumed that when soil mineral N was sufficiently depleted by plant uptake, nodule formation, development or function could be reinstated.

Our findings indicated that the N_2-fixing symbiotic relationships between plants and bacteria do not respond to N fertilisation rate in the same manner across species. As *Rhizobium* strains likely differ in their ability to induce nodulation and fix nitrogen, crop species differed in their nodulation susceptibility to fertilisation. Mineral-N inhibition would thus seem to be primarily host-plant dependent. Further research is needed to determine the best N rate for cereal/legume intercropping and the most suitable phenological phase to perform N fertilisation.

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Table 1. Grain and straw dry matter, harvest index, mean seed weight (MSW), and seed number as affected by "Crop x N rate" interaction. Values followed by different letters within column are significantly different ($P < 0.05$).

Crop	N rate	Dry matter				Seed number
		Grain	Straw	Harvest Index	MSW	
	kg ha^{-1}	g m^{-2}	g m^{-2}	%	mg	n m^{-2}
Chickpea	0	328.0 fg	610.6 efg	34.9 d	357.4 cd	917.8 m
	40	327.7 fgh	646.7 cdef	33.6 def	366.0 c	895.4 mn
	80	327.8 fg	630.1 def	34.2 de	401.9 a	815.6 mn
	120	285.9 ghi	644.5 cdef	30.7 defg	390.0 ab	733.2 mn
	160	276.4 hi	678.5 bcd	28.9 fgh	394.0 a	701.4 n
Field bean	0	537.5 a	729.6 ab	42.4 bc	330.6 e	1625.7 h
	40	562.2 a	705.1 abc	44.4 abc	355.8 cd	1580.4 hi
	80	539.2 a	650.9 cdef	45.3 abc	336.8 de	1601.3 h
	120	540.2 a	664.1 bcde	44.9 abc	369.0 bc	1464.0 hi
	160	510.7 a	642.7 cdef	44.3 abc	352.4 cde	1449.3 i
Pea	0	463.0 abc	527.7 hi	46.7 ab	158.5 h	2921.1 f
	40	436.8 bc	507.3 i	46.3 abc	153.6 hg	2843.1 fg
	80	465.5 abc	544.4 ghi	46.1 abc	148.3 hg	3139.9 e
	120	428.3 cd	548.3 ghi	43.9 abc	146.5 hg	2923.7 ef
	160	362.3 ef	513.4 i	41.4 c	135.0 g	2683.8 g
White Lupin	0	266.1 i	761.7 a	25.9 gh	212.4 fg	1252.8 il
	40	283.5 ghi	681.9 bcd	29.4 efgh	235.5 f	1203.8 l
	80	264.3 i	657.1 cde	28.7 fgh	218.1 fg	1211.5 l
	120	235.9 il	684.5 bcd	25.6 gh	209.7 g	1124.7 l
	160	195.6 l	586.9 fgh	25.0 h	165.6 h	1181.4 l
Durum wheat	0	279.9 ghi	296.2 l	48.6 a	40.7 i	6874.2 d
	40	380.2 de	480.9 i	44.2 abc	41.4 i	9190.8 c
	80	440.6 bc	609.4 efg	42.0 bc	41.7 i	10556.7 b
	120	478.0 abc	662.7 bcde	41.9 bc	42.5 i	11255.4 a
	160	486.4 ab	673.0 bcde	42.0 bc	43.2 i	11249.7 a

Figure captions

Figure 1. Root dry weight (a) and root length (b) as affected by “Crop x N rate” interaction. Vertical bars indicate LSD at $P<0.05$. Open circles, chickpea; solid circles, field bean; open squares, pea; solid squares, white lupin; open triangles, durum wheat.

Figure 2. Relationship between nodule dry weight and N rate in chickpea (a), field bean (b), pea (c), and white lupin (d). Vertical bars indicate standard error.

Figure 3. Nitrogen content of grain (a), straw (b), roots (c), and nodules (d) as affected by “Crop x N rate” interaction. Vertical bars indicate LSD at $P<0.05$. Open circles, chickpea; solid circles, field bean; open squares, pea; solid squares, white lupin; open triangles, durum wheat.

Figure 4. Relationship between nitrogen fixed and N rate in chickpea (a), field bean (b), pea (c), and white lupin (d). Vertical bars indicate standard error.

Figure 5. Relationship between nitrogen fixed and nodule dry weight in chickpea (a), field bean (b), pea (c), and white lupin (d). Data from two years, five N rates and three replications.

Figure 6. Relationship between percentage of nitrogen fixed on total N and N rate in chickpea (a), field bean (b), pea (c), and white lupin (d). Vertical bars indicate standard error.

Figure 7. Relationship between nodule fixation activity (NFA) and N rate in chickpea (a), field bean (b), pea (c), and white lupin (d). Vertical bars indicate standard error.

Figure 1

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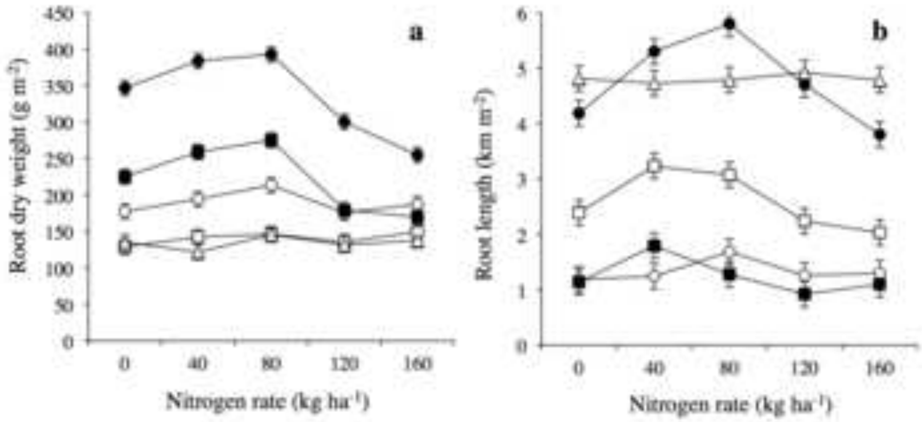

Figure 2

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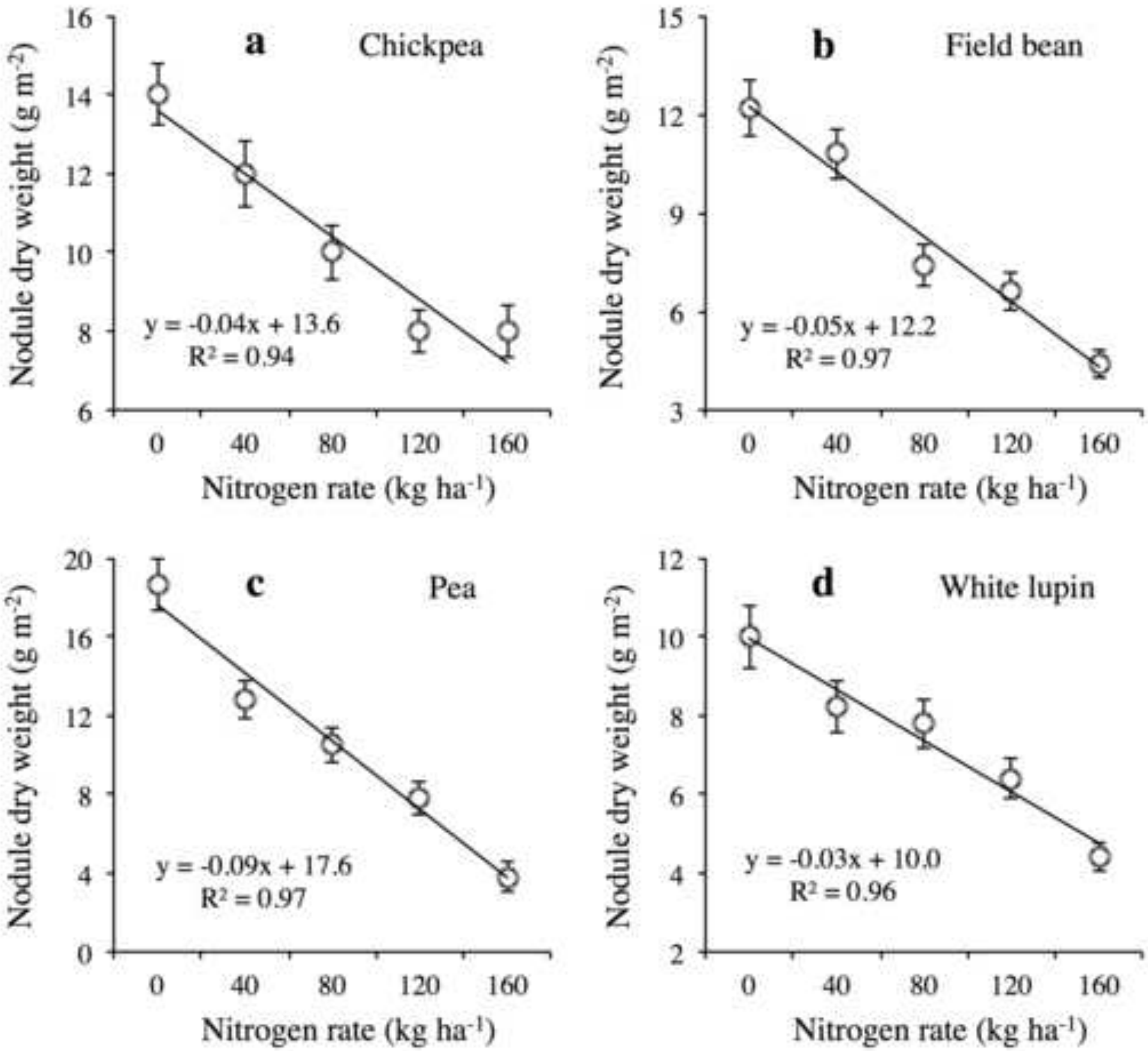

Figure 3

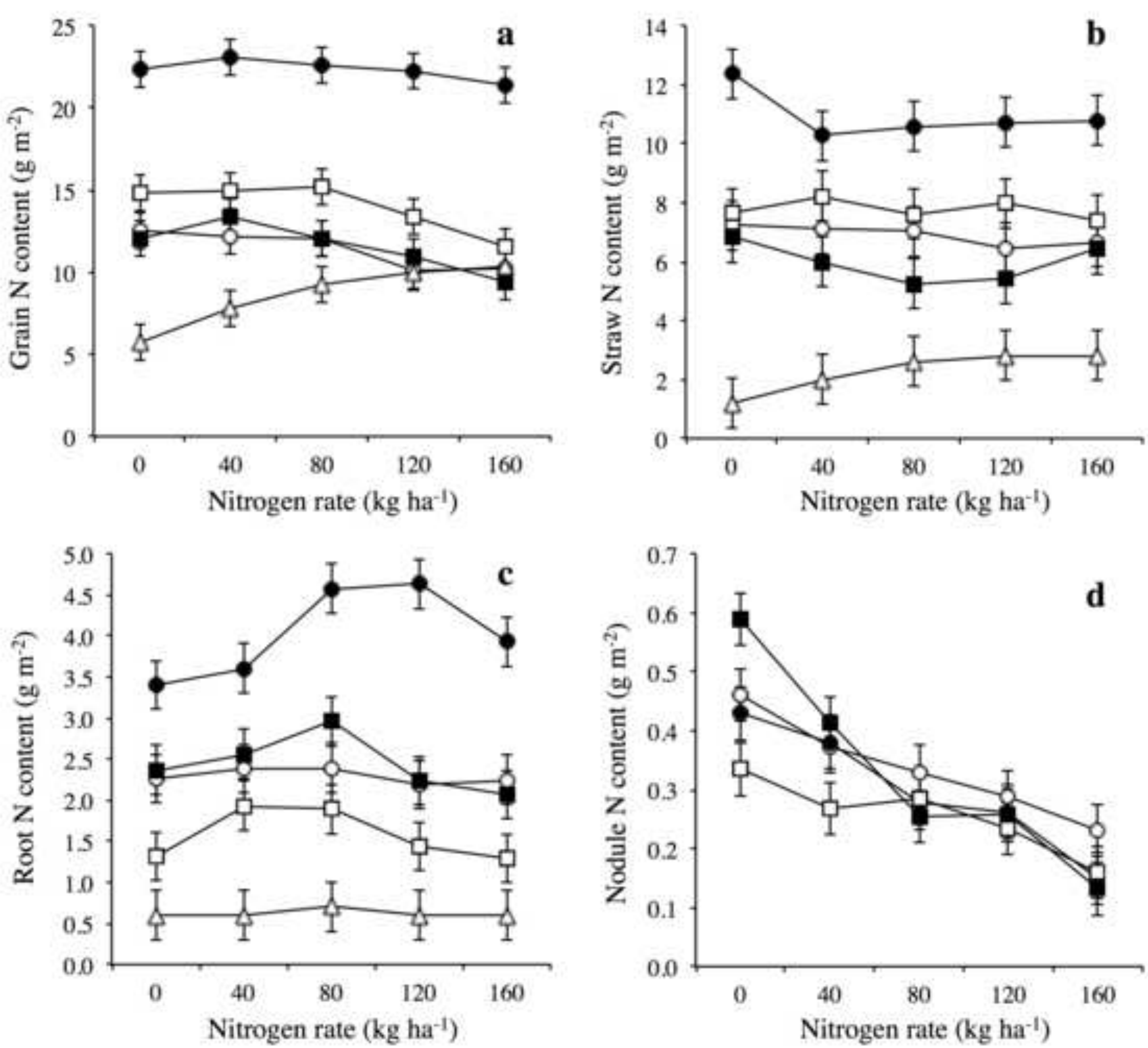
a
Grain N content (g m-2)
25
20
15
10
5
0
0
40
80
120
160
Nitrogen rate (kg ha-1)
b
Straw N content (g m-2)
14
12
10
8
6
4
2
0
0
40
80
120
160
Nitrogen rate (kg ha-1)
c
Root N content (g m-2)
5.0
4.5
4.0
3.5
3.0
2.5
2.0
1.5
1.0
0.5
0.0
0
40
80
120
160
Nitrogen rate (kg ha-1)
d
Nodule N content (g m-2)
0.7
0.6
0.5
0.4
0.3
0.2
0.1
0.0
0
40
80
120
160
Nitrogen rate (kg ha-1)

Figure 4

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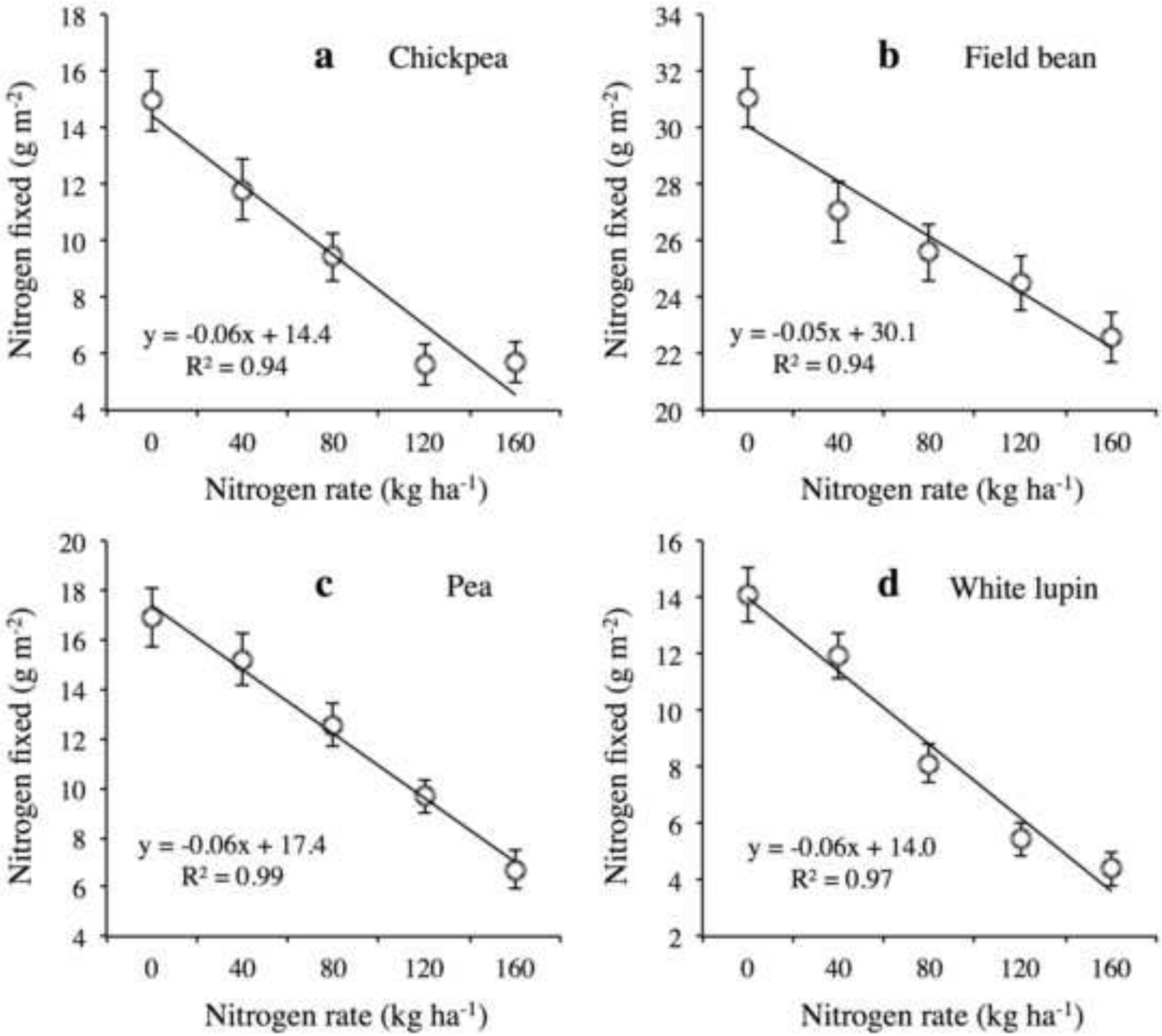

Figure 5

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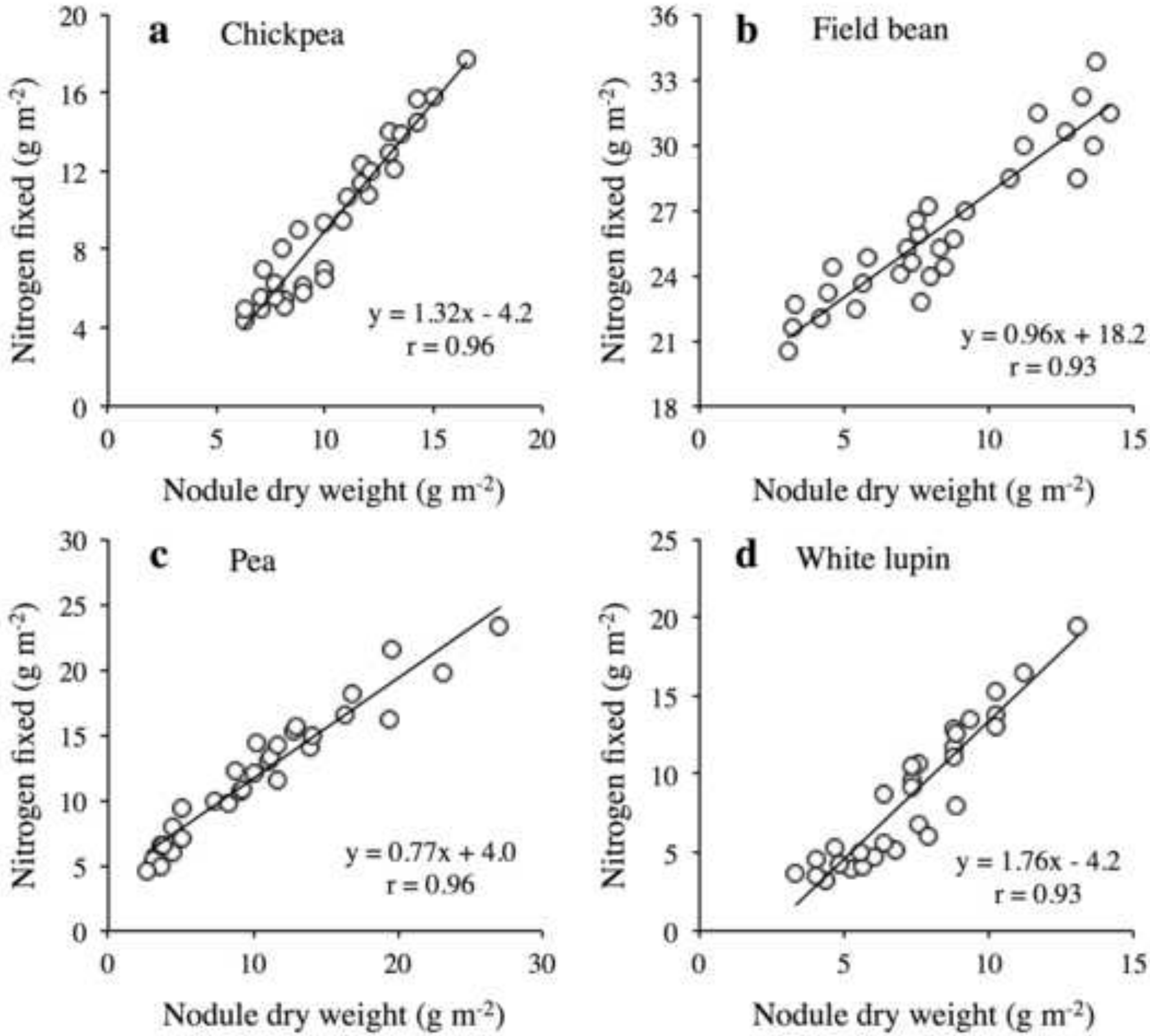

Figure 6

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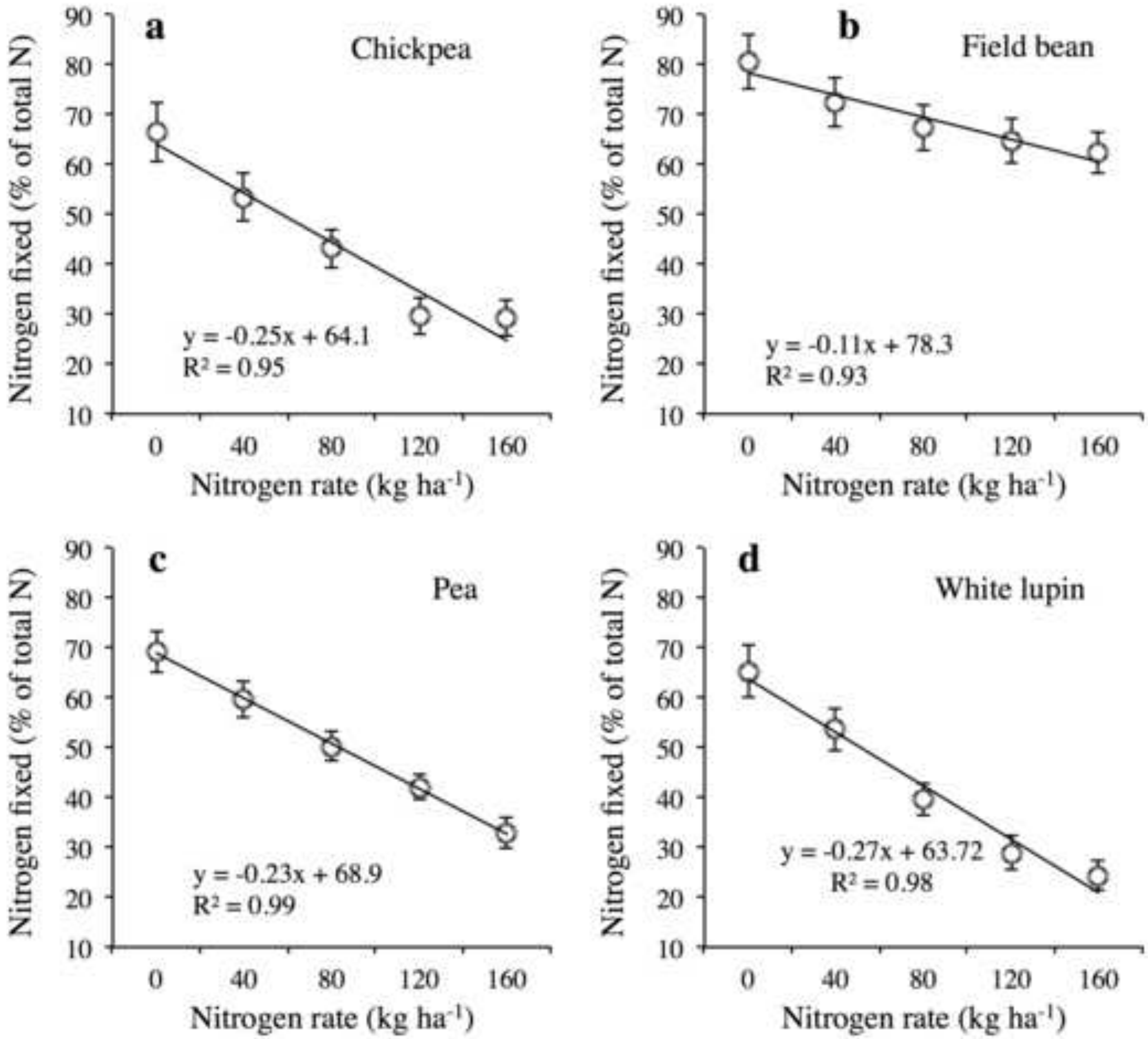

Figure 7

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