

Review Article

IL-6 as a Druggable Target in Psoriasis: Focus on Pustular Variants

Andrea Saggini,¹ Sergio Chimenti,¹ and Andrea Chiricozzi^{1,2}

¹ Department of Dermatology, University of Rome Tor Vergata, Via Montpellier 1, 00133 Rome, Italy

² Laboratory for Investigative Dermatology, The Rockefeller University, 1230 York Avenue, New York, NY 10065, USA

Correspondence should be addressed to Andrea Saggini; andreasaggini@gmail.com

Received 30 November 2013; Accepted 8 May 2014; Published 13 July 2014

Academic Editor: Clive Liu

Copyright © 2014 Andrea Saggini et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Psoriasis vulgaris (PV) is a cutaneous inflammatory disorder stemming from abnormal, persistent activation of the interleukin-(IL)-23/Th17 axis. Pustular psoriasis (PP) is a clinicopathological variant of psoriasis, histopathologically defined by the predominance of intraepidermal collections of neutrophils. Although PP pathogenesis is thought to largely follow that of (PV), recent evidences point to a more central role for IL-1, IL-36, and IL-6 in the development of PP. We review the role of IL-6 in the pathogenesis of PV and PP, focusing on its cross-talk with cytokines of the IL-23/Th17 axis. Clinical inhibitors of IL-6 signaling, including tocilizumab, have shown significant effectiveness in the treatment of several inflammatory rheumatic diseases, including rheumatoid arthritis and juvenile idiopathic arthritis; accordingly, anti-IL-6 agents may potentially represent future promising therapies for the treatment of PP.

1. Introduction

Psoriasis is an immune-mediated cutaneous disease with an estimated prevalence of approximately 2% in the European and North American population [1, 2]. The most common clinical presentation of psoriasis, namely, psoriasis vulgaris (PV), is defined by multiple erythematous plaques, histologically characterized by (1) epidermal acanthosis, hyperkeratosis, and parakeratosis; (2) dilated capillary network in the papillary dermis; (3) a mixed inflammatory infiltrate including polymorphonuclear cells, as well as intraepidermal collections of neutrophils [3]. Epidermal clusters of neutrophils have been given eponymous names such as Munro's microabscesses and Kogoj pustules [3]. Various evidences deriving from genetic studies, adoptive transfer models, and molecular evaluation of human samples point to a key pathogenetic role for T helper-1 (Th1)/Th17 cells and related cytokines (including TNF-alpha, IL-17, and IL-22), as well as for myeloid cell-derived cytokines such as IL-12 and IL-23 [1, 2, 4–8].

Pustular psoriasis (PP) is a clinicopathological variant of psoriasis distinguished by the following features: (1) clinically, presence of pustules on variably erythematous

skin; (2) histopathologically, predominance of intraepidermal collections of neutrophils [9–11]. Any bioptic sample presenting the histologic picture of PP should always undergo further investigations to rule out the eventuality of superficial dermatophytosis or *Candida albicans* infection, whose histopathologic features are often indistinguishable from those of PP [12, 13].

PP has been classified into generalized and localized forms [14]. Generalized PP is a life-threatening, systemic inflammatory condition characterized by repeated attacks of diffuse, erythematous, pustular rash associated with high-grade fever, general malaise, and frequent extracutaneous organ involvement; possible laboratory testing abnormalities include leukocytosis with left shift, increased erythrocyte sedimentation rate (ESR), or increased C-reactive protein (CRP) [14, 15]. Acute flare-ups of generalized PP may be triggered by pregnancy status, infection, or exposure to drugs [15]. Though generalized PP formally belongs to the psoriasis spectrum because of its frequent clinical association with PV and multiple similarities in molecular pathogenesis, it is debated whether it may represent a distinct clinicopathological entity [16, 17]. Another controversy is related to the classification of generalized PP alone or accompanied by PV

as distinct subtypes with different etiologies [17]. Likewise, localized PP, which is often limited to palms and soles (i.e., palmoplantar pustulosis), has been regarded by several authors as a separate entity rather than a clinical variant of psoriasis [17, 18]. However, a close relationship between localized PP and PV is likely suggested by lack of significant epidemiologic differences, frequent coexistence in the same patients, and largely shared genetic background [18].

Conventional first-line therapies for PP include topical corticosteroids, phototherapy, acitretin, cyclosporine, and methotrexate [14, 16]. Because the use of therapeutics is often hampered by low efficacy and/or adverse effect profile, a need to develop novel therapeutic approaches for PP is arising [14]. Infliximab is actually recognized by many experts as a first-line treatment option for PP, especially in severe cases [14, 19, 20]. Nonetheless, paradoxical TNF-alpha inhibitor-induced PP is a newly occurrence, whose pathogenic mechanism is still relatively unclear [21, 22].

The pathogenic process underlining PP development is only partially shared with PV [16, 17]. The efficacy of TNF-alpha inhibitors in most patients with PP or PV points to a crucial role of TNF-alpha in their pathogenesis [14]. In addition to TNF-alpha, alternative signaling pathways relevant to PP include those mediated by IL-17 and the IL-1/IL-36 family [17, 23–25]. Furthermore, recent evidence seems to indicate IL-6 as a new druggable target for PP [23].

2. Psoriasis Pathogenesis: Current Concepts

2.1. The IL-23/Th17 Axis in the Pathogenesis of Psoriasis. A distinct lineage of IL-23-responsive CD4⁺ T cells secreting IL-17A and IL-17F and expressing the lineage-specific transcription factor RORC has been recently identified as Th17 cells [1, 5, 26–28]. Additional effector cytokines produced by Th17 cells include IL-21 and IL-22, as well as other non-Th17-specific cytokines, such as IL-6 [29–31]. Cytokine requirements for inducing Th17 differentiation are similar in mice and humans [26, 32]. Naive CD4⁺ T-cell activation in the presence of both TGF-beta and IL-6 is key to priming the initial differentiation into Th17 cells [2, 27]. TGF-beta also exerts an indirect action through suppression of T-bet-dependent Th1 differentiation [2, 26]. IL-6-dependent STAT3 activation plays an essential role in Th17 differentiation by initially inducing the transcription of *RORC*, *IL17*, and *IL23R* genes and later promoting the expansion of differentiated and memory Th17 cells [26, 32]. However, TGF-beta and IL-6-driven Th17 cells are weakly functional without further exposure to IL-23; the latter cytokine is crucial for differentiation into effector cells, lineage stabilization, and full maturation to inflammatory Th17 cells [2, 5, 27, 28, 33].

Psoriasis skin lesions are the result of complex interactions between dendritic cells (DCs), keratinocytes, and Th1/Th17 lymphocytes [30, 34, 35]. Recent pathogenic models of psoriasis emphasized the role of IL-23/Th17 axis [1, 2, 5, 36]. IL-23 production by inflammatory DCs and activated keratinocytes stimulates Th17 cells within the dermis to release proinflammatory mediators such as IL-17 and IL-22 that, in turn, activate resident tissue cells, particularly keratinocytes

[33, 35]. Psoriatic plaques harbor higher levels of IL-23p19 and IL-12/23p40 than those of IL-12p35 [1, 27]; polymorphisms in *IL12/23p40* and *IL23R* genes are associated with increased risk of developing psoriasis, and injection of recombinant IL-23 into healthy skin results in inflammatory changes with histologic features of psoriasis [5, 30]. According to this evidence, the pathogenic relevance of IL-23 has been also confirmed by the high efficacy of both anti-IL-12/IL-23p40 monoclonal antibodies (i.e., ustekinumab) and IL-23p19 neutralizing agents (i.e., tildrakizumab) [8, 27, 33, 37, 38].

IL-17A (simply known as IL-17) belongs to the IL-17 cytokine family, which includes six members (from IL-17A to IL-17F) [1, 2]. IL-17A shows similar pleiotropic effects acting on a wide range of nonimmune cells, resulting in the induction of different proinflammatory cytokines, chemokines, antimicrobial peptides, nitric oxide, and matrix metalloproteinases [1, 2, 30, 34]. IL-17 is able to induce IL-6, IL-8, and CXCL5 in human skin keratinocytes, indirectly promoting the differentiation, activation, and migration of neutrophils [5, 34, 35]. Biopsies from PV plaques show elevated levels of IL-17 in parallel with increased expression of IL-23 and IL-22, while serum levels of IL-17 are correlated to psoriasis severity [2, 6, 30, 39]. IL-22 is another key downstream cytokine in the IL-23/Th17 axis, being upregulated in psoriatic skin as compared to normal skin [5, 29, 40, 41]; IL-22 mediates keratinocyte hyperplasia via STAT3 activation, leading to psoriasisiform hyperplasia. In the absence of IL-22, severity of both IL-23-mediated and imiquimod-induced psoriasis-like dermatitis in corresponding mouse models is markedly reduced [40, 42, 43].

A significant increase in IL-17 expression has been detected in lesional skin of PP, despite the absence of any significant increase in IL-12/IL-23 levels [44]; this is strikingly different from PV, where increased IL-17 levels are typically mirrored by analogous changes in IL-12/IL-23 expression [7, 37, 43]. Accordingly, conventional Th17 may not be the main driver for increased IL-17 expression in PP, with neutrophils being a possible, alternative source of IL-17 [23, 44]. Indeed, the anti-IL-23 agent ustekinumab appears to be significantly less effective in the treatment of PP than that of PV [44–46]. Of note, the immunopathology of two well-known histologic mimics of PP, that is, superficial dermatophytosis and mucocutaneous *Candida albicans* infection, relies heavily on the production of IL-17, as suggested by mouse models and rare human patients with loss-of-function defects in the *IL17* gene [47–50]. It is now clear that IL-17-dependent recruitment of neutrophils and secretion of antimicrobial peptides are crucial for cutaneous protection against dermatophytic infections and *Candida albicans* [47, 49–51]. Importantly, the cellular sources of IL-17 production in this setting are not limited to conventional CD4⁺T cells, as several components of the innate immunity (gamma/delta T cells, mast cells, and neutrophils) appear to be capable of immediate IL-17 secretion prior to the contribution of IL-23-dependent Th17 adaptive immunity [42, 48–52].

2.2. IL-36 and Pustular Psoriasis. Pathogenic *IL36RN* gene mutations have been identified in familial and sporadic cases of PP, either generalized or localized [25, 53, 54]; *IL36RN*

encodes the IL-36 receptor antagonist (IL-36Ra), a soluble mediator that antagonizes the proinflammatory activity of IL-36 cytokines (IL-36-alpha, IL-36-beta, and IL-36-gamma) through binding IL-36R (IL-1RL2) and inhibiting IL-36-dependent activation of NF-kappaB signaling [25, 55–57].

Several authors have detected elevation of keratinocyte-derived IL-36 cytokines levels in psoriatic lesional skin, as a result of keratinocyte stimulation by IL-17, IL-22, and TNF-alpha [58–60]. Primary epidermal IL-36 overexpression in transgenic mouse models results in PV-like phenotype histopathologically characterized by acanthosis, hyperkeratosis, and mixed inflammatory infiltration with predominance of neutrophils [55, 59]; further crossing with *IL36RN*-knockout strain augments IL-36 signaling leading to increased neutrophil infiltration and a histopathological picture more akin to classic PP [25, 55, 61]. Furthermore, loss of IL-36R signaling successfully counteracts development of imiquimod-induced psoriasiform dermatitis, pointing to a crucial role of IL-36 ligands in the proinflammatory activity of the IL-23/Th17 axis [61, 62]. Indeed, IL-36R signaling is relevant for the expansion of IL-17-producing T helper cells [25, 55].

IL-36 cytokines may exert a direct effect on immune cells [55]; activation of IL-36R, which is expressed constitutively on DCs, CD4+ T cells, and macrophages, promotes maturation of monocyte-derived DCs and induction of several cytokines, including IL-1, IL-6, IL-23, TNF-alpha, and IFN-gamma [59, 61, 63]. In addition, keratinocytes in psoriasis as well as synoviocytes in RA are capable of responding to direct IL-36 ligands stimulation with production of IL-6, IL-8, and antimicrobial peptides, which cooperate with IL-17A and TNF-alpha promoting neutrophil activation and migration [11, 54, 56, 60].

Thus, IL-36 ligands not only act as effector cytokines of the IL23/Th17 axis, but also induce several proinflammatory mediators (including IL-6, IL-8, and IL-23) that reinforce the Th17-driven inflammatory milieu [25, 59, 60, 63]. The cross-talk between IL-36 ligands and Th17 mediators establishes a positive feedback loop involving keratinocytes, DCs, macrophages, and Th17 [60, 61]; as a consequence, activation of T cells is enhanced, recruitment of immune cells in psoriatic lesions is augmented, and the IL-23/Th17 axis is reinforced [55, 60]. In keeping, elevation of IL-36R ligands in psoriatic plaques is closely correlated with increased levels of TNF-alpha, IL-17, and IL-22, confirming the existence of a proinflammatory, self-reinforcing gene expression loop [56, 59].

Pathogenetic *IL36RN* mutations associated with PP abolish the antagonistic effect of IL-36Ra, enhancing the IL-36-dependent production of IL-1, IL-6, and IL-8 [25, 54]. Indeed, patients with *IL36RN*-dependent genetic predisposition to PP have been treated effectively with anakinra, an IL-1 antagonist [64]. Nonetheless, so far no specific data regarding effectiveness of IL-6 inhibitors in *IL36RN*-dependent PP are available. Overall, recessive *IL36RN* mutations are associated with increased risk of PP alone, but not PV [57, 65–67]; both phenotypic variance and incomplete penetrance have been observed, supporting the notion that *IL36RN* mutations are able to induce manifest disease only in the presence of specific

environmental factors and/or further genetic defects at a second disease locus [25, 53, 65]. All genetic follow-up studies of PP patients have found evidence of genetic heterogeneity, proving that *IL36RN* mutations account for only a minority of sporadic PP cases [25, 57, 66].

3. IL-6 Signaling and Pustular Psoriasis

3.1. IL-6 Signaling and Selective IL-6 Inhibition. IL-6, a pleiotropic, proinflammatory cytokine, is the archetypal member of the gp130-related cytokine family, which also includes IL-11, IL-27, OSM, CNTF, CT-1, LIF, and CLC [68, 69]. IL-6 exerts its activity through interaction with a receptor complex composed of the nonsignaling alpha subunit IL-6R (CD126) and the common, ubiquitously expressed, beta subunit gp130 (CD130), resulting in immediate activation of receptor-associated kinases (JAK1/JAK2 and TYK2) and subsequent regulation of STAT1/STAT3 and SHP2-MAPK signaling pathways (Figure 1) [68, 70, 71]. The IL-6R subunit functions *in vivo* as both a conventional membrane-bound receptor, expressed on the surface of hepatocytes and certain inflammatory cells, and a soluble form (sIL-6R) which forms active IL-6/sIL-6R complexes (IL-6 transsignaling) [72, 73]; this property is unique to IL-6 among currently known cytokines [68–70].

In addition to being a major stimulus for the synthesis of acute-phase proteins, IL-6 promotes differentiation of B cells into mature plasma cells as well as T-cell differentiation and activation [69, 72]. Importantly, recent evidence demonstrated that IL-6 exerts a positive influence in initiating Th17 cell development, whereas it inhibits TGF-beta-dependent differentiation of regulatory T cells [32, 74]. IL-6 is also a downstream target gene of IL-17 signaling in nonimmune cells such as keratinocytes and fibroblasts [35, 72, 75]; this positive IL-6/IL-17 loop plays a key role in proinflammatory interactions between the immune system and nonimmune tissues [32, 76]. Additionally, IL-6 exerts a significant influence on myeloid precursor cells and circulating neutrophils [69, 77–79]: IL-6 promotes differentiation from myeloid progenitors to neutrophils as well as neutrophilia [80]. Furthermore, IL-6 secretion results in secondary production of chemokines such as IL-8 and MCP-1 by mononuclear cells/macrophages as well as expression of ICAM-1 and other adhesion molecules on endothelial cells, leading to enhanced neutrophil migration [75, 77, 79]. Last, mature neutrophils respond to IL-6 via membrane-bound IL-6R, releasing proinflammatory cytokines such as IL-23 and IL-17 and establishing a Th17-polarizing positive feedback loop [32, 76].

Transgenic *IL6*-KO mouse models are characterized by a unique resistance to several inflammatory conditions such as experimental autoimmune arthritis or encephalomyelitis [69, 70]; accordingly, IL-6 plays a central role in the pathogenesis of several autoimmune diseases, including rheumatoid arthritis, juvenile idiopathic arthritis, adult onset Still's disease, systemic lupus erythematosus, Takayasu's arteritis, and inflammatory bowel disease [69, 72, 75]. As a consequence, IL-6 has gained attention as an attractive therapeutic target

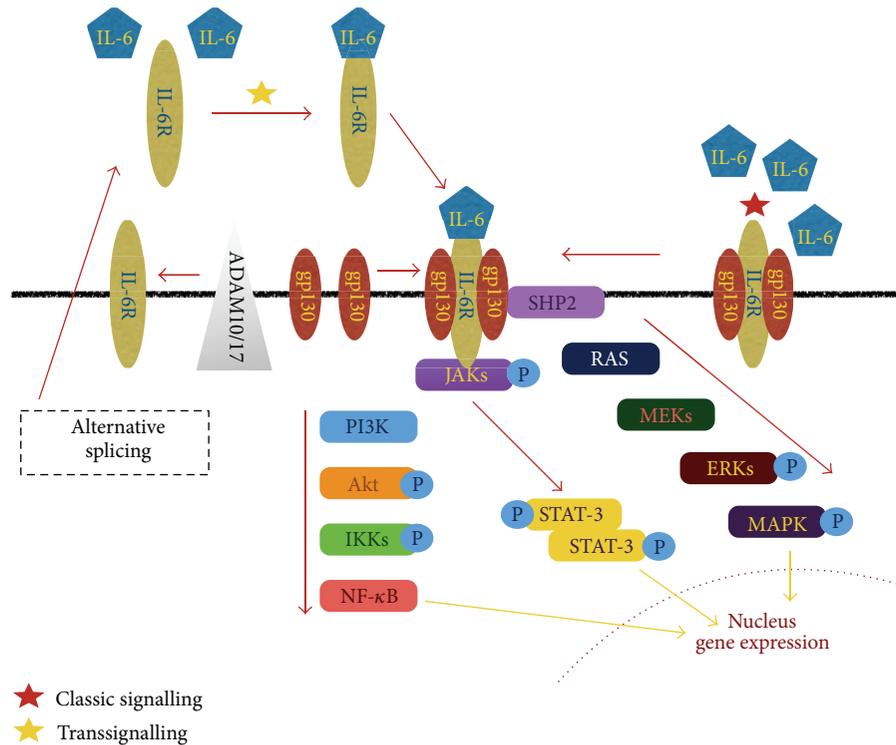


FIGURE 1: IL-6 signalling pathways. In classical signalling (red star), cells expressing membranous IL-6R are responsive to IL-6; in transsignalling (yellow star), cells lacking IL-6R are activated by IL-6/sIL-6R complexes (sIL-6R is generated by proteolytic shedding from IL-6R via ADAM10 and ADAM17 or by mRNA alternative splicing). Cellular events initiated by IL-6/IL-6R activity include activation of JAK, MEKs-ERKs, and PI3K/Akt kinases, resulting in changes in nuclear gene expression. IL-6: interleukin 6; sIL-6R: soluble interleukin 6 receptor.

for autoimmunity, leading to the clinical development of anti-IL-6R agents such as tocilizumab [72, 81]. Tocilizumab is a monoclonal antibody which globally blocks IL-6 biologic activity by antagonizing both conventional membrane-bound signaling and sIL-6/IL-6R transsignaling, resulting in a strong inhibition of IL-6-dependent STAT1/STAT3 activation [70]. Tocilizumab is an established therapeutic option for rheumatoid arthritis and juvenile idiopathic arthritis, although the field of tocilizumab-responsive autoimmune conditions is still expanding [68, 69, 81, 82].

3.2. IL-6 in the Pathogenesis of Psoriasis. IL-6 has long been associated with psoriasis pathogenesis [83–85]. In addition to known psoriasis susceptibility loci encoding proteins engaged in the TNF-alpha, IL-23, and IL-17 signaling pathways (including *HLA-Cw6*, *IL23R*, *IL12B*, *IL23A*, and *TNFAIP3* genes), *IL6* and *STAT3* polymorphisms have been linked with hereditary predisposition of developing psoriasis and response to TNF-alpha inhibitors [33, 86–89]. Increased skin and serum IL-6 levels are a feature of psoriasis [39, 84, 90]. Serum levels of IL-6 are regarded as a marker of the inflammatory activity in psoriasis as well as an indicator of treatment response [4, 39, 84, 85]; a positive correlation between IL-6 serum levels and clinical severity of PV before treatment has been described [4, 90]. Additionally, serum

IL-6 levels have been reported to decrease after effective treatment with methotrexate or UVB phototherapy [91, 92]. Furthermore, the likelihood of a positive K obner reaction has been reported to correlate with higher proportions of IL-6+ mast cells and IL-6R+ cells in the dermis [93].

IL-6 is produced by a wide range of cell types in psoriatic plaques (including keratinocytes, fibroblasts, endothelial cells, DCs, and macrophages) in response to several stimuli, such as IL-1, TNF-alpha, IL-17, and IL-36 (Figure 2) [84, 94–96]. Human keratinocytes stimulated by IL-17 or IL-36 may serve as a significant source of IL-6 [35, 76, 85, 94]; furthermore, a population of dermal slan-DCs has been recently identified as proinflammatory myeloid DCs in psoriatic skin lesions, which is capable of producing significant levels of IL-6 together with TNF-alpha, IL-1b, IL-23p19, and IL-12p70, all of which have proven crucial for the polarization of pathogenic Th17 and Th1 cells [95]. Importantly, the synergistic effects of IL-17 and TNF-alpha are capable of further upregulating IL-6 in psoriasis lesional skin; hence, selective targeting of either IL-17 or TNF-alpha exerts additional beneficial effects by indirectly reducing IL-6 levels [32, 35, 94, 96].

The key pathogenetic role of IL-6 signaling pathway in psoriasis is supported by evidence deriving from mouse models of psoriasis-like skin disease relying on constitutive

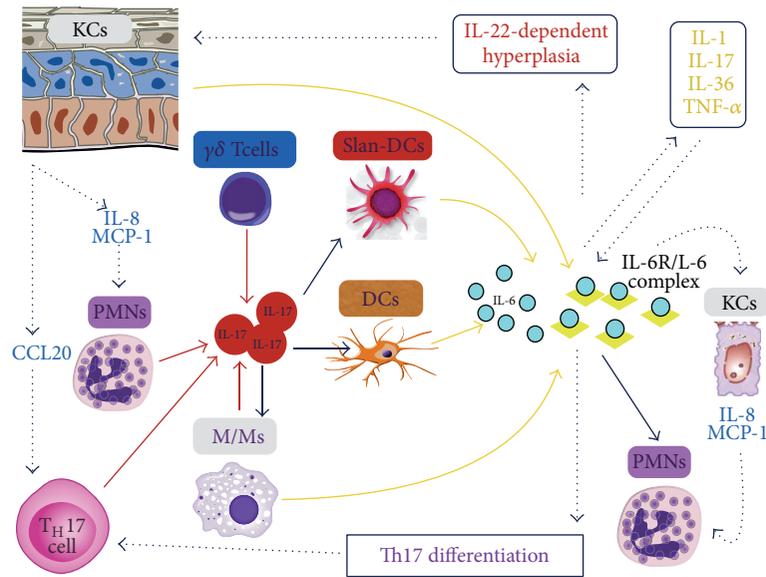


FIGURE 2: IL-17/IL-6 axis in the pathogenesis of pustular psoriasis. Both innate (gamma/delta T cells, neutrophils, and macrophages) and adaptive (Th17 cells) immunities contribute to cutaneous IL-17 production. Macrophages, conventional DCs, and slan-DCs respond to IL-17 by releasing IL-6, which in turn plays a key role in neutrophils recruitment and pustules formation; additional IL-6-dependent effects include reinforcement of Th1/Th17 inflammatory cytokines production, facilitation of IL-22-mediated epidermal hyperplasia, and naive CD4+ T cells differentiation into Th17. Activated keratinocytes amplify the IL-17/IL-6 axis by producing IL-6, recruiting Th17 cells through CCL20, and inducing neutrophils chemotaxis via IL-8 and MCP-1. DCs: dendritic cells; IL: interleukin; KCs: keratinocytes; M/Ms: monocytes/macrophages; PMNs: neutrophils; Th17: T helper 17 cells.

activation of STAT3 in keratinocytes [71, 97, 98]. Increased activation of STAT3 (pSTAT3) has been detected in lesional skin of psoriatic patients [98]; several cytokines upregulated in psoriasis, including IL-6, IL-20, and IL-22, signal through STAT3 activation [71, 98]. STAT3 phosphorylation influences the expression of genes controlling keratinocyte survival and proliferation through interactions with other transcription factors such as NF-kappaB [96, 99]. STAT3 activation has a key role in the psoriasis-associated IL-23 signaling cascade [71, 97, 99]. Accordingly, JAK inhibition is being assessed as a novel therapeutic strategy for treatment of psoriasis. Importantly, IL-6 produced by DCs, macrophages, T cells, and keratinocytes further augments the IL-6-rich microenvironment in psoriatic plaque, resulting in the robust induction of pSTAT3 in effector and memory Th17 cells [76]. Persistent pSTAT3 signaling in T cells is required for initial Th17 differentiation and promotion of Th17 cytokines production, unleashes unrestrained activation of effector T cells, and prevents suppressive activity of T regulatory cells [76]. Additionally, IL-6-mediated pSTAT3 signaling is capable of enhancing keratinocyte growth and proliferation, promoting psoriasis epidermal hyperplasia [96, 98]; IL-6 signaling on keratinocytes also induces chemoattractant proteins via AP-1 downstream activation [97].

IL-6 is a key mediator of IL-23/Th17-driven cutaneous inflammation [37, 94]. IL-23-induced dermal inflammation in psoriasis mouse models relies on T cells and IL-6 [96]. In IL-6-deficient mice, intradermal injections of IL-23 lead to increased IL-22 production compared with WT mice, but this response is not sufficient for effective dermal inflammation

and epidermal hyperplasia [96]. This finding seems to be secondary to insufficient expression of IL-22R1A in the absence of IL-6. The increased level of IL-6 in the skin of imiquimod-treated *IL17RA*-del mice compared with treated WT skin confirms the role of IL-6 in disease development in the absence of IL-17 signaling [41]. Accordingly, imiquimod is thought to indirectly activate the preexisting IL-17-producing T cells, which are capable of secreting other cytokines such as IL-6 that drive development of psoriasiform dermatitis independent of IL-17 [41, 43].

3.3. IL-6 and Pustular Psoriasis. Recent evidence points to an unexpected, central role of IL-6 in driving the abnormal recruitment of neutrophils into lesional skin of PP [23]; accordingly, IL-6 would be the key downstream mediator acting together with IL-17 to induce excessive skin infiltration by neutrophils resulting in intraepidermal pustules typical of PP (Figure 2) [23]. Importantly, IL-6 could be a novel, attractive target for the treatment of PP, in the light of the current availability of biologic agents safely and effectively antagonizing IL-6.

IL-6 has been long known to favor neutrophil differentiation and activation both *in vivo* and *in vitro* [79, 80]. Positive correlations have been recorded between IL-6 serum levels and clinical severity of PP, as well as associated leukocytosis, ESR, and CRP levels [100, 101]. Clinical improvement of PP following tonsillectomy has been paralleled by reduction of serum IL-6 levels [102]; in keeping, *in vitro* exposure of tonsillar mononuclear cells to streptococcal antigens resulted in increased production of IL-6 [91, 103].

The K14-*IL17A*-ind/+ transgenic mouse represents an animal model of psoriasiform dermatitis characterized by deregulated, persistent overexpression of IL-17A in epidermal keratinocytes leading to prominent development of intraepidermal neutrophil microabscesses in addition to dermal T-cell infiltration, hyperkeratosis, and parakeratosis [23]. The immunopathogenesis observed in the K14-*IL17A*-ind/+ strain strongly supports a mechanism whereby IL-6 propagates IL-17-induced inflammation, as confirmed by the noticeable presence of IL-6R α -expressing monocytes and neutrophils in the affected skin [23].

In this setting, the inflammatory cascade starts with epidermal IL-17A expression in the absence of IL-23 overexpression; similar conditions (i.e., a high IL-17A/IL-23 ratio) have been described as characteristic of bioptic samples of human PP compared to conventional PV (whereby IL-17A levels appear to follow those of IL-23). The persistent expression of IL-17A in basal keratinocytes seems to induce target cell to secrete significant amounts of IL-6, resulting in high levels of circulating IL-6 and sIL-6/IL-6R heterodimers [23]; increased levels of local and systemic IL-6 influence IL-6R- α + neutrophils and monocytes activity, leading to aberrant chemotaxis into lesional skin and formation of intraepidermal neutrophil microabscesses [23].

Importantly, administration of anti-IL-6 neutralizing antibody in K14-*IL17A*-ind/+ mice is sufficient to reduce and prevent the extent of leukocyte infiltration, leading to a sizeable decrease in cutaneous accumulation of myeloperoxidase+ CD11b+ cells, intraepidermal neutrophil microabscesses formation, and epidermal changes [23]. Hence, IL-6 seems to play a key role in the innate component of IL-17-driven PP-like dermatitis, and blockade of IL-6 activity may result in dramatic clinicopathological improvements despite the persistent activation of the IL-17 signaling.

Interestingly, gene expression evaluation of psoriatic plaques in the initial 48 hours after anti-TNF- α infliximab administration revealed significant inhibition of slan-DC-derived IL-1b, TNF- α , IFN- γ , IL-12, and IL-23 but not IL-6, suggesting that direct TNF- α blockade is less effective in targeting IL-6 production by inflammatory dermal DCs [95]. If IL-6 signaling was more relevant to PP development than to PV, such data would provide an explanation to clinical evidence that efficacy rates of TNF- α inhibitors in PP are lower as compared to PV [14].

4. Conclusions

So far, the experience with IL-6 inhibitors in psoriasis is limited, as other signaling pathways have been successfully investigated as therapeutic targets (i.e., TNF- α , IL-23, and IL-17) [8, 36, 38, 104]. Furthermore, paradoxical cases of biologic-induced psoriasiform dermatitis have been reported also for patients undergoing treatment with tocilizumab for RA [105, 106]. Tofacitinib and other Janus kinase inhibitors (targeting, among the others, also the IL-6R signaling pathway) are gaining significant attention as therapeutic options in psoriasis, but their efficacy in PP is still unclear [107, 108].

Only occasional patients with generalized PP, including paradoxical anti-TNF-induced cases, have been effectively treated with the anti-IL-6 agent tocilizumab [109, 110]. A larger amount of data exists with regard to the role of IL-1 antagonist anakinra in PP, especially in cases secondary to *IL36RN* mutations [24, 62, 64]. Nonetheless, it seems reasonable that IL-6 may play a crucial role as well as IL-1 independently from the persistent IL-36R activation in the epidermis [62]. If this evidence will be confirmed, agents neutralizing IL-1 and IL-6 may be effective in treating PP, similarly to juvenile idiopathic arthritis, which has been successfully treated with either anti-IL-1 agents or IL-6 inhibitors [82].

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- [1] M. A. Lowes, T. Kikuchi, J. Fuentes-Duculan et al., "Psoriasis vulgaris lesions contain discrete populations of Th1 and Th17 T cells," *Journal of Investigative Dermatology*, vol. 128, no. 5, pp. 1207–1211, 2008.
- [2] D. A. Martin, J. E. Towne, G. Kricorian et al., "The emerging role of IL-17 in the pathogenesis of psoriasis: preclinical and clinical findings," *Journal of Investigative Dermatology*, vol. 133, no. 1, pp. 17–26, 2013.
- [3] A. Ragaz and A. B. Ackerman, "Evolution, maturation, and regression of lesions of psoriasis. New observations and correlation of clinical and histologic findings," *The American Journal of Dermatopathology*, vol. 1, no. 3, pp. 199–214, 1979.
- [4] A. Balato, M. Schiattarella, R. di Caprio et al., "Effects of adalimumab therapy in adult subjects with moderate-to-severe psoriasis on Th17 pathway," *Journal of the European Academy of Dermatology and Venereology*, 2013.
- [5] A. Di Cesare, P. Di Meglio, and F. O. Nestle, "The IL-23/Th17 axis in the immunopathogenesis of psoriasis," *Journal of Investigative Dermatology*, vol. 129, no. 6, pp. 1339–1350, 2009.
- [6] X. Shi, L. Jin, E. Dang et al., "IL-17A upregulates keratin 17 expression in keratinocytes through STAT1- and STAT3-dependent mechanisms," *Journal of Investigative Dermatology*, vol. 131, no. 12, pp. 2401–2408, 2011.
- [7] S. Kagami, H. L. Rizzo, J. J. Lee, Y. Koguchi, and A. Blauvelt, "Circulating Th17, Th22, and Th1 cells are increased in psoriasis," *Journal of Investigative Dermatology*, vol. 130, no. 5, pp. 1373–1383, 2010.
- [8] N. H. Shear, J. Prinz, K. Papp, R. G. B. Langley, and W. P. Gulliver, "Targeting the interleukin-12/23 cytokine family in the treatment of psoriatic disease," *Journal of Cutaneous Medicine and Surgery*, vol. 12, supplement 1, pp. S1–S10, 2008.
- [9] S. H. Kardaun, H. Kuiper, V. Fidler, and M. F. Jonkman, "The histopathological spectrum of acute generalized exanthematous pustulosis (AGEP) and its differentiation from generalized pustular psoriasis," *Journal of Cutaneous Pathology*, vol. 37, no. 12, pp. 1220–1229, 2010.
- [10] N. Sanchez and A. B. Ackerman, "Subcorneal pustular dermatosis: a variant of pustular psoriasis," *Acta Dermato-Venereologica*, vol. 59, no. 85, pp. 147–151, 1979.
- [11] L. Skov, F. J. Beurskens, C. O. C. Zachariae et al., "IL-8 as antibody therapeutic target in inflammatory diseases: reduction

- of clinical activity in palmoplantar pustulosis," *The Journal of Immunology*, vol. 181, no. 1, pp. 669–679, 2008.
- [12] A. B. Ackerman, "Subtle clues to diagnosis by conventional microscopy. Neutrophils within the cornified layer as clues to infection by superficial fungi," *The American Journal of Dermatopathology*, vol. 1, no. 1, pp. 69–75, 1979.
- [13] A. Feily, M. R. Namazi, and H. Seifmanesh, "Generalized psoriasis-like dermatophytosis due to *Trichophyton rubrum*," *Acta Dermatovenerologica Croatica*, vol. 19, no. 3, pp. 209–211, 2011.
- [14] A. Robinson, A. S. van Voorhees, S. Hsu et al., "Treatment of pustular psoriasis: From the medical board of the National Psoriasis Foundation," *Journal of the American Academy of Dermatology*, vol. 67, no. 2, pp. 279–288, 2012.
- [15] J. Borges-Costa, R. Silva, L. Gonçalves, P. Filipe, L. S. De Almeida, and M. M. Gomes, "Clinical and laboratory features in acute generalized pustular psoriasis: a retrospective study of 34 patients," *American Journal of Clinical Dermatology*, vol. 12, no. 4, pp. 271–276, 2011.
- [16] S. Ikeda, H. Takahashi, Y. Suga et al., "Therapeutic depletion of myeloid lineage leukocytes in patients with generalized pustular psoriasis indicates a major role for neutrophils in the immunopathogenesis of psoriasis," *Journal of the American Academy of Dermatology*, vol. 68, no. 4, pp. 609–617, 2013.
- [17] H. B. Naik and E. W. Cowen, "Autoinflammatory pustular neutrophilic diseases," *Dermatologic Clinics*, vol. 31, no. 3, pp. 405–425, 2013.
- [18] A. M. G. Brunasso, M. Puntoni, W. Aberer, C. Delfino, L. Fancelli, and C. Massone, "Clinical and epidemiological comparison of patients affected by palmoplantar plaque psoriasis and palmoplantar pustulosis: a case series study," *British Journal of Dermatology*, vol. 168, no. 6, pp. 1243–1251, 2013.
- [19] M. Viguier, F. Aubin, E. Delaporte et al., "Efficacy and safety of tumor necrosis factor inhibitors in acute generalized pustular psoriasis," *Archives of Dermatology*, vol. 148, no. 12, pp. 1423–1425, 2012.
- [20] N. Smith, K. L. Harms, A. C. Hines et al., "Acute treatment of generalized pustular psoriasis of von Zumbusch with single-dose infliximab," *Journal of the American Academy of Dermatology*, vol. 68, no. 6, pp. e187–e189, 2013.
- [21] G. Egnatios, M. M. Warthan, R. Pariser, and A. F. Hood, "Pustular psoriasis following treatment of rheumatoid arthritis with TNF-alpha inhibitors," *Journal of Drugs in Dermatology*, vol. 7, no. 10, pp. 975–977, 2008.
- [22] E. Rallis, C. Korfitis, E. Stavropoulou, and M. Papaconstantis, "Onset of palmoplantar pustular psoriasis while on adalimumab for psoriatic arthritis: a "class effect" of TNF- α antagonists or simply an anti-psoriatic treatment adverse reaction?" *Journal of Dermatological Treatment*, vol. 21, no. 1, pp. 3–5, 2010.
- [23] A. L. Croxford, S. Karbach, F. C. Kurschus, S. Wortge, A. Nikolaev, and N. Yogeve, "IL-6 regulates neutrophil microabscess formation in IL-17A-driven psoriasiform lesions," *The Journal of Investigative Dermatology*, vol. 134, pp. 728–735, 2014.
- [24] U. Huffmeier, M. Watzold, J. Mohr, M. P. Schon, and R. Mossner, "Successful therapy with anakinra in a patient with generalized pustular psoriasis carrying IL36RN mutations," *The British Journal of Dermatology*, vol. 170, no. 1, pp. 202–204, 2014.
- [25] F. Capon, "IL36RN mutations in generalized pustular psoriasis: just the tip of the iceberg?" *Journal of Investigative Dermatology*, vol. 133, no. 11, pp. 2503–2504, 2013.
- [26] K. Hirahara, K. Ghoreschi, A. Laurence, X. Yang, Y. Kanno, and J. J. O'Shea, "Signal transduction pathways and transcriptional regulation in Th17 cell differentiation," *Cytokine and Growth Factor Reviews*, vol. 21, no. 6, pp. 425–434, 2010.
- [27] E. Lee, W. L. Trepicchio, J. L. Oestreicher et al., "Increased expression of interleukin 23 p19 and p40 in lesional skin of patients with psoriasis vulgaris," *Journal of Experimental Medicine*, vol. 199, no. 1, pp. 125–130, 2004.
- [28] S. Aggarwal, N. Ghilardi, M. Xie, F. J. De Sauvage, and A. L. Gurney, "Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17," *Journal of Biological Chemistry*, vol. 278, no. 3, pp. 1910–1914, 2003.
- [29] L. A. Zenewicz and R. A. Flavell, "Recent advances in IL-22 biology," *International Immunology*, vol. 23, no. 3, pp. 159–163, 2011.
- [30] H. L. Rizzo, S. Kagami, K. G. Phillips, S. E. Kurtz, S. L. Jacques, and A. Blauvelt, "IL-23-mediated psoriasis-like epidermal hyperplasia is dependent on IL-17A," *Journal of Immunology*, vol. 186, no. 3, pp. 1495–1502, 2011.
- [31] M. Sarra, R. Caruso, M. L. Cupi et al., "IL-21 promotes skin recruitment of CD4⁺ cells and drives IFN- γ -dependent epidermal hyperplasia," *Journal of Immunology*, vol. 186, no. 9, pp. 5435–5442, 2011.
- [32] A. Camporeale and V. Poli, "IL-6, IL-17 and STAT3: a holy trinity in auto-immunity?" *Frontiers in Bioscience*, vol. 17, no. 6, pp. 2306–2326, 2011.
- [33] P. di Meglio, F. Villanova, L. Napolitano et al., "The IL23R A/Gln381 allele promotes IL-23 unresponsiveness in human memory T-helper 17 cells and impairs Th17 responses in psoriasis patients," *Journal of Investigative Dermatology*, vol. 1, 2013.
- [34] K. E. Nograles, L. C. Zaba, E. Guttman-Yassky et al., "Th17 cytokines interleukin (IL)-17 and IL-22 modulate distinct inflammatory and keratinocyte-response pathways," *British Journal of Dermatology*, vol. 159, no. 5, pp. 1092–1102, 2008.
- [35] A. Chiricozzi, E. Guttman-Yassky, M. Suárez-Farías et al., "Integrative responses to IL-17 and TNF- α in human keratinocytes account for key inflammatory pathogenic circuits in psoriasis," *Journal of Investigative Dermatology*, vol. 131, no. 3, pp. 677–687, 2011.
- [36] M. A. Lowes, C. B. Russell, D. A. Martin, J. E. Towne, and J. G. Krueger, "The IL-23/Th17 pathogenic axis in psoriasis is amplified by keratinocyte responses," *Trends in Immunology*, vol. 34, no. 4, pp. 174–181, 2013.
- [37] A. Chiricozzi and J. G. Krueger, "IL-17 targeted therapies for psoriasis," *Expert Opinion on Investigational Drugs*, vol. 22, no. 8, pp. 993–1005, 2013.
- [38] W. Alexander, "American academy of dermatology cardiovascular research technologies 2013 American college of cardiology," *P & T*, vol. 38, no. 5, pp. 288–292, 2013.
- [39] O. Arican, M. Aral, S. Sasmaz, and P. Ciragil, "Serum levels of TNF- α , IFN- γ , IL-6, IL-8, IL-12, IL-17, and IL-18 in patients with active psoriasis and correlation with disease severity," *Mediators of Inflammation*, vol. 2005, no. 5, pp. 273–279, 2005.
- [40] A. B. Van Belle, M. De Heusch, M. M. Lemaire et al., "IL-22 is required for imiquimod-induced psoriasiform skin inflammation in mice," *Journal of Immunology*, vol. 188, no. 1, pp. 462–469, 2012.
- [41] K. El Malki, S. H. Karbach, J. Huppert et al., "An alternative pathway of imiquimod-induced psoriasis-like skin inflammation in the absence of interleukin-17 receptor signaling," *Journal of Investigative Dermatology*, vol. 133, no. 2, pp. 441–451, 2013.

- [42] T. Mabuchi, T. Takekoshi, and S. T. Hwang, "Epidermal CCR6⁺ $\gamma\delta$ T cells are major producers of IL-22 and IL-17 in a murine model of psoriasisform dermatitis," *Journal of Immunology*, vol. 187, no. 10, pp. 5026–5031, 2011.
- [43] L. van der Fits, S. Mourits, J. S. A. Voerman et al., "Imiquimod-induced psoriasis-like skin inflammation in mice is mediated via the IL-23/IL-17 axis," *The Journal of Immunology*, vol. 182, no. 9, pp. 5836–5845, 2009.
- [44] R. Bissonnette, S. Nigen, R. G. Langley et al., "Increased expression of IL-17A and limited involvement of IL-23 in patients with palmo-plantar (PP) pustular psoriasis or PP pustulosis; results from a randomised controlled trial," *Journal of the European Academy of Dermatology and Venereology*, 2013.
- [45] S. Gregoriou, C. Kazakos, E. Christofidou, G. Kontochristopoulos, G. Vakis, and D. Rigopoulos, "Pustular psoriasis development after initial ustekinumab administration," *European Journal of Dermatology*, vol. 21, no. 1, pp. 104–105, 2011.
- [46] K. S. Wenk, J. M. Claros, and A. Ehrlich, "Flare of pustular psoriasis after initiating ustekinumab therapy," *Journal of Dermatological Treatment*, vol. 23, no. 3, pp. 212–214, 2012.
- [47] A. Puel, S. Cypowyj, L. Maródi, L. Abel, C. Picard, and J. Casanova, "Inborn errors of human IL-17 immunity underlie chronic mucocutaneous candidiasis," *Current Opinion in Allergy and Clinical Immunology*, vol. 12, no. 6, pp. 616–622, 2012.
- [48] H. Zhang, H. Li, Y. Li, Y. Zou, X. Dong, and W. Song, "IL-17 plays a central role in initiating experimental *Candida albicans* infection in mouse corneas," *European Journal of Immunology*, vol. 43, no. 10, pp. 2671–2682, 2013.
- [49] S. Cypowyj, C. Picard, L. Maródi, J. Casanova, and A. Puel, "Immunity to infection in IL-17-deficient mice and humans," *European Journal of Immunology*, vol. 42, no. 9, pp. 2246–2254, 2012.
- [50] A. Gladiator, N. Wangler, K. Trautwein-Weidner, and S. LeibundGut-Landmann, "Cutting edge: IL-17-secreting innate lymphoid cells are essential for host defense against fungal infection," *The Journal of Immunology*, vol. 190, no. 2, pp. 521–525, 2013.
- [51] N. Hernández-Santos and S. L. Gaffen, "Th17 cells in immunity to *Candida albicans*," *Cell Host and Microbe*, vol. 11, no. 5, pp. 425–435, 2012.
- [52] A. M. Lin, C. J. Rubin, R. Khandpur et al., "Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis," *Journal of Immunology*, vol. 187, no. 1, pp. 490–500, 2011.
- [53] M. Li, J. Han, Z. Lu et al., "Prevalent and rare mutations in IL-36RN gene in chinese patients with generalized pustular psoriasis and psoriasis vulgaris," *Journal of Investigative Dermatology*, vol. 133, no. 11, pp. 2637–2639, 2013.
- [54] S. Marrakchi, P. Guigue, B. R. Renshaw et al., "Interleukin-36-receptor antagonist deficiency and generalized pustular psoriasis," *The New England Journal of Medicine*, vol. 365, no. 7, pp. 620–628, 2011.
- [55] J. Towne and J. Sims, "IL-36 in psoriasis," *Current Opinion in Pharmacology*, vol. 12, no. 4, pp. 486–490, 2012.
- [56] D. Tripodi, F. Conti, M. Rosati et al., "IL-36 a new member of the IL-1 family cytokines," *Journal of Biological Regulators and Homeostatic Agents*, vol. 26, no. 1, pp. 7–14, 2012.
- [57] A. Körber, R. Mössner, R. Renner et al., "Mutations in IL36RN in patients with generalized pustular psoriasis," *Journal of Investigative Dermatology*, vol. 133, no. 11, pp. 2634–2637, 2013.
- [58] P. Muhr, J. Zeitvogel, I. Heitland, T. Werfel, and M. Wittmann, "Expression of interleukin (IL)-1 family members upon stimulation with IL-17 differs in keratinocytes derived from patients with psoriasis and healthy donors," *The British Journal of Dermatology*, vol. 165, no. 1, pp. 189–193, 2011.
- [59] H. Blumberg, H. Dinh, C. Dean Jr. et al., "IL-1RL2 and its ligands contribute to the cytokine network in psoriasis," *The Journal of Immunology*, vol. 185, no. 7, pp. 4354–4362, 2010.
- [60] Y. Carrier, H.-L. Ma, H. E. Ramon et al., "Inter-regulation of Th17 cytokines and the IL-36 cytokines *in vitro* and *in vivo*: implications in psoriasis pathogenesis," *Journal of Investigative Dermatology*, vol. 131, no. 12, pp. 2428–2437, 2011.
- [61] L. Tortola, E. Rosenwald, B. Abel et al., "Psoriasisform dermatitis is driven by IL-36-mediated DC-keratinocyte crosstalk," *Journal of Clinical Investigation*, vol. 122, no. 11, pp. 3965–3976, 2012.
- [62] M. Uribe-Herranz, L. Lian, K. M. Hooper, K. A. Milora, and L. E. Jensen, "IL-1R1 signaling facilitates murine microabscess formation in psoriasisform imiquimod-induced skin inflammation," *Journal of Investigative Dermatology*, vol. 133, no. 6, pp. 1541–1549, 2013.
- [63] A. Johnston, X. Xing, A. M. Guzman et al., "IL-1F5, -F6, -F8, and -F9: a novel IL-1 family signaling system that is active in psoriasis and promotes keratinocyte antimicrobial peptide expression," *Journal of Immunology*, vol. 186, no. 4, pp. 2613–2622, 2011.
- [64] L. Rossi-Semerano, M. Piram, C. Chiaverini, D. De Ricaud, A. Smahi, and I. Kone-Paut, "First clinical description of an infant with interleukin-36-receptor antagonist deficiency successfully treated with anakinra," *Pediatrics*, vol. 132, no. 4, pp. e1043–e1047, 2013.
- [65] N. Setta-Kaffetzi, A. A. Navarini, V. M. Patel et al., "Rare pathogenic variants in IL36RN underlie a spectrum of psoriasis-associated pustular phenotypes," *Journal of Investigative Dermatology*, vol. 133, no. 5, pp. 1366–1369, 2013.
- [66] K. Sugiura, A. Takemoto, M. Yamaguchi et al., "The majority of generalized pustular psoriasis without psoriasis vulgaris is caused by deficiency of interleukin-36 receptor antagonist," *Journal of Investigative Dermatology*, vol. 133, no. 11, pp. 2514–2521, 2013.
- [67] D. M. Berki, S. K. Mahil, A. David Burden et al., "Loss of IL36RN function does not confer susceptibility to psoriasis vulgaris," *Journal of Investigative Dermatology*, vol. 134, pp. 271–273, 2014.
- [68] M. F. Neurath and S. Finotto, "IL-6 signaling in autoimmunity, chronic inflammation and inflammation-associated cancer," *Cytokine and Growth Factor Reviews*, vol. 22, no. 2, pp. 83–89, 2011.
- [69] M. Rincon, "Interleukin-6: from an inflammatory marker to a target for inflammatory diseases," *Trends in Immunology*, vol. 33, no. 11, pp. 571–577, 2012.
- [70] S. A. Jones, J. Scheller, and S. Rose-John, "Therapeutic strategies for the clinical blockade of IL-6/gp130 signaling," *Journal of Clinical Investigation*, vol. 121, no. 9, pp. 3375–3383, 2011.
- [71] K. Miyoshi, M. Takaishi, K. Nakajima et al., "Stat3 as a therapeutic target for the treatment of psoriasis: a clinical feasibility study with STA-21, a Stat3 Inhibitor," *Journal of Investigative Dermatology*, vol. 131, no. 1, pp. 108–117, 2011.
- [72] P. Ataie-Kachoei, M. H. Pourgholami, and D. L. Morris, "Inhibition of the IL-6 signaling pathway: a strategy to combat chronic inflammatory diseases and cancer," *Cytokine and Growth Factor Reviews*, vol. 24, no. 2, pp. 163–173, 2013.

- [73] S. Rose-John, "IL-6 trans-signaling via the soluble IL-6 receptor: importance for the proinflammatory activities of IL-6," *International Journal of Biological Sciences*, vol. 8, no. 9, pp. 1237–1247, 2012.
- [74] W. A. Goodman, A. D. Levine, J. V. Massari, H. Sugiyama, T. S. McCormick, and K. D. Cooper, "IL-6 signaling in psoriasis prevents immune suppression by regulatory T cells," *Journal of Immunology*, vol. 183, no. 5, pp. 3170–3176, 2009.
- [75] K. Ishihara and T. Hirano, "IL-6 in autoimmune disease and chronic inflammatory proliferative disease," *Cytokine and Growth Factor Reviews*, vol. 13, no. 4–5, pp. 357–368, 2002.
- [76] H. Ogura, M. Murakami, Y. Okuyama et al., "Interleukin-17 promotes autoimmunity by triggering a positive-feedback loop via interleukin-6 induction," *Immunity*, vol. 29, no. 4, pp. 628–636, 2008.
- [77] M. Romano, M. Sironi, C. Toniatti et al., "Role of IL-6 and its soluble receptor in induction of chemokines and leukocyte recruitment," *Immunity*, vol. 6, no. 3, pp. 315–325, 1997.
- [78] R. M. McLoughlin, J. Witowski, R. L. Robson et al., "Interplay between IFN- γ and IL-6 signaling governs neutrophil trafficking and apoptosis during acute inflammation," *Journal of Clinical Investigation*, vol. 112, no. 4, pp. 598–607, 2003.
- [79] E. Bartoccioni, F. Scuderi, M. Marino, and C. Provenzano, "IL-6, monocyte infiltration and parenchymal cells," *Trends in Immunology*, vol. 24, no. 6, pp. 299–301, 2003.
- [80] G. Kaplanski, V. Marin, F. Montero-Julian, A. Mantovani, and C. Farnarier, "IL-6: a regulator of the transition from neutrophil to monocyte recruitment during inflammation," *Trends in Immunology*, vol. 24, no. 1, pp. 25–29, 2003.
- [81] M. Murakami and N. Nishimoto, "The value of blocking IL-6 outside of rheumatoid arthritis: current perspective," *Current Opinion in Rheumatology*, vol. 23, no. 3, pp. 273–277, 2011.
- [82] G. Horneff, "Update on biologicals for treatment of juvenile idiopathic arthritis," *Expert Opinion on Biological Therapy*, vol. 13, no. 3, pp. 361–376, 2013.
- [83] A. Castells-Rodellas, J. V. Castell, A. Ramirez-Bosca, J. F. Nicolas, F. Valcuende-Cavero, and J. Thivolet, "Interleukin-6 in normal skin and psoriasis," *Acta Dermato-Venereologica*, vol. 72, no. 3, pp. 165–168, 1992.
- [84] P. Neuner, A. Urbanski, F. Trautinger et al., "Increased IL-6 production by monocytes and keratinocytes in patients with psoriasis," *Journal of Investigative Dermatology*, vol. 97, no. 1, pp. 27–33, 1991.
- [85] R. M. Grossman, J. Krueger, D. Yourish et al., "Interleukin 6 is expressed in high levels of psoriatic skin and stimulates proliferation of cultured human keratinocytes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 86, no. 16, pp. 6367–6371, 1989.
- [86] W. Baran, J. C. Szepietowski, G. Mazur, and E. Baran, "IL-6 and IL-10 promoter gene polymorphisms in psoriasis vulgaris," *Acta Dermato-Venereologica*, vol. 88, no. 2, pp. 113–116, 2008.
- [87] A. N. Boca, M. L. Talamonti, M. Galluzzo et al., "Genetic variations in IL6 and IL12B decreasing the risk for psoriasis," *Immunology Letters*, vol. 156, no. 1–2, pp. 127–131, 2013.
- [88] T. Tejasvi, P. E. Stuart, V. Chandran et al., "TNFAIP3 gene polymorphisms are associated with response to TNF blockade in psoriasis," *Journal of Investigative Dermatology*, vol. 132, no. 3, part 1, pp. 593–600, 2012.
- [89] L. Di Renzo, A. Bianchi, R. Saraceno et al., "174G/C IL-6 gene promoter polymorphism predicts therapeutic response to TNF- α blockers," *Pharmacogenetics and Genomics*, vol. 22, no. 2, pp. 134–142, 2012.
- [90] B. Toruniowa, D. Krasowska, M. Koziol, A. Ksiazek, and A. Pietrzak, "Serum levels of IL-6 in mycosis fungoides, psoriasis, and lichen planus," *Annals of the New York Academy of Sciences*, vol. 762, pp. 432–434, 1995.
- [91] H. Mizutani, Y. Ohmoto, T. Mizutani, M. Murata, and M. Shimizu, "Role of increased production of monocytes TNF- α , IL-1 β and IL-6 in psoriasis: Relation to focal infection, disease activity and responses to treatments," *Journal of Dermatological Science*, vol. 14, no. 2, pp. 145–153, 1997.
- [92] Y. Lo, K. Torii, C. Saito, T. Furuhashi, A. Maeda, and A. Morita, "Serum IL-22 correlates with psoriatic severity and serum IL-6 correlates with susceptibility to phototherapy," *Journal of Dermatological Science*, vol. 58, no. 3, pp. 225–227, 2010.
- [93] M.-M. Suttle, G. Nilsson, E. Snellman, and I. T. Harvima, "Experimentally induced psoriatic lesion associates with interleukin (IL)-6 in mast cells and appearance of dermal cells expressing IL-33 and IL-6 receptor," *Clinical and Experimental Immunology*, vol. 169, no. 3, pp. 311–319, 2012.
- [94] S. Fujishima, H. Watanabe, M. Kawaguchi et al., "Involvement of IL-17F via the induction of IL-6 in psoriasis," *Archives of Dermatological Research*, vol. 302, no. 7, pp. 499–505, 2010.
- [95] A. Hänsel, C. Günther, J. Ingwersen et al., "Human slan (6-sulfo LacNAc) dendritic cells are inflammatory dermal dendritic cells in psoriasis and drive strong Th17/Th1 T-cell responses," *Journal of Allergy and Clinical Immunology*, vol. 127, no. 3, pp. 787–794, 2011.
- [96] J. Lindroos, L. Svensson, H. Norsgaard et al., "IL-23-mediated epidermal hyperplasia is dependent on IL-6," *Journal of Investigative Dermatology*, vol. 131, no. 5, pp. 1110–1118, 2011.
- [97] K. Nakajima, T. Kanda, M. Takaishi et al., "Distinct roles of IL-23 and IL-17 in the development of psoriasis-like lesions in a mouse model," *The Journal of Immunology*, vol. 186, no. 7, pp. 4481–4489, 2011.
- [98] S. Sano, K. S. Chan, S. Carbajal et al., "Stat3 links activated keratinocytes and immunocytes required for development of psoriasis in a novel transgenic mouse model," *Nature Medicine*, vol. 11, no. 1, pp. 43–49, 2005.
- [99] R. M. Andrés, M. C. Montesinos, P. Navalón, M. Payá, and M. C. Terencio, "NF- κ B and STAT3 Inhibition as a therapeutic strategy in psoriasis: in vitro and in vivo effects of BTH," *Journal of Investigative Dermatology*, vol. 133, no. 10, pp. 2362–2371, 2013.
- [100] J. Kamarashev, P. Lor, A. Forster, L. Heinzerling, G. Burg, and F. O. Nestle, "Generalised pustular psoriasis induced by cyclosporin a withdrawal responding to the tumour necrosis factor alpha inhibitor etanercept," *Dermatology*, vol. 205, no. 2, pp. 213–216, 2002.
- [101] M. Yamamoto, Y. Imai, Y. Sakaguchi, T. Haneda, and K. Yamaniishi, "Serum cytokines correlated with the disease severity of generalized pustular psoriasis," *Disease Markers*, vol. 34, no. 3, pp. 153–161, 2013.
- [102] T. Nakamura, M. Oishi, M. Johno, T. Ono, and M. Honda, "Serum levels of interleukin 6 in patients with pustulosis palmaris et plantaris," *Journal of Dermatology*, vol. 20, no. 12, pp. 763–766, 1993.
- [103] H. Murakata, Y. Harabuchi, Y. Kukuminato, Y. Yokoyama, and A. Kataura, "Cytokine production by tonsillar lymphocytes stimulated with alpha-streptococci in patients with pustulosis palmaris et plantaris," *Acta Oto-Laryngologica, Supplement*, vol. 523, pp. 201–203, 1996.
- [104] K. A. Papp, C. Leonardi, A. Menter et al., "Brodalumab, an anti-interleukin-17-receptor antibody for psoriasis," *The New England Journal of Medicine*, vol. 366, no. 13, pp. 1181–1189, 2012.

- [105] A. Grasland, E. Mahé, E. Raynaud, and I. Mahé, "Psoriasis onset with tocilizumab," *Joint Bone Spine*, vol. 80, no. 5, pp. 541–542, 2013.
- [106] D. Wendling, H. Letho-Gyselinck, X. Guillot, and C. Prati, "Psoriasis onset with tocilizumab treatment for rheumatoid arthritis," *Journal of Rheumatology*, vol. 39, no. 3, p. 657, 2012.
- [107] K. A. Papp, A. Menter, B. Strober et al., "Efficacy and safety of tofacitinib, an oral Janus kinase inhibitor, in the treatment of psoriasis: a Phase 2b randomized placebo-controlled dose-ranging study," *British Journal of Dermatology*, vol. 167, no. 3, pp. 668–677, 2012.
- [108] K. Ghoreschi and M. Gadina, "Jakpot! new small molecules in autoimmune and inflammatory diseases," *Experimental Dermatology*, vol. 23, no. 1, pp. 7–11, 2013.
- [109] S. Younis, D. Rimar, G. Slobodin, and I. Rosner, "Tumor necrosis factor-associated palmoplantar pustular psoriasis treated with interleukin 6 blocker," *Journal of Rheumatology*, vol. 39, no. 10, pp. 2055–2056, 2012.
- [110] J. Rueda-Gotor, M. A. González-Gay, R. Blanco Alonso, C. Gonzalez-Vela, C. Lopez-Obregon, and M. A. González-López, "Successful effect of tocilizumab in anti-TNF- α -induced palmoplantar pustulosis in rheumatoid arthritis," *Joint Bone Spine*, vol. 79, no. 5, pp. 510–513, 2012.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

