

1 **Effect of ball-milling on crystallinity index, degree of polymerization and thermal stability**
2 **of cellulose: ~~a systematic study~~**

3 Marco Mattonai^a, Dominika Pawcenis^b, Serena del Seppia^a, Joanna Łojewska^b, Erika
4 Ribechini^{a*}

5 ^aDepartment of Chemistry and Industrial Chemistry, University of Pisa, Via G. Moruzzi 13,
6 56124 Pisa, Italy

7 ^bDepartment of Chemistry, Jagiellonian University, Gronostajowa 2, 30-387 Kraków, Poland

8 *Corresponding Author, erika.ribechini@unipi.it

9

10 **ABSTRACT**

11 A combined study of crystallinity index (CI), degree of polymerisation (DP) and thermal
12 stability of cellulose was carried out for monitoring the effect of ball-milling. DP and CI are
13 two fundamental quantities that describe the physico-chemical behaviour of cellulose. Milling
14 is a common strategy to reduce cellulose crystallinity. In this work, four different
15 commercially available celluloses were milled at 30, 60 and 120 min, and the changes in DP
16 and CI were monitored using spectroscopic, diffraction and chromatographic techniques.
17 Evolved gas analysis-mass spectrometry (EGA-MS) was also used to evaluate the changes in
18 apparent activation energy (E_a) of the pyrolysis reaction upon different milling times by using
19 model-free isoconversional methods. The results showed substantial decrease in CI values
20 and moderate changes in DP after two-hours of ball-milling. E_a were found in the range 110-
21 140 kJ/mol, and were reduced by 10% on average after two hours of ball-milling.

22

23 **Keywords:** *Cellulose; Crystallinity index; Degree of polymerization; Isoconversional methods;*

24 *EGA-MS*

25

26

27 **1. INTRODUCTION**

28 Cellulose, the most abundant organic polymer on Earth, is an infinitely surprising material,
29 with application potential in many research fields including renewable energy, material
30 development and medicine (Kian et al., 2018; Lavoine et al., 2012; Qiu & Hu, 2013; Siqueira
31 et al., 2010; Yang et al., 2018). Two main parameters describe the structure of cellulose: the
32 degree of polymerisation (DP), which is the average length of the polysaccharide chains, and
33 the crystallinity index (CI), which is the percentage of crystalline cellulose. The chemical and
34 physico-chemical behaviour of cellulose is deeply influenced by these parameters. Celluloses
35 with high DPs and CIs are more resistant to hydrolysis and show increased density and tensile
36 strength (Bledzki & Gassan, 1999; Hallac & Ragauskas, 2011; Marx-Figini & Maximino, 1993;
37 Schenzel et al., 2005; Siqueira et al., 2010). A recent paper by Mukarakate and co-workers
38 also provides a systematic study of the effect of crystallinity on the thermal stability and
39 products distribution of cellulose allomorphs during pyrolysis (Mukarakate et al., 2016).

40 The knowledge of DP, CI and thermal stability of a cellulosic material is fundamental for the
41 optimization of its conversion processes, on both the laboratory and the industrial scale. The
42 polymerization degree is usually estimated using viscosimetry or size-exclusion
43 chromatography (SEC) (De Silva & Byrne, 2017; Hallac & Ragauskas, 2011; Łojewski et al.,
44 2010; Ni et al., 2015; Podzimek, 2014; Rebière et al., 2016). SEC analysis with multi-angle light-
45 scattering (MALS) detection constitutes a special case, since this technique provides the so
46 called absolute DP values and requires no calibration with molar mass standards such as
47 polystyrene samples with different hydrodynamic radii (Pawcenis et al., 2015; Podzimek,
48 2011). The crystallinity index is typically measured by XRD spectroscopy (French & Santiago
49 Cintrón, 2013; Lee & Mani, 2017; Zhao et al., 2017), but reliable estimations can also be

50 obtained by infrared and Raman-infrared spectroscopy (Agarwal et al., 2010; Schenzel et al.,
51 2005). The thermal stability can be estimated by thermogravimetric analysis (TGA-MS or TGA-
52 FTIR) by using model-free or model-based isoconversional methods. The advantage of model-
53 free isoconversional methods is that no assumptions are made on the thermal degradation
54 mechanism. This allows obtaining meaningful data even for samples with very complex
55 thermal degradation pathways such as cellulose (Ali et al., 2017; Kaur et al., 2018; Sánchez-
56 Jiménez et al., 2011). Recently, model-free isoconversional methods were also applied to
57 evolved gas analysis-mass spectrometry (EGA-MS) (Kim et al., 2017), showing that this
58 technique can provide the same information as TGA.

59 Milling is a very common strategy to reduce the CI of cellulose (Alvira et al., 2010; da Silva et
60 al., 2010; Gu et al., 2018; Wang et al., 2017). However, it is difficult to find a consensus in the
61 literature regarding the milling time required to eliminate the crystalline phase. Avolio et al.
62 (Avolio et al., 2012) showed that 60 minutes of ball-milling reduce the CI of cellulose from
63 0.53 to 0.15, but are not sufficient to achieve total amorphization. Schenzel and co-workers
64 (Schenzel et al., 2005) obtained a totally amorphous cellulose after 30 hours of milling, and
65 mixed amorphous and un-milled cellulose to obtain samples at different CIs. A similar strategy
66 was used by Agarwal and co-workers (Agarwal et al., 2010), but totally amorphous cellulose
67 was obtained after only 120 minutes of milling. The paper by Agarwal and co-workers also
68 provides viscosimetric measurements of the DP before and after milling, but the variation is
69 considered as negligible.

70 In the present work, four commercially available celluloses were milled at 30, 60 and 120
71 minutes. The effect of ball milling on both the CI and the DP was investigated using ATR-FTIR,
72 XRD and SEC-MALS. EGA-MS was used to calculate the apparent activation energy for the

73 pyrolysis reaction of all the samples by application of model-free isoconversional methods.
74 To the best of our knowledge, this work is a first systematic study of the effects of milling on
75 CI, DP and thermal stability of cellulose using multiple analytical techniques and
76 isoconversional methods.

77

78 **2. MATERIALS AND METHODS**

79 **2.1 Materials:** Pyridine ($\geq 99.9\%$, Sigma, USA), phenyl-isocyanate ($\geq 99.9\%$, Sigma, USA),
80 methanol (Stanlab, Poland) and tetrahydrofuran (Chempur, Poland) were used for cellulose
81 derivatisation and SEC analyses. Four different cellulose samples were purchased from Sigma,
82 USA: Sigmacell type 20 (labelled "S20", from cotton linters, product n° 310697), Sigmacell
83 type 50 (labelled "S50", from cotton linters, product n° S5504), Microcrystalline cellulose
84 (labelled "SMC", from cotton linters, product n° 435263), and acid-washed cellulose (labelled
85 "SAW", from fir, product n° 22182). All cellulose samples were milled for 30, 60 and 120 min
86 using a Pulverisette 23 ball-mill (Fritsch, Germany).

87

88 **2.2 ATR-FTIR:** Infrared spectra were recorded for all samples in ATR mode using a Nicolet
89 8700 FT-IR spectrometer (Thermo Scientific, USA) and a GladiATR sample holder (PIKE
90 Technologies, USA). The sample holder was equipped with a heating plate, which was kept at
91 105 °C during all measurements to ensure the removal of bound water, which gives the
92 bending vibrational modes at around 1640 cm^{-1} . Spectra were recorded in the range 650–
93 4000 cm^{-1} with a resolution of 4 cm^{-1} . Background spectra were collected before every
94 sample, and all spectra were corrected by subtraction of atmospheric H₂O and CO₂ signals. All
95 spectra were collected in triplicates and averaged. The spectra were processed with OMNIC

96 (Thermo Scientific, USA) and Essential FTIR (Operant LLC, USA). After baseline subtraction,
97 absorbance values were normalized using the area of the C-H stretching band at 2900 cm^{-1}
98 which was used as an internal standard according to the calibration method described by
99 Lojewska et al. (Łojewska et al., 2005). The area was calculated in the range $2700\text{-}3000\text{ cm}^{-1}$.
100 Attribution of the peaks in the IR spectra was based on literature references (Ogundare et al.,
101 2017; Schwanninger et al., 2004; Xu et al., 2013; Zhao et al., 2017).

102

103 **2.3 XRD:** X-ray diffractograms were recorded with a D2-PHASER spectrometer (Bruker, USA).
104 Spectra were recorded in the range $2\theta = 4\text{-}60$ degrees, with a resolution of 0.016 degrees.
105 The $\text{CuK}\alpha$ emission at 1.54 \AA was used as X-ray source. Cumulative spectra were obtained by
106 recording three diffractograms for each sample. During each measurement, the sample was
107 kept under rotation to prevent iso-orientation effects. The spectra were processed with
108 DIFFRAC (Bruker, USA). Baseline subtraction and peak fitting were performed using PeakFit
109 (Systat Software Inc., USA). Simulated diffraction patterns were used as reference to establish
110 the appropriate number of peaks (French, 2014).

111 Crystallinity indices were evaluated with the peak fitting method (Park et al., 2010), assuming
112 the presence of five Gaussian peaks for the crystalline phase and a sixth Gaussian peak for
113 the amorphous phase. Once the peak areas were calculated from the peak fitting, CI was
114 determined with equation (1), where A_{AM} is the area of the amorphous cellulose peak, and
115 A_{TOT} is the total area of all peaks. The r^2 values for the fitted peaks were always greater than
116 0.95. The dimensions of the crystallites in the un-milled samples were also estimated using
117 the Scherrer equation (2), where τ is the crystallite dimension, K a shape factor which was set

118 to 1.0 (Yue et al., 2015), λ the wavelength of incident light, θ the nominal diffraction angle for
119 a given diffraction peak, and b the half-height width of the peak.

120

121 (1) $CI = 1 - A_{AM}/A_{TOT}$

122 (2) $\tau = K\lambda/b\cos\theta$

123

124 **2.4 SEC-UV-MALS-DRI:** Before SEC analysis, all cellulose samples were converted into
125 cellulose tricarbaniolate (CTC) with a previously optimised procedure (Pawcenis et al., 2015).
126 Briefly, 5 mg of each sample were weighted in a glass vial and dried at 105 °C for 30 min. The
127 derivatisation was performed by adding 1 mL of pyridine and 0.1 mL of phenyl-isocyanate to
128 the vials, and by keeping the vials at 80 °C for 48 h. After this time, 0.1 mL of methanol were
129 added to stop the reaction. Finally, the solution was diluted with 10 mL of tetrahydrofuran
130 and filtered on 0.45 μm PTFE syringe filters (Equimed, Poland) and injected in the
131 chromatographic system.

132 The SEC apparatus consisted of a 1515 isocratic pump and a 717+ auto-sampler (Waters, USA).
133 Detection was performed using a 2487 UV detector (Waters, USA), Dawn Heleos multi-angle
134 light scattering detector (Wyatt Technology, USA), and Optilab T-rEX differential refractive
135 index detector (Wyatt Technology, USA). The separation was achieved using tetrahydrofuran
136 as mobile phase, and two Jordi Gel DVB columns (25 cm x 1 cm, 500 Å packing, Jordi Labs,
137 USA) equipped with a Styragel guard column (4.6 mm x 30 mm, 20 μm , Waters, USA). The
138 chromatographic columns were kept at 35 °C during all experiments. Mobile flow rate was 1
139 mL min⁻¹, and injection volume was 100 μL . Replicates were made for all samples to evaluate
140 uncertainty. Data elaboration was performed using Astra 6.1.7.17 (Wyatt Technology, USA).

141 SEC analyses provide the weight-average molar mass of the sample M_w , which can be related
142 to the intensity of scattered light using equation (3), where c is the polymer concentration,
143 K^* an instrumental constant, R_G the radius of gyration of the polymer particles, λ the
144 wavelength of incident light and A_2 the second virial coefficient of the polymer solution. R_θ is
145 the excess Rayleigh ratio, which is directly proportional to the intensity of scattered light at
146 the angle θ . This equation shows that for a given polymer concentration, cK^*/R_θ is linearly
147 dependent from $\sin^2\theta/2$. The intercept of this line equals the second bracket in the equation,
148 which can be used to calculate M_w if A_2 and c are known.

149

$$150 \quad (3) \quad \frac{cK^*}{R_\theta} = \left(1 + \frac{16\pi^2 R_G^2}{3\lambda^2} \sin^2 \theta/2\right) \left(\frac{1}{M_w} + 2A_2c\right)$$

151

152 Once M_w is obtained, the degree of polymerization can be calculated from M_w dividing by the
153 molar mass of a monomer. The yield of the carbanilation of cellulose is known to be 97.5%
154 (Daňhelka et al., 1976), and therefore the molar mass of the monomer is approximately 510.5
155 g mol⁻¹.

156

157 **2.5 EGA-MS:** Evolved gas analyses were carried out with an EGA/Py-3030D micro-furnace
158 pyrolyzer (Frontier Lab, Japan) coupled to a 6890 gas chromatograph (Agilent Technologies,
159 USA) equipped with a deactivated and uncoated stainless steel column (UADTM-2.5N, 2.5 m
160 x 0.15 mm, Frontier Lab, Japan). 50 μ g of sample were used for each EGA. In all experiments,
161 the temperature of the pyrolysis furnace was raised from 100 to 600 °C. Heating rates of 5,
162 10 and 20 °C min⁻¹ were used for all samples. Helium (1 mL min⁻¹) was used as carrier gas.

163 Injection was performed in split mode with a split ratio of 100:1. The GC oven and injector
164 were kept at 300 °C during all analyses. Detection was carried out with a 5973 Mass Selective
165 detector (Agilent Technologies, USA), operating in EI positive mode (70 eV, mass range m/z
166 50-300). The thermograms were processed using MSD ChemStation (Agilent Technologies,
167 USA). Experiments were repeated three times to assess the reproducibility of the
168 thermograms. Relative standard deviations were lower than 10%.

169 The thermograms were used to obtain apparent activation energy values using model-free
170 isoconversional methods. For this purpose, conversion profiles were calculated from the
171 thermograms using $\alpha(T) = I(T)/I_{tot}$, where α is the conversion, $I(T)$ the cumulative integral of
172 the thermogram up to the temperature T , and I_{tot} is the total integral of the thermogram. Two
173 different isoconversional data processing methods were used: the Kissinger-Akahira-Sunose
174 method (KAS) and the Flynn-Wall-Ozawa method (FWO). The equations for these methods
175 are (4) and (5), where β is the heating rate, T the temperature, E_a the apparent activation
176 energy, R the gas constant, and $q(\alpha)$ and $r(\alpha)$ are functions of the conversion. For a fixed value
177 of α , these equations give a linear trend of the left side with respect to $1/T$, and the slope can
178 be directly related to the apparent activation energy of the thermal degradation process.

179

180
$$(1) \ln \frac{\beta}{T^2} = -\frac{E_a}{RT} + q(\alpha)$$

181
$$(2) \ln \beta = -1.052 \frac{E_a}{RT} + r(\alpha)$$

182

183 **3. RESULTS AND DISCUSSION**

184 **3.1 ATR-FTIR:** ~~The ATR-FTIR spectra obtained for Sigmacell 20 cellulose at all milling times are~~
185 ~~reported in Figure 1a, and peak attributions are reported in Table 1.~~ ATR-FTIR spectra
186 obtained for Sigmacell 20 cellulose at all milling times and peak attributions are reported in
187 Figure S1a and Table S1 in Supplementary Materials, respectively. When milling time was
188 increased, a significant loss in resolution for some of the absorption bands was observed. This
189 alteration of the spectra was attributed to a reduction in the cellulose crystallinity. While the
190 orderly structure of crystalline cellulose generates sharp and resolved peaks, amorphous
191 cellulose generates broad peaks, most likely due to broader distributions of the transition
192 energies (Široký et al., 2010). In addition, ATR is sensitive to crystal size: upon the decrease in
193 crystal size more light scattering is observed which is manifested by the increase of the base
194 line absorption. The changes after ball-milling were particularly evident for the bands at 1051,
195 1103, 1317, 1335, 1362 and 1418 cm^{-1} . Similar changes were reported by Schwanninger and
196 co-workers in FTIR transmittance analyses of spruce wood after short-time ball milling
197 (Schwanninger et al., 2004).

198 Other changes in the spectral profile were observed for bands at 1027-1019 and 1159-1155
199 cm^{-1} (#3 and 6 in ~~Figure 1a and Table 1~~ Figure S1a and Table S1), which can be attributed to
200 C-O-C vibrations. While the bands intensities remained unaltered, the wavenumber of
201 maximum absorption shifted to smaller values as the milling time was increased. It is likely
202 that the cleavage of hydrogen bonds due to ball-milling allowed these bonds to change their
203 equilibrium distances and conformations, resulting in different vibrational energies. The shift
204 in absorption maxima was also reported by Schwanninger and co-workers for most of the
205 cellulose signals (Schwanninger et al., 2004).

206

207

208 Crystallinity of all samples was estimated from the ATR-FTIR spectra by calculating the ratio
209 of peaks at 1426 and 895 cm^{-1} (Nelson & O'Connor, 1964). The results are reported in Figure
210 1ea and ~~Table 2~~ Table 1. The CI derived from FTIR cannot be treated as absolute measure of
211 crystallinity, but it can be used for comparative assessment of the samples. All samples
212 showed a decreasing trend in the obtained CI values upon milling time. The most profound
213 changes were noted for the initial period of milling (30 min). This is in accordance with the
214 band broadening, which is more evident after 30 minutes of milling.

215

216 **3.2 XRD:** The XRD spectra obtained for Sigmacell 20 cellulose are reported in ~~Figure 1b~~ Figure
217 S1b in the Supplementary Materials. The diffraction pattern corresponds to cellulose I β phase,
218 with diffraction peaks at $2\theta = 14.5^\circ, 16.2^\circ, 20.5^\circ, 22.4^\circ$ and 33.9° , corresponding to the (101),
219 (10-1), (201), (002) and (040) diffraction planes, respectively. Cellulose I β ~~which~~ is the most
220 common cellulose allomorph in nature (French, 2014; French & Santiago Cintrón, 2013). As
221 expected, the intensity of the reflection signals decreased upon milling time. Similarly to the
222 ATR-FTIR results, the smoothing of the diffractograms was most evident in the first 30 min of
223 milling, and less evident for the other milling times. The intensity reduction of the peaks is
224 also manifested by the decrease in the signal-to-noise ratio going from 60 to 120 min milled
225 samples. ~~After 2 hours of milling, the diffractograms were dominated by the signal of the~~
226 ~~amorphous phase, which is centred at approximately $2\theta = 19^\circ$.~~

227 The diffractograms were processed by peak fitting, using five Gaussian functions
228 corresponding to each of the ~~four~~ five reflection functions of crystalline cellulose, and a sixth
229 Gaussian function for the amorphous phase (Park et al., 2010). The resulting CI values are

230 reported in ~~Table 2~~ Table 1, while a graph showing the CI trends for all samples is reported in
231 Figure 1d. The initial CIs were higher than 0.6 for Sigmacell 50 and Sigmacell 20, and higher
232 than 0.7 for microcrystalline and acid-washed celluloses. ~~After 120 minutes of milling, the CI~~
233 ~~values decreased by 20% on average.~~ Similarly to the FTIR results, the most significant drop
234 in CI was observed for all samples after 30 minutes of milling, while all subsequent milling
235 times had less profound effects. ~~After 120 minutes of milling, the CI values decreased by 80%~~
236 ~~on average.~~ Comparing cellulose samples from different sources, the greatest decrease in CI
237 was observed for microcrystalline cellulose (~~22%-93%~~), while the smallest - for Sigmacell 20
238 Sigmacell 50 and acid-washed cellulose (~~16%-73%~~). There is a high positive correlation ($r =$
239 0.94) between the percentage decrease in CI and the starting CI value for all samples. This
240 suggests that ball milling tends to reduce the crystallinity of cellulose up to an asymptotic
241 value, which means that the samples with higher starting CIs will be more affected. We have
242 found also a good correlation between the CI values obtained from ATR-FTIR and from XRD (r
243 = 0.97), which indicates that both methods can be used to monitor the loss of crystallinity.
244 The dimensions of the crystalline domains obtained by using the Scherrer equation for the
245 un-milled samples are reported in ~~Table 2~~ Table 1. The diffraction peak at $2\theta = 22.4^\circ$ was used
246 in all four cases for the calculation. Estimation of the crystallite dimensions for all milled
247 samples was difficult and unreliable due to the loss in peak intensity. Acid-washed cellulose
248 showed the greatest crystallite dimension, while Sigmacell 50 cellulose showed the smallest.
249 There is no apparent correlation between the crystallite dimensions and the CI decrease after
250 two hours of milling, suggesting that the efficiency of ball-milling does not depend on τ .

251

252 **3.3 SEC-UV- MALS-DRI:** Figure 2a shows the molar mass distribution for Sigmacell 20 at all
253 milling times. The DP values derived from the SEC results for all samples are reported in ~~Table~~
254 **2 Table 1**, and a graphic representation is given in Figure 2b. All un-milled cellulose samples
255 showed similar degrees of polymerization, although the plant source is not the same for every
256 sample. This could be due to similar extraction or purification processes. Acid-washed
257 cellulose showed the highest DP which amounts to 283, while microcrystalline cellulose
258 showed the lowest DP - 232. Both Sigmacell celluloses had initial DPs values of 251.

259 After milling, both a slight shift of the peak maxima and a peak broadening in the molecular
260 mass distribution curves were observed (Figure 2a). These results are consistent with the
261 decrease in degree of polymerisation. The extent of this decrease was different for all
262 samples: acid-washed cellulose was the most affected, and after two hours its DP dropped to
263 192; Sigmacell 20 was the least affected, reaching 218 after two hours of milling;
264 microcrystalline cellulose remained the least polymerized at all milling times, dropping from
265 114 to 102 (10%).

266 Coefficients of variations (CVs) on the DP values were generally lower than 3%, with few
267 exceptions. If the un-milled and 120 minutes milled samples are compared using Student's t-
268 test, the variations in DP prove to be statistically significant for all cellulose samples at a 95%
269 confidence. Although the effect of ball-milling on the DP is usually overlooked, these results
270 show that it should be considered during pre-treatment of cellulose samples. Finally, it is
271 interesting to note that the trends in DP and CI with milling time are different. While the
272 decrease in CI is more drastic in the first 30 minutes of milling, the decrease in DP is more
273 homogeneous along the whole investigated time range. This is also shown by the average

274 correlation coefficients between milling time and CI ($r = -0.88$) and between milling time and
275 DP ($r = -0.98$).

276

277 **3.4 EGA-MS:** The set of thermograms obtained from EGA-MS for un-milled Sigmacell 20
278 cellulose is reported ~~at the top of Figure 3 in Figure 3a~~. As the heating rate was increased, the
279 gas evolution peak moved to higher temperatures. The thermograms were processed with
280 model-free isoconversional methods, to obtain apparent activation energy values for
281 cellulose pyrolysis. ~~To use isoconversional methods, conversion profiles were calculated from~~
282 ~~the thermograms using $\alpha(T) = I(T)/I_{tot}$, where α is the conversion, $I(T)$ the cumulative integral~~
283 ~~of the thermogram up to the temperature T , and I_{tot} is the total integral of the thermogram.~~
284 ~~Two different isoconversional data processing methods were used: the Kissinger-Akahira-~~
285 ~~Sunose method (KAS) and the Flynn-Wall-Ozawa method (FWO). The equations for these~~
286 ~~methods are (4) and (5), where β is the heating rate, T the temperature, E_a the apparent~~
287 ~~activation energy, R the gas constant, and $q(\alpha)$ and $r(\alpha)$ are functions of the conversion. For a~~
288 ~~fixed value of α , these equations give a linear trend of the left side with respect to $1/T$, and~~
289 ~~the slope can be directly related to the apparent activation energy of the thermal degradation~~
290 ~~process.~~

291

292 ~~(3) $\ln \frac{\beta}{T^2} = -\frac{E_a}{RT} + q(\alpha)$~~

293 ~~(4) $\ln \beta = -1.052 \frac{E_a}{RT} + r(\alpha)$~~

294

295 For each sample, seven different conversion values were chosen to perform linear regression,
296 ranging from $\alpha = 0.2$ to $\alpha = 0.8$. The seven slopes were averaged to obtain an estimation of
297 E_a . The results are reported in ~~Table 3~~ Table 1, and ~~a graphical representation~~ graphical
298 ~~representations~~ of the trends ~~is are~~ shown in ~~the bottom row of Figure 3~~ Figure 3e and f.
299 Sigmacell 50 cellulose showed the highest E_a at all milling times, with a starting value of 136-
300 139 kJ/mol. On the other hand, microcrystalline cellulose showed the lowest starting E_a with
301 114-118 kJ/mol. After 120 minutes of milling, the E_a decreased by about 10% on average.
302 Sigmacell 50 cellulose showed the greatest drop (16-17%), while microcrystalline cellulose
303 showed the smallest (9-10%). The E_a values for the un-milled samples are similar to those
304 found by previous studies using isoconversional methods (Ali et al., 2017; Dahiya et al., 2008;
305 Kim et al., 2017; Sánchez-Jiménez et al., 2011). However, it should be pointed out that the
306 activation energy values obtained by isoconversional methods are only an approximation of
307 the intrinsic activation energy involved in the whole pyrolysis process. This is because the
308 intrinsic activation energy can be derived only if the pyrolysis mechanism is known or at least
309 the kinetic equation is derived. Therefore, the values of E_a obtained in this work should only
310 be used for comparison of samples treated under the same conditions.

311 CVs on the E_a values were lower by 5% in most of the cases, with few exceptions. The
312 differences between E_a values were evaluated with Student's and Welch's tests at a 95%
313 confidence. The differences between the un-milled and the 120 minutes milled samples
314 proved to be statistically relevant in all four cases. Moreover, all the E_a variations with milling
315 time were statistically relevant for both Sigmacell 50 and Sigmacell 20 cellulose. An
316 anomalous trend was observed for acid-washed cellulose, for which the E_a value after two
317 hours of milling was greater than the one after one hour. However, the differences among E_a

318 values of acid-washed cellulose at 30, 60 and 120 minutes of milling proved to be statistically
319 negligible (Student's t-test, 95% confidence).

320 The two isoconversional methods provided E_a values with good correlation ($r = 0.99$).
321 However, it is evident by looking at the values themselves that the FWO method provides
322 higher values than the KAS method. The 2003 paper by Starink (Starink, 2003) can be used to
323 estimate the error of the two isoconversional methods depending on the value of E_a/RT . In
324 this case, the error associated with FWO method is approximately one order of magnitude
325 bigger than the error associated with the KAS method (3% and 0.3%, respectively).

326 To obtain additional insight into the effect of milling on the cellulose strands, average mass
327 spectra were obtained from the EGA thermograms. The average mass spectra of un-milled
328 and 2-hours milled Sigmacell 20 are shown in Figure S2 in the Supplementary Materials.
329 Similar results were obtained for all other cellulose samples. As can be noted, the two spectra
330 showed the same profiles. The main signals at m/z 60 and 73 can be attributed to fragment
331 ions of levoglucosan, which is the main pyrolysis product of cellulose (Tamburini et al., 2015).
332 This result suggests that, although ball-milling reduced the apparent activation energy of
333 cellulose pyrolysis, it had no effect on the chemical structure of the cellulose strands, and
334 therefore the pyrolysis mechanism of the samples was not altered. This means that the
335 decrease in apparent activation energy is due to the decrease of CI and DP.

336 A recent publication by Sanchez-Jimenez and co-workers (Sánchez-Jiménez et al., 2011) used
337 model-based isoconversional methods and observed that random-chain scission is the most
338 suitable model to represent the pyrolysis mechanism of cellulose. This result is very
339 important, as it suggests that the degree of polymerisation has little impact on the energy
340 required for the pyrolysis of cellulose. This brings us to the conclusion that the main cause for

341 the decrease in apparent activation energy for the pyrolysis of cellulose is the decrease in
342 crystallinity. This is consistent with the fact that isoconversional methods provide an energy
343 value that considers all transformations taking place in the sample during pyrolysis, and
344 therefore the apparent activation energy values must include the energy required to cleave
345 the crystalline lattice.

346 Following this hypothesis, the apparent activation energies obtained with the KAS method,
347 which proved to be the most accurate, were plotted against the CIs obtained from the XRD
348 measurements, and linear regression was performed on the four cellulose samples separately
349 using equation (6).

350

$$351 \quad (5) \ E_a = E_1 + E_2 \cdot CI$$

352

353 Equation (6) shows that a hypothetical fully-crystalline cellulose sample (CI = 1) would have
354 an apparent activation energy equal to $E_1 + E_2$, while a fully-amorphous sample (CI = 0) would
355 have an apparent activation energy equal to E_1 . The value of E_2 is therefore an estimation of
356 the apparent activation energy for the cleavage of the crystalline lattice of cellulose.

357 The results are reported in Figure 4 and Table 2. Good R^2 values were obtained for all
358 cellulose samples. The slopes of the regression curves ranged from 18.1 kJ/mol for
359 Microcrystalline cellulose to 44.8 kJ/mol for Sigmacell 50. Averaging of the four slopes gave
360 30 kJ/mol. On the other hand, the four intercepts showed much smaller variation, and
361 averaging of the four intercepts gave 102 kJ/mol.

362 The lower error on the average intercept suggests that the apparent activation energy of the
363 pyrolysis process has been estimated with more accuracy than the apparent activation energy
364 for the amorphization process. A likely reason for this result is that there are some additional

365 contributions to the energy of the crystalline lattice, which are different among the various
366 samples and are not considered in the proposed model. A possible additional factor could be
367 a different spatial arrangement of the crystallites inside the amorphous phase, or a
368 heterogeneous distribution of the two phases in the cellulose powder.

369

370 **4. CONCLUSIONS**

371 Ball-milling of cellulose gave rise to an average decrease of 80% in crystallinity indices
372 derived from FTIR and XRD analyses. The crystallinity drop showed an asymptotic trend.

373 The DP decreased by 20% on average, with approximately linear trend. This is an important
374 observation since the effect of ball-milling on the DP is usually neglected.

375 Apparent activation energy values decreased by 10% on average, indicating that
376 depolymerization and amorphization significantly reduce thermal stability.

377 We believe that the results provided here will contribute to the discussion on the effects of
378 ball-milling on cellulose, and could be used to optimize cellulose conversion processes.

379

380 **ACKNOWLEDGEMENTS**

381 The authors would like to thank the European Regional Development Fund in the framework
382 of the Polish Innovation Economy Operational Program (contract no. POIG.02.01.00-12-
383 023/08) for the purchase of the Optilab t-rEX detector. In addition, the University of Pisa is
384 acknowledgment for having funded the project “Advanced analytical pyrolysis to study
385 polymers in renewable energy, environment, cultural heritage” (PRA_2018_26).

386 The authors would also like to thank Carlo Gini of the Department of Earth Sciences of the
387 University of Pisa for the XRD measurements.

- 389 [1] Agarwal, U.P., Reiner, R.S., Ralph, S.A. 2010. Cellulose I crystallinity determination using FT-Raman
390 spectroscopy: univariate and multivariate methods. *Cellulose*, **17**(4), 721-733.
- 391 [2] Ali, I., Bahaitham, H., Naebulharam, R. 2017. A comprehensive kinetics study of coconut shell waste
392 pyrolysis. *Bioresource Technology*, **235**, 1-11.
- 393 [3] Alvira, P., Tomás-Pejó, E., Ballesteros, M., Negro, M. 2010. Pretreatment technologies for an
394 efficient bioethanol production process based on enzymatic hydrolysis: a review. *Bioresource
395 Technology*, **101**(13), 4851-4861.
- 396 [4] Avolio, R., Bonadies, I., Capitani, D., Errico, M., Gentile, G., Avella, M. 2012. A multitechnique
397 approach to assess the effect of ball milling on cellulose. *Carbohydrate Polymers*, **87**(1), 265-
398 273.
- 399 [5] Bledzki, A.K., Gassan, J. 1999. Composites reinforced with cellulose based fibres. *Progress in
400 Polymer Science*, **24**(2), 221-274.
- 401 [6] da Silva, A.S.A., Inoue, H., Endo, T., Yano, S., Bon, E.P.S. 2010. Milling pretreatment of sugarcane
402 bagasse and straw for enzymatic hydrolysis and ethanol fermentation. *Bioresource
403 Technology*, **101**(19), 7402-7409.
- 404 [7] Dahiya, J.B., Kumar, K., Muller-Hagedorn, M., Bockhorn, H. 2008. Kinetics of isothermal and non-
405 isothermal degradation of cellulose: model-based and model-free methods. *Polymer
406 International*, **57**(5), 722-729.
- 407 [8] Daňhelka, J., Kössler, I., Boháčková, V. 1976. Determination of molecular weight distribution of
408 cellulose by conversion into tricarbonyl and fractionation. *Journal of Polymer Science Part
409 A: Polymer Chemistry*, **14**(2), 287-298.
- 410 [9] De Silva, R., Byrne, N. 2017. Utilization of cotton waste for regenerated cellulose fibres: influence
411 of degree of polymerization on mechanical properties. *Carbohydrate Polymers*.
- 412 [10] French, A.D. 2014. Idealized powder diffraction patterns for cellulose polymorphs. *Cellulose*,
413 **21**(2), 885-896.
- 414 [11] French, A.D., Santiago Cintrón, M. 2013. Cellulose polymorphism, crystallite size, and the Segal
415 Crystallinity Index. *Cellulose*, **20**(1), 583-588.
- 416 [12] Gu, B.-J., Wang, J., Wolcott, M.P., Ganjyal, G.M. 2018. Increased sugar yield from pre-milled
417 Douglas-fir forest residuals with lower energy consumption by using planetary ball milling.
418 *Bioresource Technology*, **251**, 93-98.
- 419 [13] Hallac, B.B., Ragauskas, A.J. 2011. Analyzing cellulose degree of polymerization and its relevancy
420 to cellulosic ethanol. *Biofuels, Bioproducts and Biorefining*, **5**(2), 215-225.
- 421 [14] Kaur, R., Gera, P., Jha, M.K., Bhaskar, T. 2018. Pyrolysis kinetics and thermodynamic parameters
422 of castor (*Ricinus communis*) residue using thermogravimetric analysis. *Bioresource
423 Technology*, **250**, 422-428.
- 424 [15] Kian, L.K., Jawaid, M., Ariffin, H., Karim, Z. 2018. Isolation and characterization of nanocrystalline
425 cellulose from roselle-derived microcrystalline cellulose. *International Journal of Biological
426 Macromolecules*.
- 427 [16] Kim, Y.-M., Han, T.U., Hwang, B., Lee, Y., Watanabe, A., Teramae, N., Kim, S.-S., Park, Y.-K., Kim, S.
428 2017. New approach for the kinetic analysis of cellulose using EGA-MS. *Polymer Testing*, **60**,
429 12-17.
- 430 [17] Lavoine, N., Desloges, I., Dufresne, A., Bras, J. 2012. Microfibrillated cellulose—Its barrier
431 properties and applications in cellulosic materials: A review. *Carbohydrate Polymers*, **90**(2),
432 735-764.
- 433 [18] Lee, H., Mani, S. 2017. Mechanical pretreatment of cellulose pulp to produce cellulose nanofibrils
434 using a dry grinding method. *Industrial Crops and Products*, **104**(Supplement C), 179-187.
- 435 [19] Łojewska, J., Miśkowiec, P., Łojewski, T., Proniewicz, L. 2005. Cellulose oxidative and hydrolytic
436 degradation: In situ FTIR approach. *Polymer Degradation and Stability*, **88**(3), 512-520.

- 437 [20] Łojewski, T., Zięba, K., Łojewska, J. 2010. Size exclusion chromatography and viscometry in paper
438 degradation studies. New Mark-Houwink coefficients for cellulose in cupri-ethylenediamine.
439 *Journal of Chromatography A*, **1217**(42), 6462-6468.
- 440 [21] Marx-Figini, M., Maximino, M. 1993. Effect of impregnation on the degree of polymerization of
441 cellulose during bleaching of sodapulped sugar cane bagasse. *Macromolecular Materials and*
442 *Engineering*, **208**(1), 167-172.
- 443 [22] Mukarakate, C., Mittal, A., Ciesielski, P.N., Budhi, S., Thompson, L., Lisa, K., Nimlos, M.R., Donohoe,
444 B.S. 2016. Influence of crystal allomorph and crystallinity on the products and behavior of
445 cellulose during fast pyrolysis. *ACS Sustainable Chemistry & Engineering*, **4**(9), 4662-4674.
- 446 [23] Nelson, M.L., O'Connor, R.T. 1964. Relation of certain infrared bands to cellulose crystallinity and
447 crystal latticed type. Part I. Spectra of lattice types I, II, III and of amorphous cellulose. *Journal*
448 *of Applied Polymer Science*, **8**(3), 1311-1324.
- 449 [24] Ni, J., Teng, N., Chen, H., Wang, J., Zhu, J., Na, H. 2015. Hydrolysis behavior of regenerated
450 celluloses with different degree of polymerization under microwave radiation. *Bioresource*
451 *Technology*, **191**, 229-233.
- 452 [25] Ogundare, S.A., Moodley, V., van Zyl, W.E. 2017. Nanocrystalline cellulose isolated from discarded
453 cigarette filters. *Carbohydrate Polymers*, **175**, 273-281.
- 454 [26] Park, S., Baker, J., Himmel, M., Parilla, P., Johnson, D. 2010. Cellulose crystallinity index:
455 measurement techniques and their impact on interpreting cellulase performance.
456 *Biotechnology for Biofuels*, **3**(1), 1-10.
- 457 [27] Pawcenis, D., Thomas, J.L., Łojewski, T., Milczarek, J.M., Łojewska, J. 2015. Towards determination
458 of absolute molar mass of cellulose polymer by size exclusion chromatography with multiple
459 angle laser light scattering detection. *Journal of Chromatography A*, **1409**, 53-59.
- 460 [28] Podzimek, S. 2011. *Light scattering, size exclusion chromatography and asymmetric flow field flow*
461 *fractionation: powerful tools for the characterization of polymers, proteins and nanoparticles.*
462 John Wiley & Sons.
- 463 [29] Podzimek, S. 2014. Truths and myths about the determination of molar mass distribution of
464 synthetic and natural polymers by size exclusion chromatography. *Journal of Applied Polymer*
465 *Science*, **131**(7).
- 466 [30] Qiu, X., Hu, S. 2013. "Smart" materials based on cellulose: a review of the preparations,
467 properties, and applications. *Materials*, **6**(3), 738-781.
- 468 [31] Rebière, J., Heuls, M., Castignolles, P., Gaborieau, M., Rouilly, A., Violleau, F., Durrieu, V. 2016.
469 Structural modifications of cellulose samples after dissolution into various solvent systems.
470 *Analytical and Bioanalytical Chemistry*, **408**(29), 8403-8414.
- 471 [32] Sánchez-Jiménez, P.E., Pérez-Maqueda, L.A., Perejón, A., Pascual-Cosp, J., Benítez-Guerrero, M.,
472 Criado, J.M. 2011. An improved model for the kinetic description of the thermal degradation
473 of cellulose. *Cellulose*, **18**(6), 1487-1498.
- 474 [33] Schenzel, K., Fischer, S., Brendler, E. 2005. New method for determining the degree of cellulose I
475 crystallinity by means of FT Raman spectroscopy. *Cellulose*, **12**(3), 223-231.
- 476 [34] Schwanninger, M., Rodrigues, J., Pereira, H., Hinterstoisser, B. 2004. Effects of short-time
477 vibratory ball milling on the shape of FT-IR spectra of wood and cellulose. *Vibrational*
478 *Spectroscopy*, **36**(1), 23-40.
- 479 [35] Siqueira, G., Bras, J., Dufresne, A. 2010. Cellulosic bionanocomposites: a review of preparation,
480 properties and applications. *Polymers*, **2**(4), 728-765.
- 481 [36] Široký, J., Blackburn, R.S., Bechtold, T., Taylor, J., White, P. 2010. Attenuated total reflectance
482 Fourier-transform Infrared spectroscopy analysis of crystallinity changes in lyocell following
483 continuous treatment with sodium hydroxide. *Cellulose*, **17**(1), 103-115.
- 484 [37] Starink, M. 2003. The determination of activation energy from linear heating rate experiments: a
485 comparison of the accuracy of isoconversion methods. *Thermochimica Acta*, **404**(1), 163-176.

- 486 [38] Tamburini, D., Łucejko, J.J., Ribechini, E., Colombini, M.P. 2015. Snapshots of lignin oxidation and
487 depolymerization in archaeological wood: an EGA-MS study. *Journal of Mass Spectrometry*,
488 **50**(10), 1103-1113.
- 489 [39] Wang, S., Dai, G., Yang, H., Luo, Z. 2017. Lignocellulosic biomass pyrolysis mechanism: A state-of-
490 the-art review. *Progress in Energy and Combustion Science*, **62**, 33-86.
- 491 [40] Xu, F., Yu, J., Tesso, T., Dowell, F., Wang, D. 2013. Qualitative and quantitative analysis of
492 lignocellulosic biomass using infrared techniques: a mini-review. *Applied Energy*, **104**, 801-
493 809.
- 494 [41] Yang, C.-H., Chen, C.-A., Chen, C.-F. 2018. Surface-Modified Cellulose Paper and Its Application in
495 Infectious Disease Diagnosis. *Sensors and Actuators B: Chemical*.
- 496 [42] Yue, Y., Han, J., Han, G., Zhang, Q., French, A.D., Wu, Q. 2015. Characterization of cellulose I/II
497 hybrid fibers isolated from energycane bagasse during the delignification process:
498 Morphology, crystallinity and percentage estimation. *Carbohydrate Polymers*, **133**, 438-447.
- 499 [43] Zhao, D., Yang, F., Dai, Y., Tao, F., Shen, Y., Duan, W., Zhou, X., Ma, H., Tang, L., Li, J. 2017. Exploring
500 crystalline structural variations of cellulose during pulp beating of tobacco stems.
501 *Carbohydrate Polymers*, **174**(Supplement C), 146-153.

502

503

504

505 **Figure 1:** ~~(a) ATR-FTIR spectra of Sigmacell 20 cellulose at different milling times. The signals~~
506 ~~are normalised by the correction factor. Peaks are numbered according to Table 1; (b) XRD~~
507 ~~spectra of Sigmacell type 20 at different milling times. The peaks are labelled using Miller~~
508 ~~indices; (c) CI trends obtained by (a) ATR-FTIR and (d)-(b) XRD spectra for all cellulose samples~~
509 ~~at all milling times. S20 = Sigmacell 20; S50 = Sigmacell 50; SMC = Microcrystalline cellulose;~~
510 ~~SAW = acid-washed cellulose.~~

511
512 **Figure 2:** (a) molar mass distribution plots for Sigmacell 20 at all milling times; (b) degrees of
513 polymerization for all samples at all milling times. S20 = Sigmacell 20; S50 = Sigmacell 50; SMC
514 = Microcrystalline cellulose; SAW = acid-washed cellulose.

515
516 **Figure 3:** (a) normalized thermograms obtained by EGA-MS for un-milled Sigmacell 20
517 cellulose at different heating rates; (b) conversion profiles obtained by integration of the
518 thermograms in (a); (c) linear regression curves obtained using the KAS and (d) FWO methods
519 for un-milled Sigmacell 20 cellulose at conversion values from 0.2 to 0.8; (e) trends in E_a values
520 obtained using the KAS and (f) FWO methods. S20 = Sigmacell 20; S50 = Sigmacell 50; SMC =
521 Microcrystalline cellulose; SAW = acid-washed cellulose.

522
523 **Figure 4:** Plots of E_a values obtained with the KAS method and CI values from XRD for all
524 cellulose samples.

525

526 **Table 1:** Crystallinity indices obtained by FTIR and XRD, crystallite dimensions, degrees of
 527 polymerization obtained by SEC and apparent activation energies obtained by EGA-MS using
 528 KAS and FWO isoconversional methods for all samples.

Sample	Milling time (min)	CI by FTIR	CI by XRD	τ (nm)	DP by SEC	E_a by KAS (kJ/mol)	E_a by FWO (kJ/mol)
Sigmacell 20	0	1.05	0.63	6.43	251 ± 5	122 ± 2	126 ± 2
	30	0.67	0.37		249 ± 6	113 ± 2	117 ± 2
	60	0.55	0.26		232 ± 3	110 ± 2	114 ± 1
	120	0.52	0.15		218 ± 1	106 ± 2	111 ± 2
Sigmacell 50	0	0.90	0.67	5.29	250 ± 10	136 ± 8	139 ± 7
	30	0.59	0.33		235 ± 1	126 ± 3	129 ± 3
	60	0.61	0.28		233 ± 3	119 ± 2	123 ± 2
	120	0.60	0.18		206 ± 6	113 ± 1	117 ± 1
Microcrystalline	0	0.87	0.73	6.00	232 ± 6	114 ± 9	118 ± 8
	30	0.56	0.37		227 ± 1	110 ± 2	114 ± 2
	60	0.46	0.17		213 ± 2	104 ± 4	108 ± 4
	120	0.46	0.05		183 ± 1	102 ± 6	106 ± 5
Acid-washed	0	0.87	0.75	8.18	283 ± 8	122 ± 8	125 ± 7
	30	0.72	0.33		266 ± 3	108 ± 4	113 ± 4
	60	0.66	0.25		250 ± 20	105 ± 3	110 ± 3
	120	0.63	0.20		192 ± 7	108 ± 1	112 ± 1

529

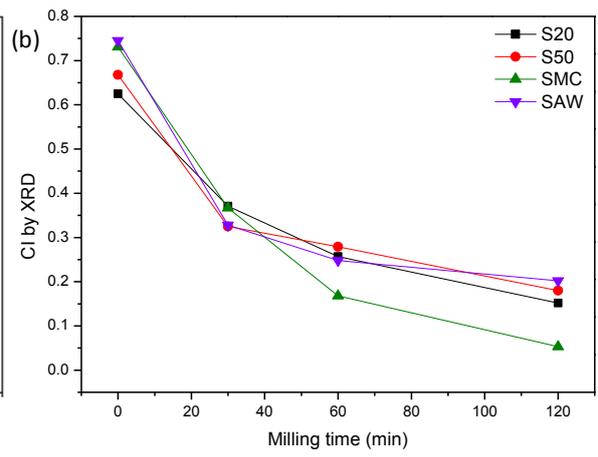
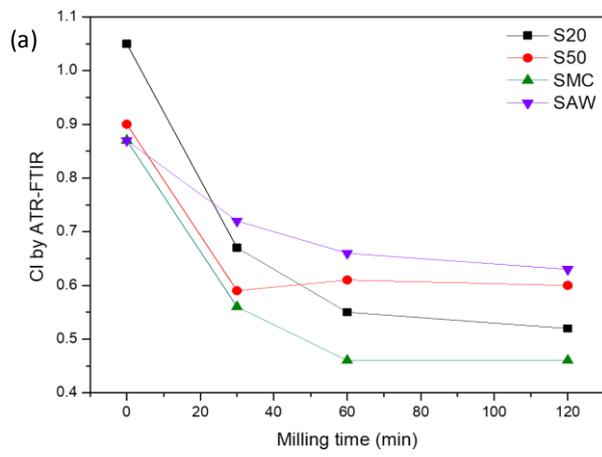
530

531 **Table 2:** Intercept, slope and R² values for the plots of apparent activation energy against CI
532 of all cellulose samples. Average intercept and slope are also reported.

Sample	INT ($\cdot 10^5$ J/mol)	SLO ($\cdot 10^4$ J/mol)	R ²
Sigmacell 20	1.01	3.30	0.98
Sigmacell 50	1.07	4.48	0.88
Microcrystalline	1.01	1.81	0.92
Acid-washed	1.00	2.86	0.91
Average	1.02 \pm 0.02	3.00 \pm 1.00	

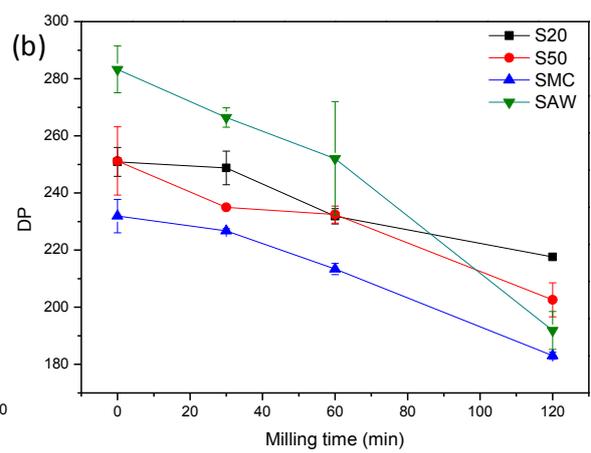
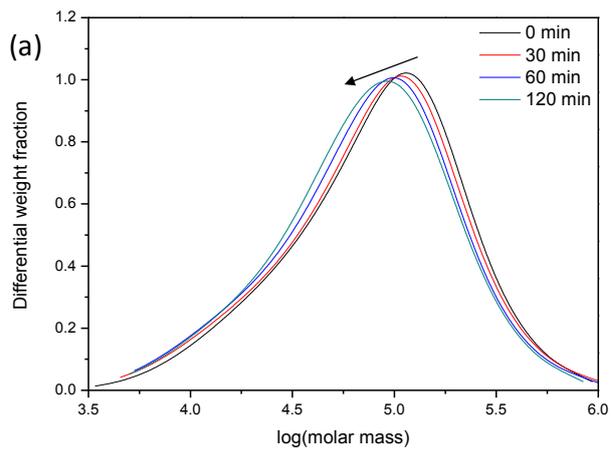
533

534



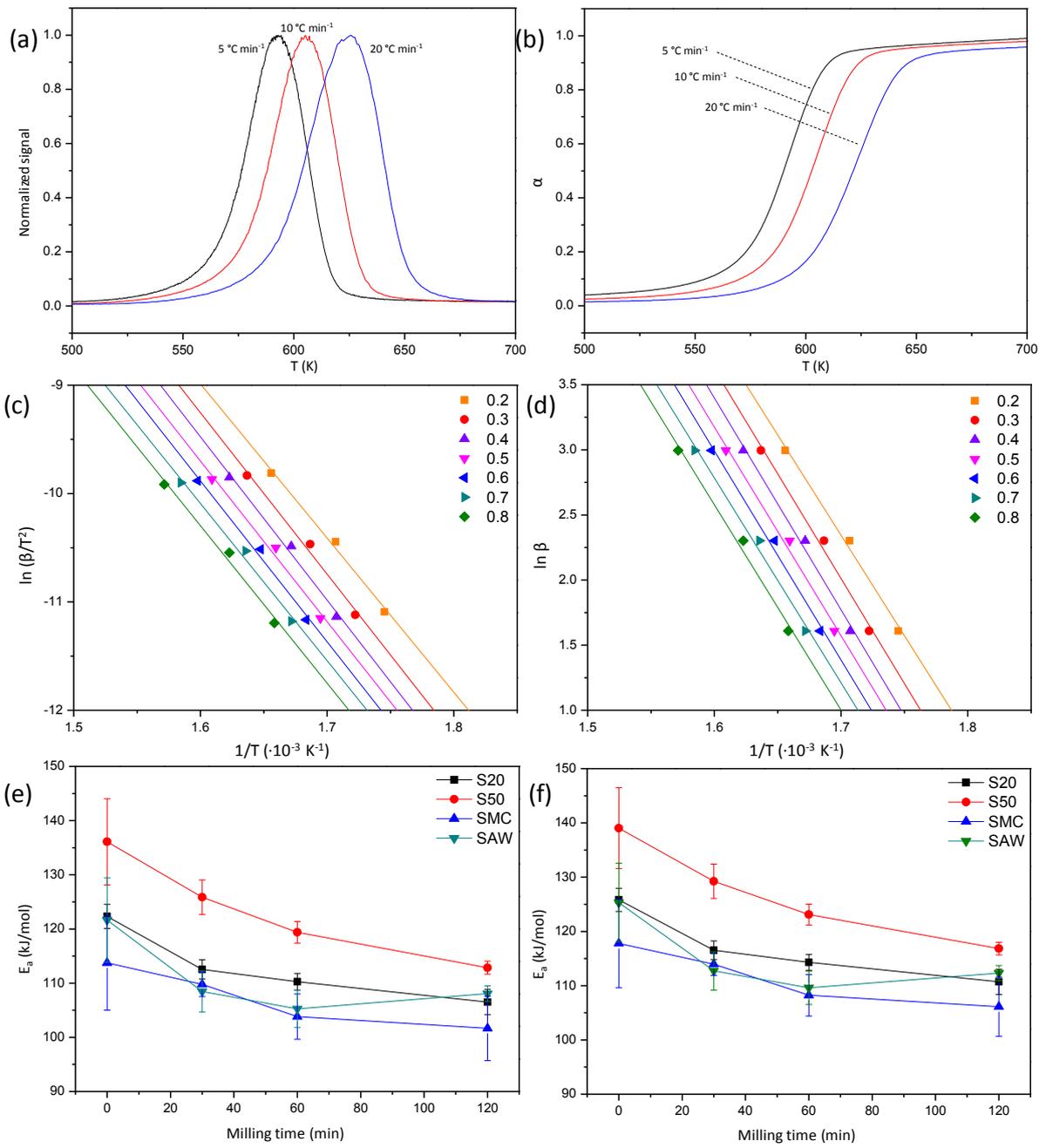
535

536



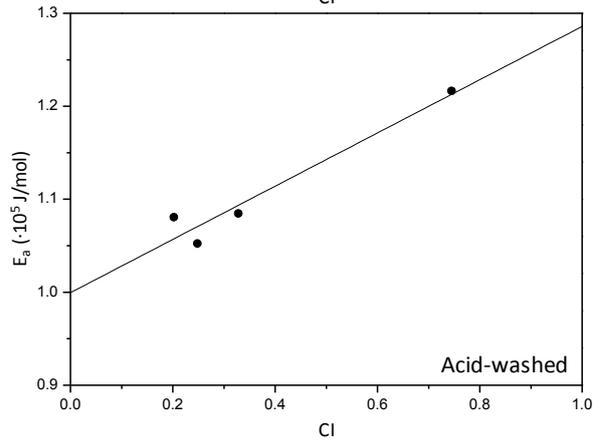
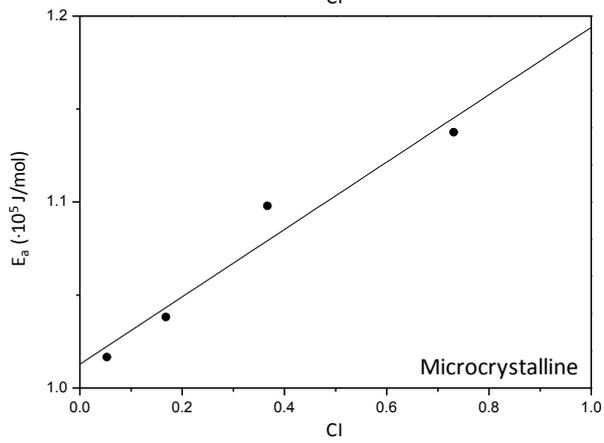
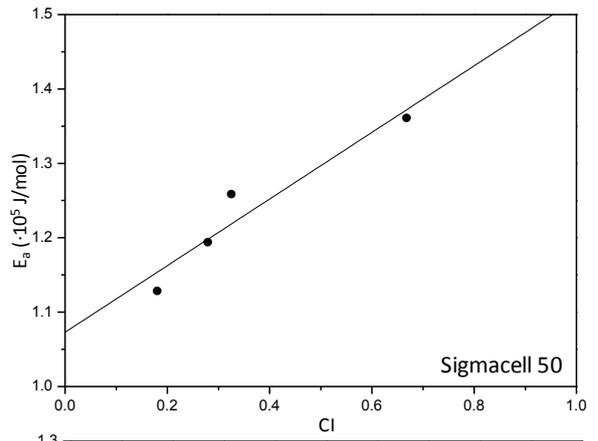
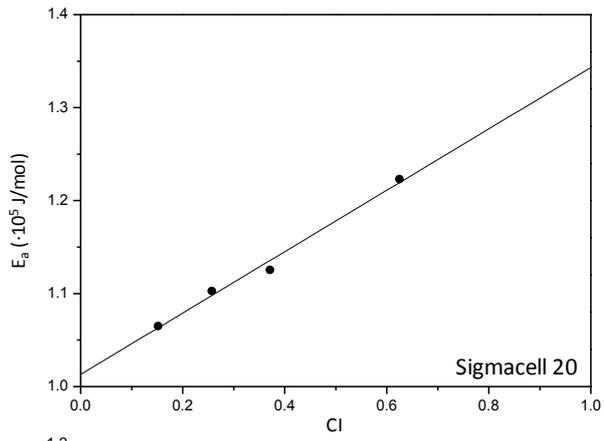
537

538



539

540



541

542

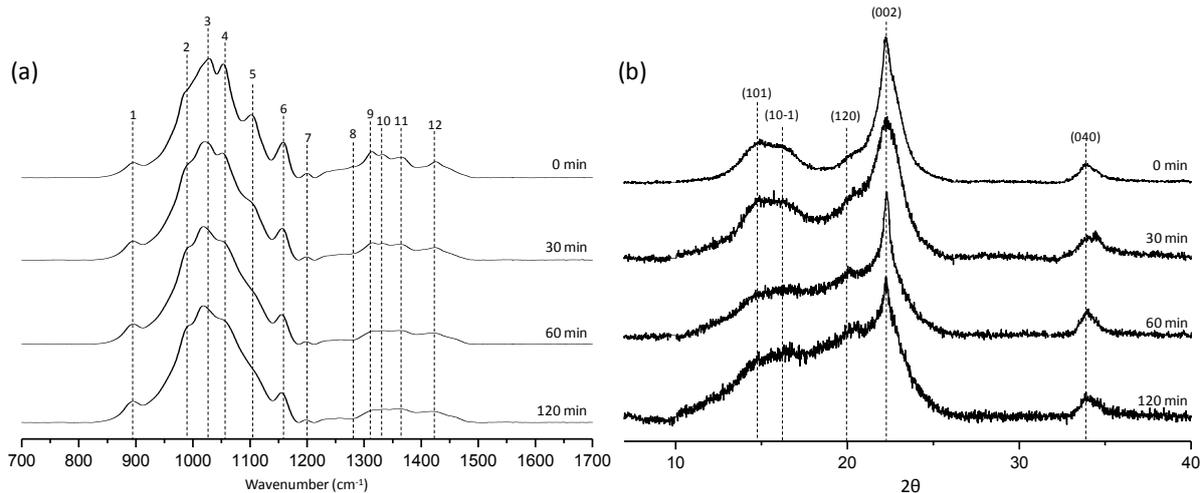
543

544 **SUPPLEMENTARY MATERIAL**545 **Table S1:** Peak attributions for the ATR-FTIR spectra.

Peak #	cm ⁻¹	Attribution	Reference
1	894	C-O-C, C-C-O, C-C-H stretching	(Schwanninger et al., 2004; Xu et al., 2013; Zhao et al., 2017)
2	985	C-O valence vibration	(Schwanninger et al., 2004; Xu et al., 2013)
3	1027-1019	C-O-C asymmetric stretching	(Ogundare et al., 2017)
4	1051	C-O valence vibration	(Zhao et al., 2017)
5	1103	Ring asymmetric valence vibration	(Zhao et al., 2017)
6	1159-1155	C-O-C asymmetric valence vibration	(Schwanninger et al., 2004; Xu et al., 2013; Zhao et al., 2017)
7	1199	O-H bending	(Schwanninger et al., 2004; Xu et al., 2013)
8	1281	C-H bending	(Schwanninger et al., 2004; Xu et al., 2013)
9	1317	CH ₂ wagging	(Schwanninger et al., 2004; Xu et al., 2013)
10	1335	C-H vibration, O-H bending	(Schwanninger et al., 2004; Xu et al., 2013; Zhao et al., 2017)
11	1362	C-H asymmetric bending	(Ogundare et al., 2017)
12	1418	H-C-H, O-C-H scissoring	(Schwanninger et al., 2004; Zhao et al., 2017)

546

547



548

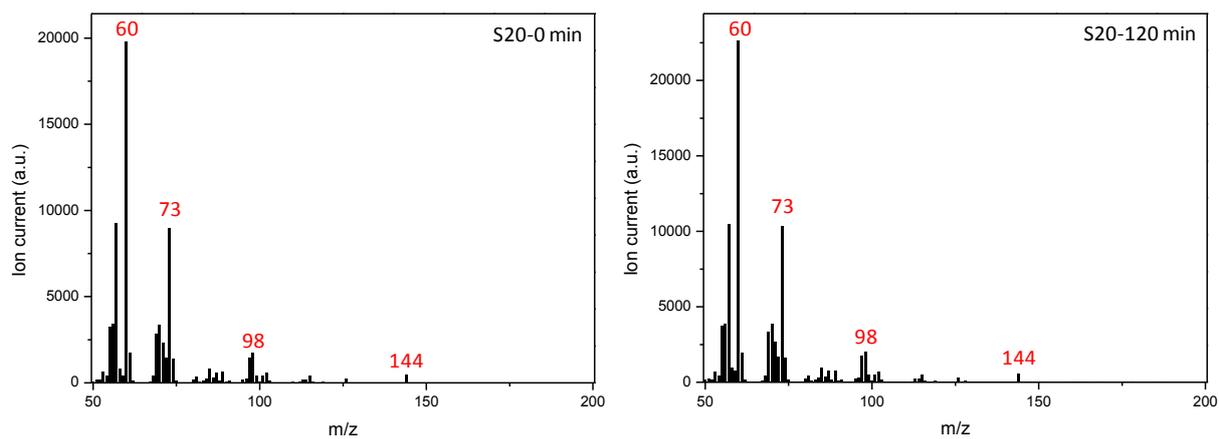
549 **Figure S1:** (a) ATR-FTIR spectra of Sigmacell 20 cellulose at different milling times. The

550 signals are normalised by the correction factor. Peaks are numbered according to Table 1;

551 (b) XRD spectra of Sigmacell type 20 at different milling times. The peaks are labelled using

552 Miller indices.

553



554

555 **Figure S2:** Average mass spectra from EGA-MS profiles of un-milled (left) and 2 hours milled
 556 (right) Sigmacell 20 samples.

557